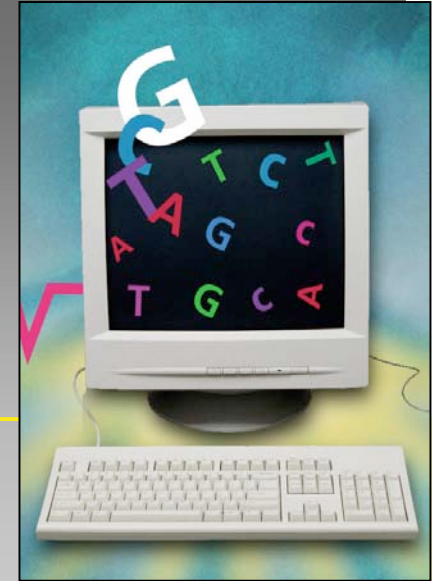


Humanbiologie 4. Semester
1-Tages Praktikum "Humangenom"



Funktionelle Einzelzell-Genomik
am Beispiel der dopaminergen Mittelhirn-Neuronen

Institut für Normale und Pathologische Physiologie
AG Molekulare Neurobiologie und Computerraum

B.Liss, A.Alexandru, J.Gründemann

I. Wie kann man methodisch Funktion und Genexpression einzelner Nervenzellen untersuchen ?

(Laser-Mikrodissection, PALM-System)

II. Wie nutzt man dazu die Informationen des Humangenom-Projektes und verwandter Internet-Datenbanken / Programme?

(Primer-Design für RT-PCR)

III. Warum ist Einzelzell-Analyse von Nervenzellen erstrebenswert?

(Theorie soviel wie gewünscht / zeitlich machbar)

Gen-Expressionsanalyse einzelner Zellen mittels RT-PCR

1. Isolation einzelner Zellen / mRNA (z.T. Funktionsanalyse)

(Laser Mikrodissektion, Patch-clamp Technik)

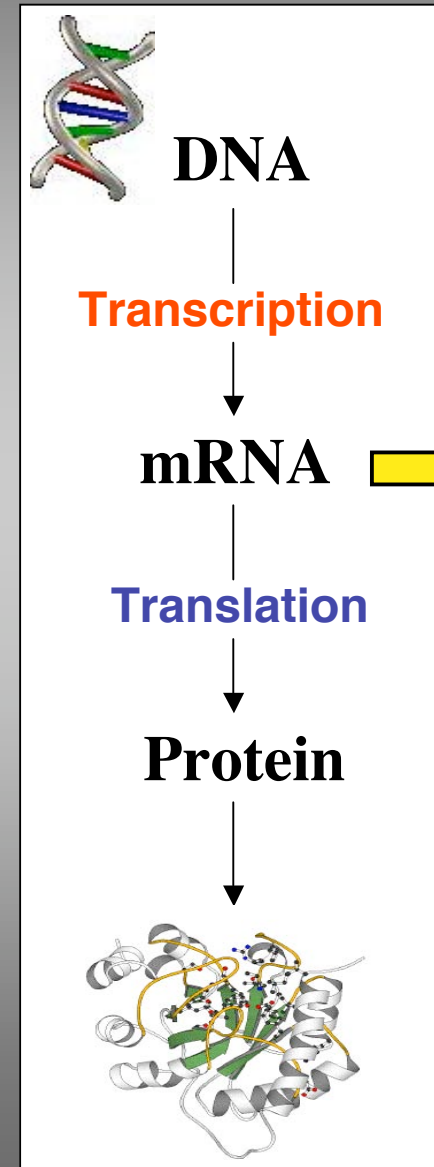
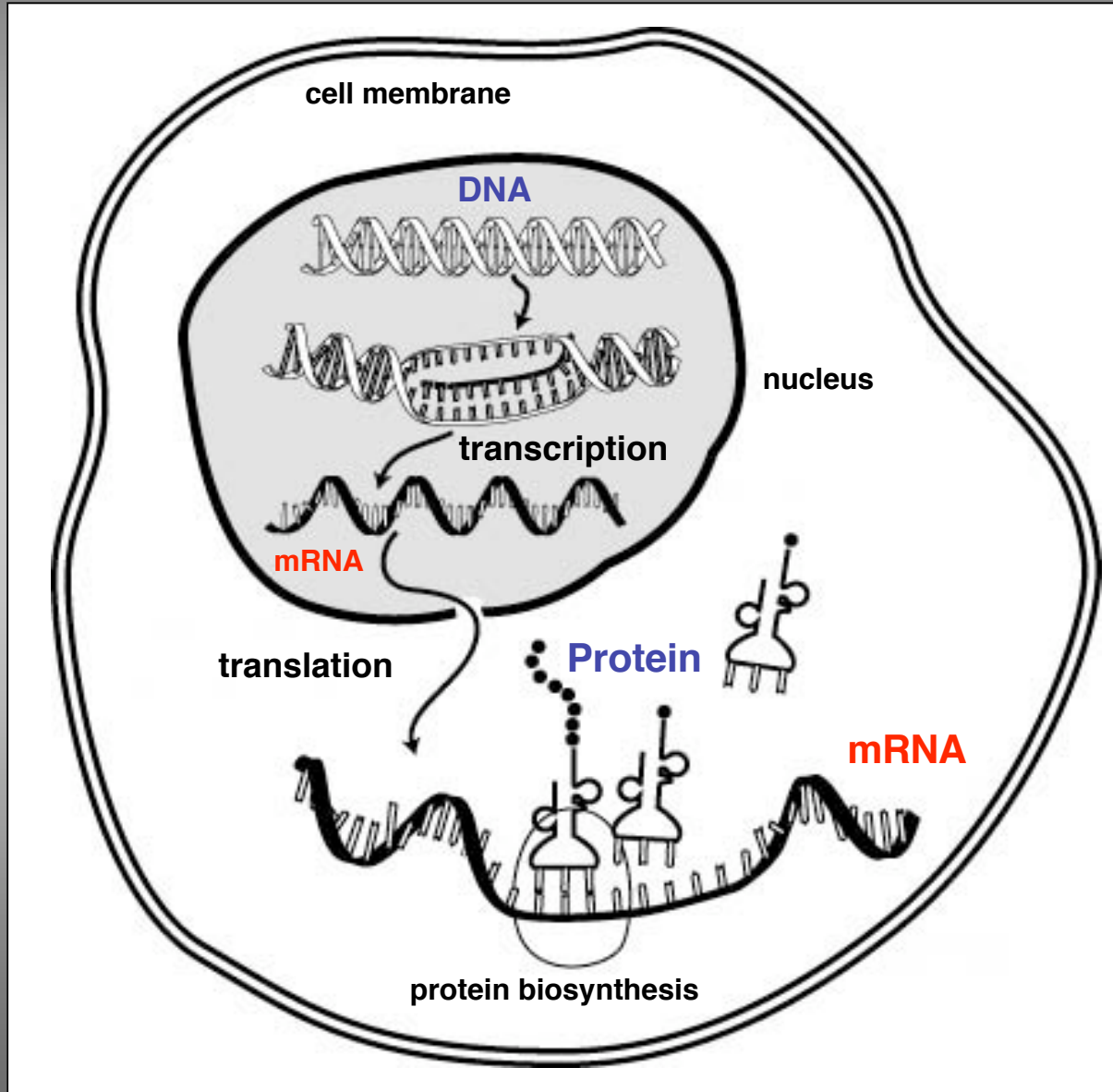
2. cDNA Synthese (Reverse Transkription der mRNA)

(Reverse Transkription -RT)

3. PCR Amplifikation der zu untersuchenden Gene/cDNAs

(Primer-Design, qualitative PCR, quantitative real-time PCR)

complementary DNA (cDNA): stabilere Negativkopie der mRNA

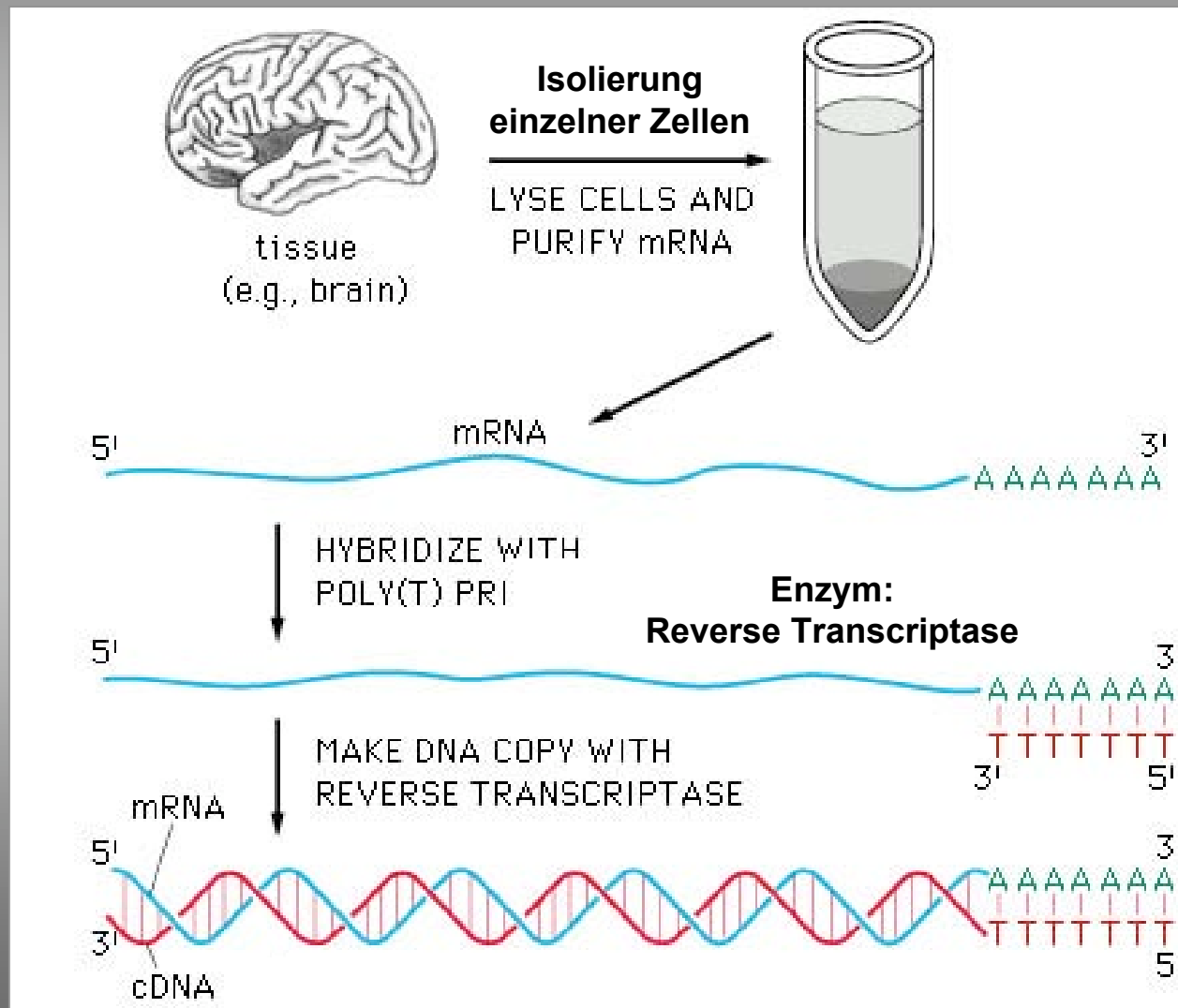


RNA ist
extrem fragil:
RNAsen!!

cDNA

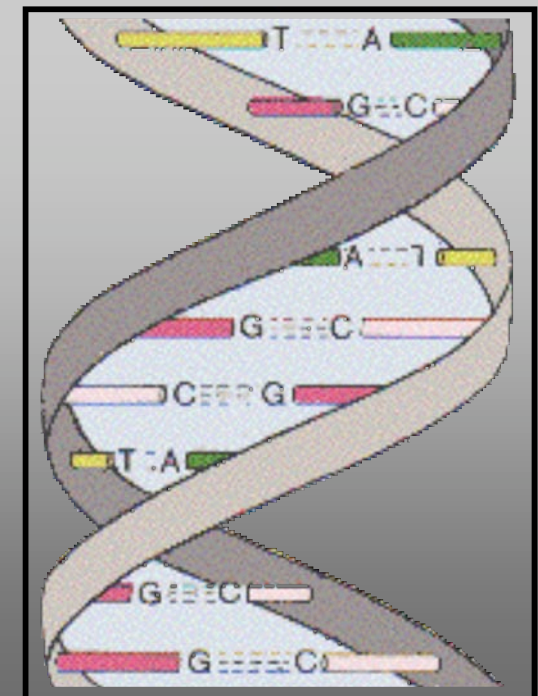
Reverse Transkription (RT) zur cDNA Synthese:

Umschreiben der mRNA in eine stabilere cDNA Form



Primer = Oligonukleotide:

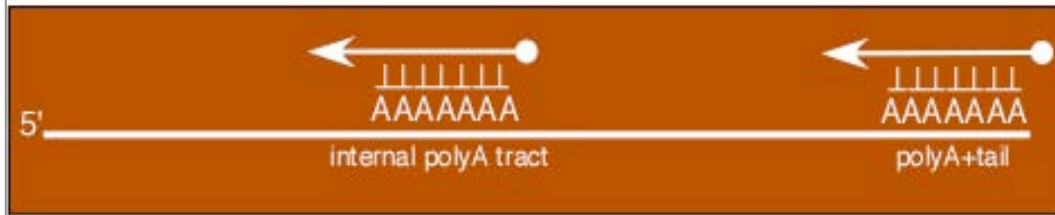
Kurze Einzelstrang-DNA (15-50 Basen) die durch Basenpaarung kurze dsDNA Bereiche bilden, die die Enzyme (zB *Reverse Transcriptase*) benötigen



Primerauswahl für cDNA Synthese / RT:

3 verschiedene Primer-Strategien

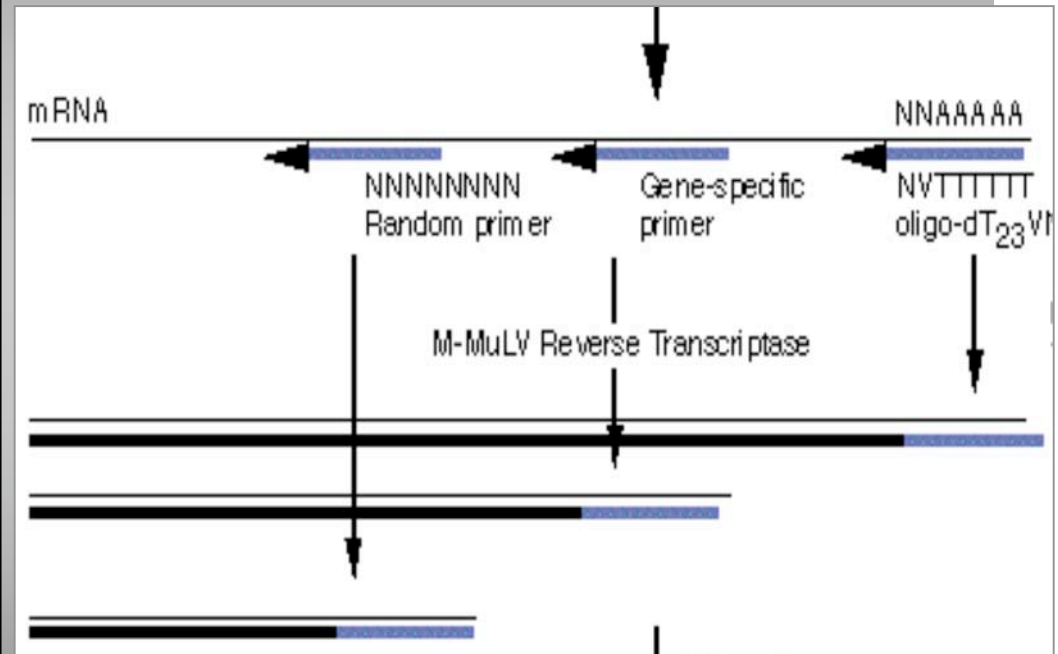
Oligo-dT priming



Random hexamer priming



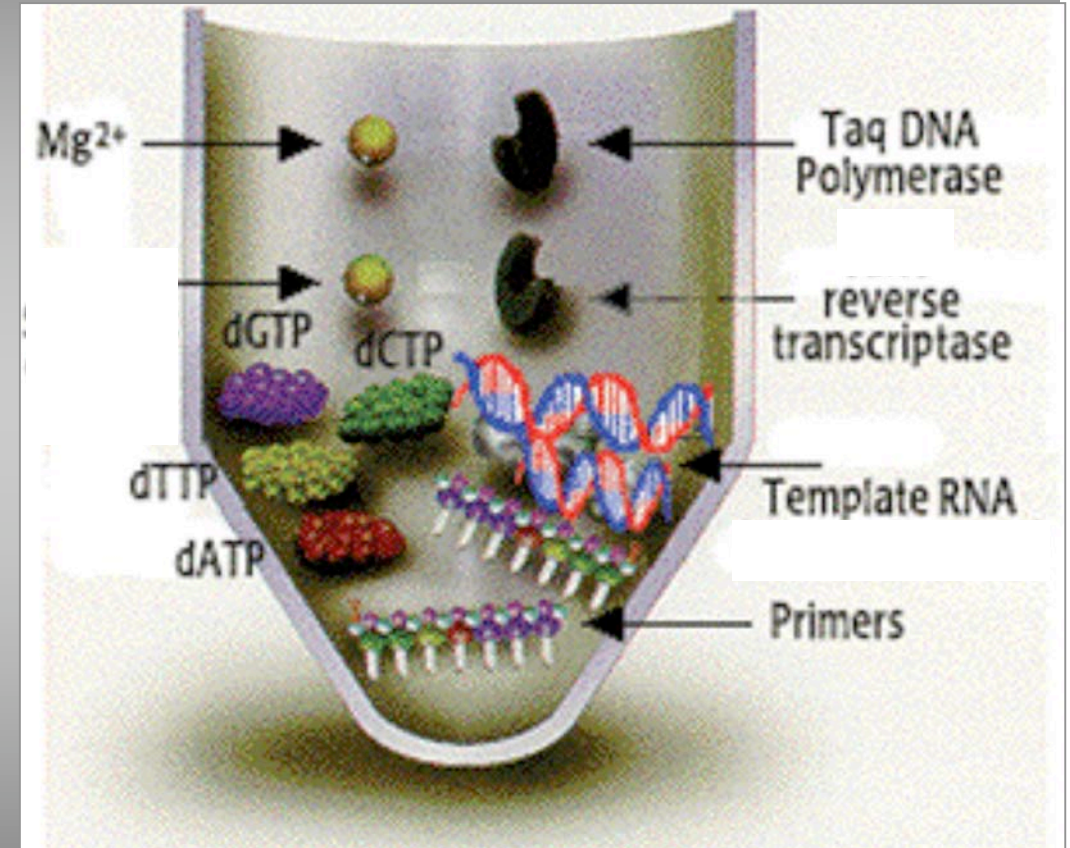
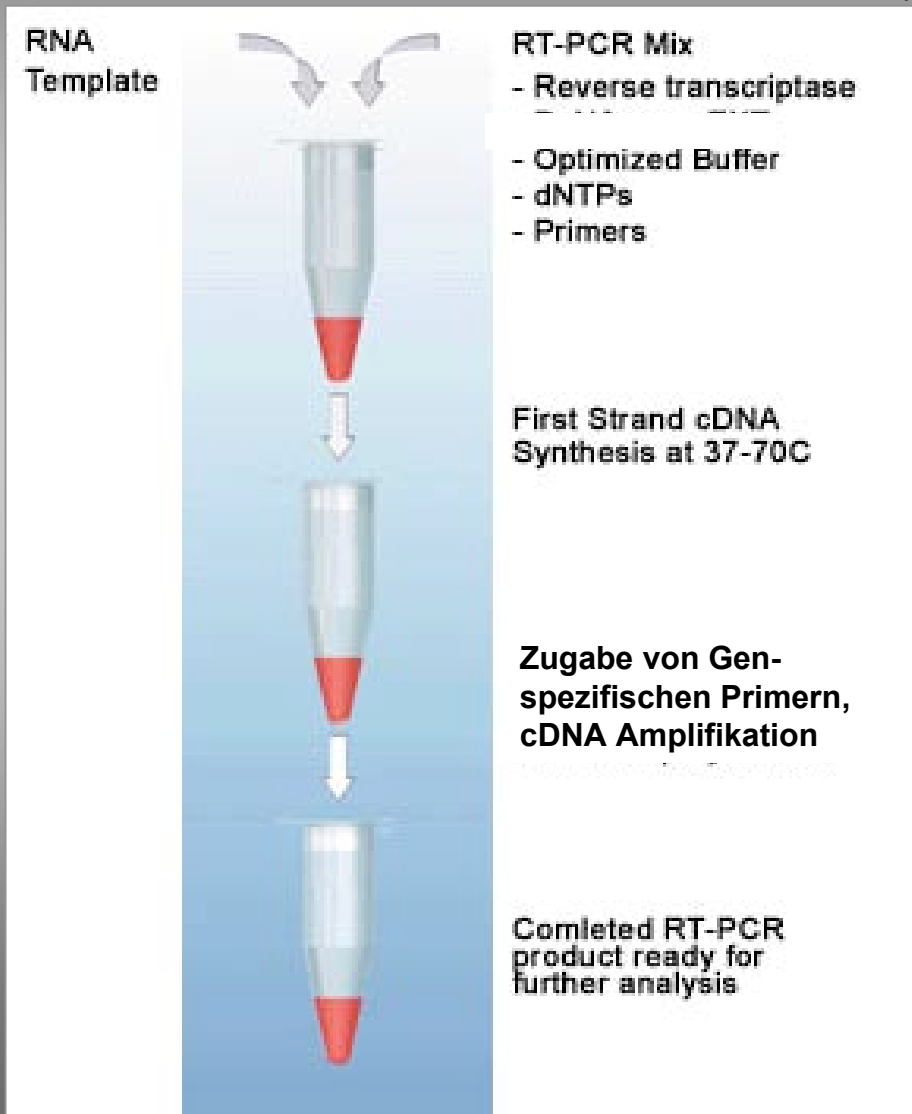
Gene-specific priming



3 verschiedene cDNA Populationen
von derselben RNA-Probe möglich:
Gen-Expressions Artefakte !

RT-PCR: reverse transcription - polymerase chain reaction:

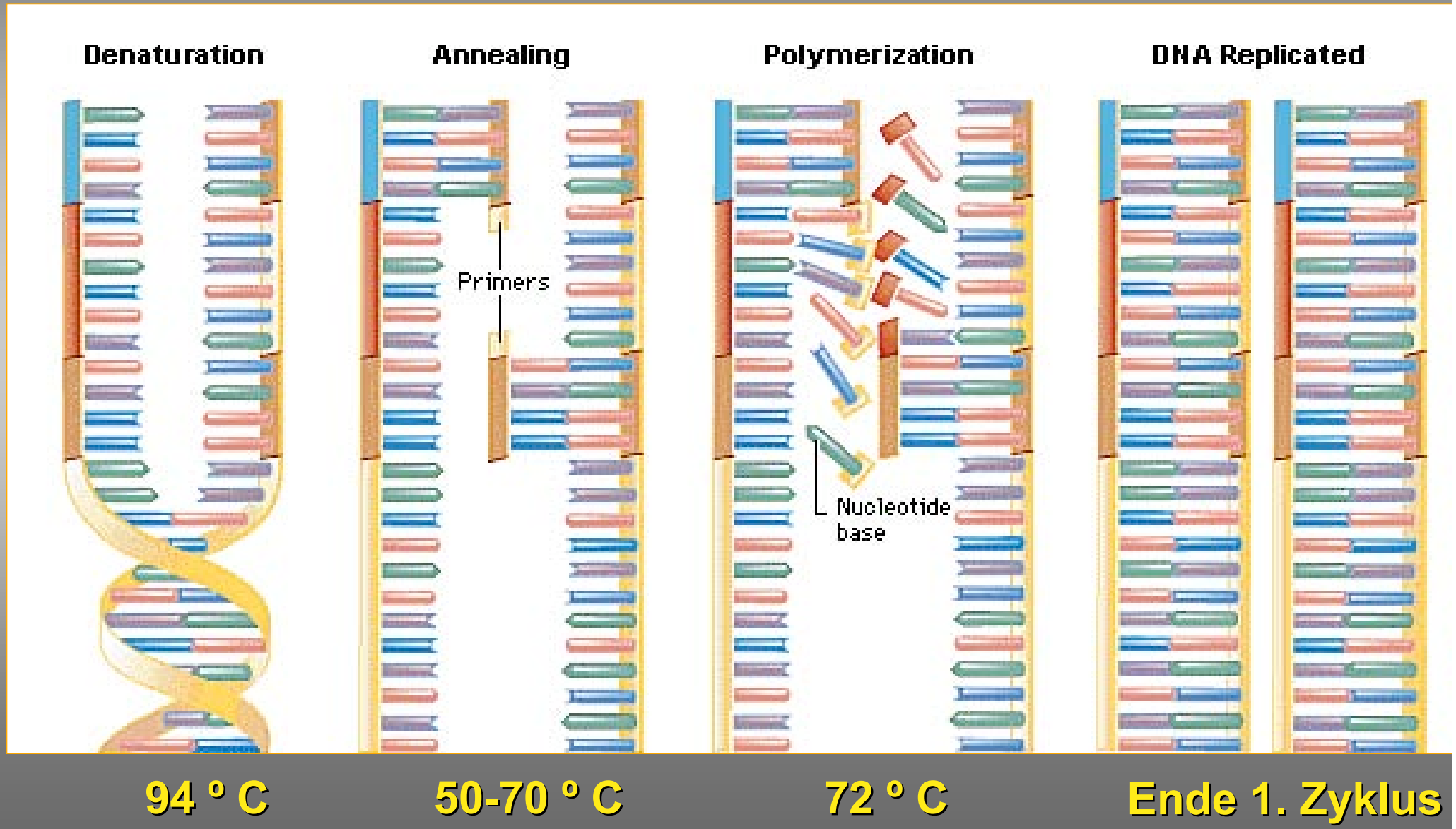
Kombination von cDNA Synthese und PCR-Amplifikation
zur Genexpressions-Detektion



Primerdesign für PCR ist entscheidend
für RT-PCR Gen-Expressionsanalysen

PCR: polymerase chain reaction:

Wiederholte Abfolge von 3 Temperatur/Inkubationsschritten

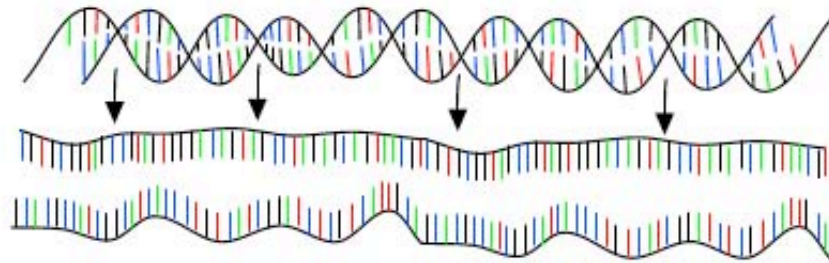


PCR : Polymerase Chain Reaction

30 - 40 cycles of 3 steps :

Step 1 : denaturation

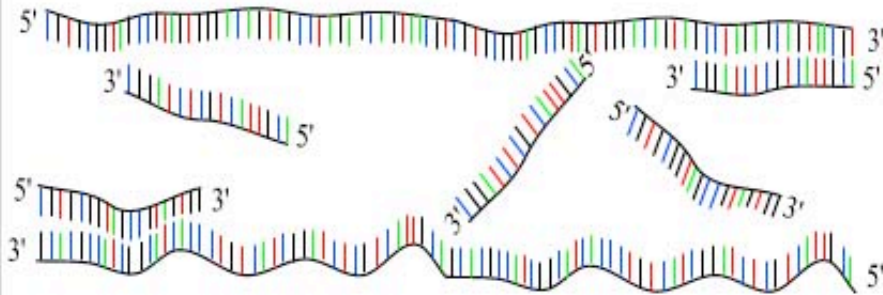
1 minut 94 °C



Step 2 : annealing

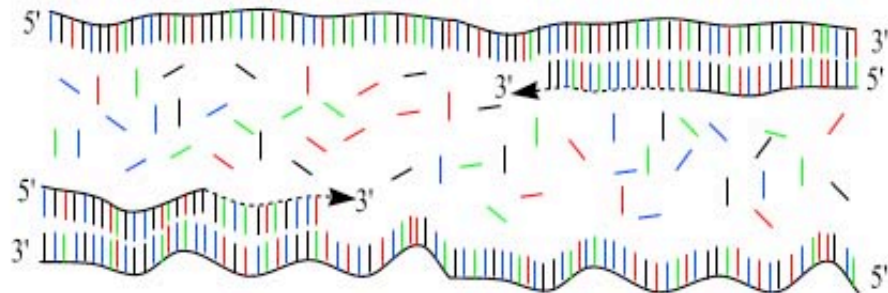
45 seconds 54 °C

forward and reverse primers !!!



Step 3 : extension

2 minutes 72 °C
only dNTP's



(Andy Vierstraete 1999)

Benötigt werden:

2 PCR-Primer
(Forward / Reverse)

dNTPs + Puffer
Taq-Polimerase
Thermozykler (PCR-Gerät)

Exponentielle DNA Vermehrung

nach jedem PCR Zyklus
Verdoppelt sich die
Zielmolekül Anzahl:

2^n
DNA Moleküle
nach n PCR-Zyklen

Exponentielle DNA-Vermehrung durch PCR

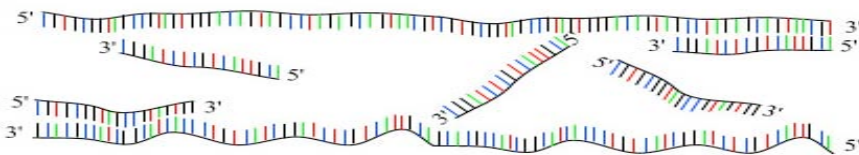
Abfolge von 3 Temperaturstufen

A

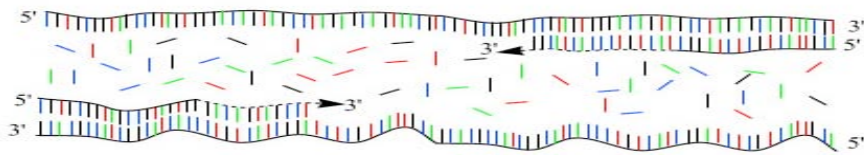
1. DNA-denaturation



2. Primer-annealing



3. Primer-extension

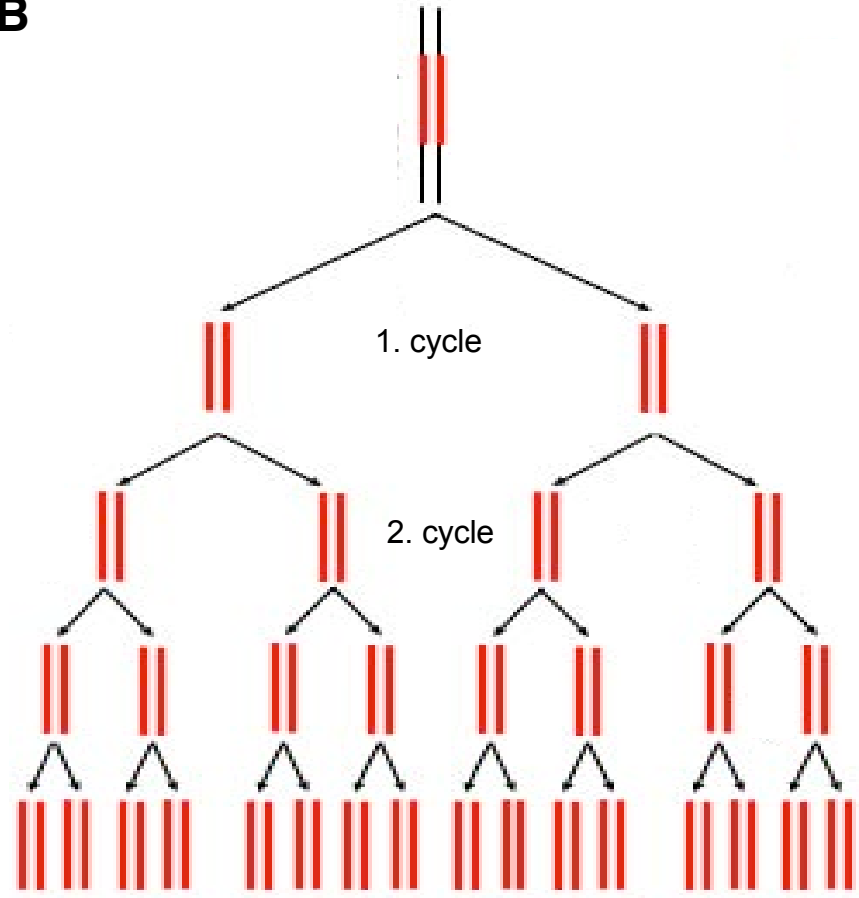


Exponentielle DNA-Vermehrung

B

1. cycle

2. cycle



Gen-Expressionsanalyse einzelner Zellen mittels RT-PCR

1. Isolation einzelner Zellen / mRNA (z.T. Funktionsanalyse)

(Laser Mikrodissektion, Patch-clamp Technik)

2. cDNA Synthese (Reverse Transkription der mRNA)

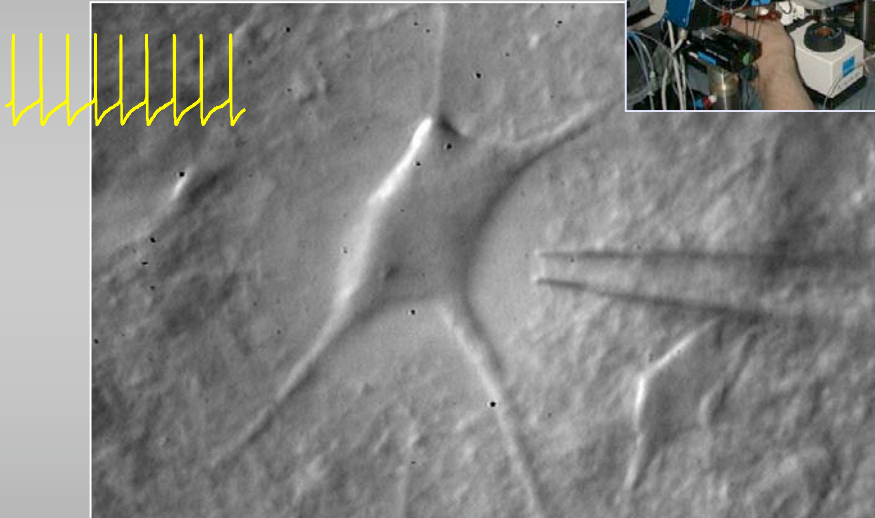
(Reverse Transkription -RT)

3. PCR Amplifikation der zu untersuchenden Gene/cDNAs

(Primer-Design, qualitative PCR, quantitative real-time PCR)

Zwei verschiedenen Methoden zur Isolierung einzelner Zellen / mRNA aus Gewebe (zB Hirnschnitte)

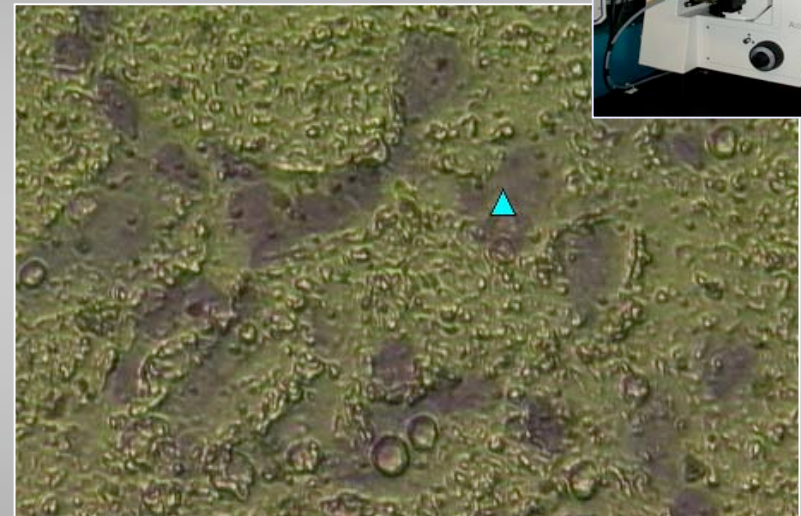
1. *Living neurons brain-slices*



Zytoplasma-Ernte mittels **Patch-Pipette**
(Patch-Clamp Set-up):

Funktion und Genexpression

2. *Post mortem fixed brain-sections*



Kontaktfreie **Laser-Microdissection** / Katapu
(PALM-Set-up)

Analyse von post mortem (humanen!) Proben

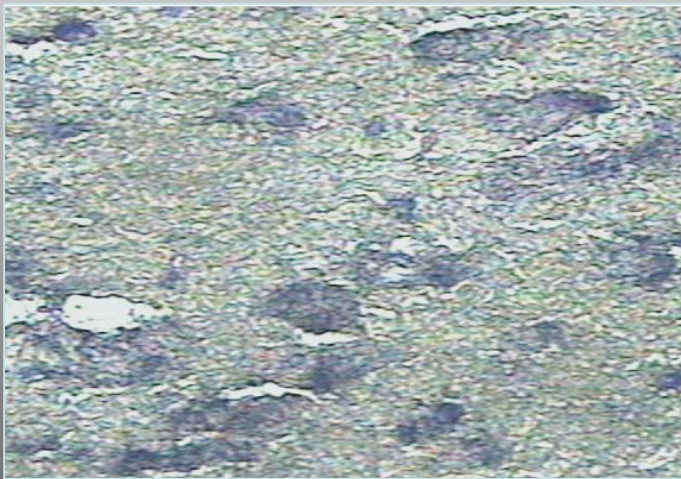
cDNA-Synthese, Gene-expression Profiling (RT-PCR)

Isolierung einzelner Zellen aus Gewebe (zB Hirnschnitte)

Methode: Laser-Mikrodissection (PALM-System)

Erlaubt schnelle "unkomplizierte" Isolierung einzelner Zellen

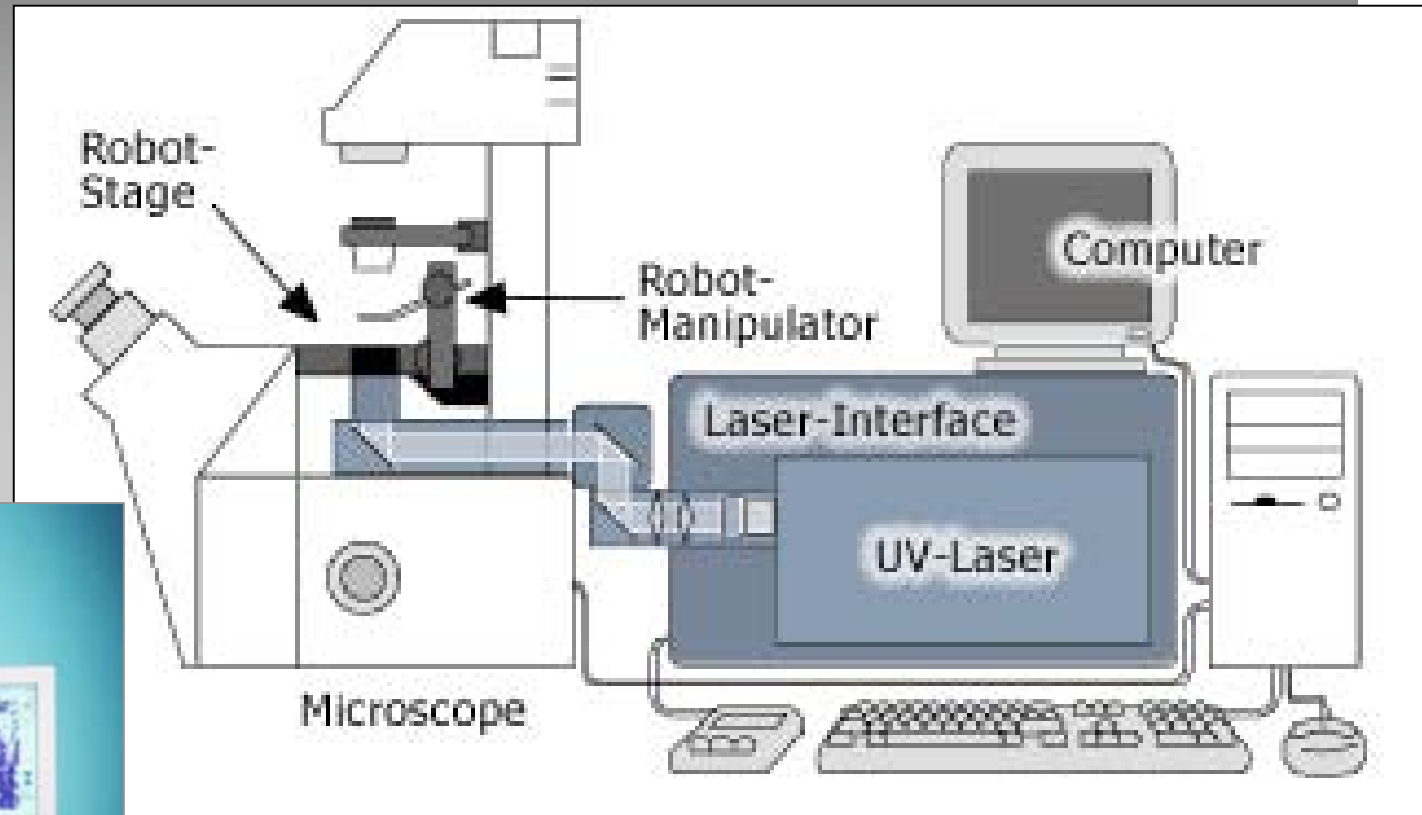
Erlaubt Analyse von fixiertem (*post mortem*) Material



PALM: Photo Ablation and Laser Microdissection

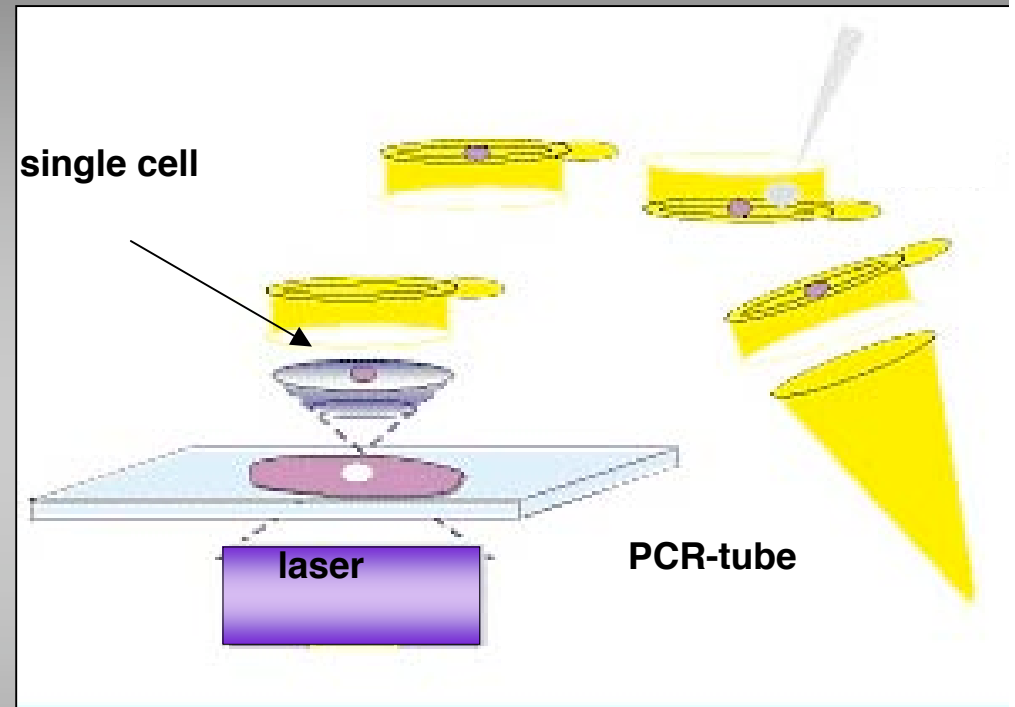
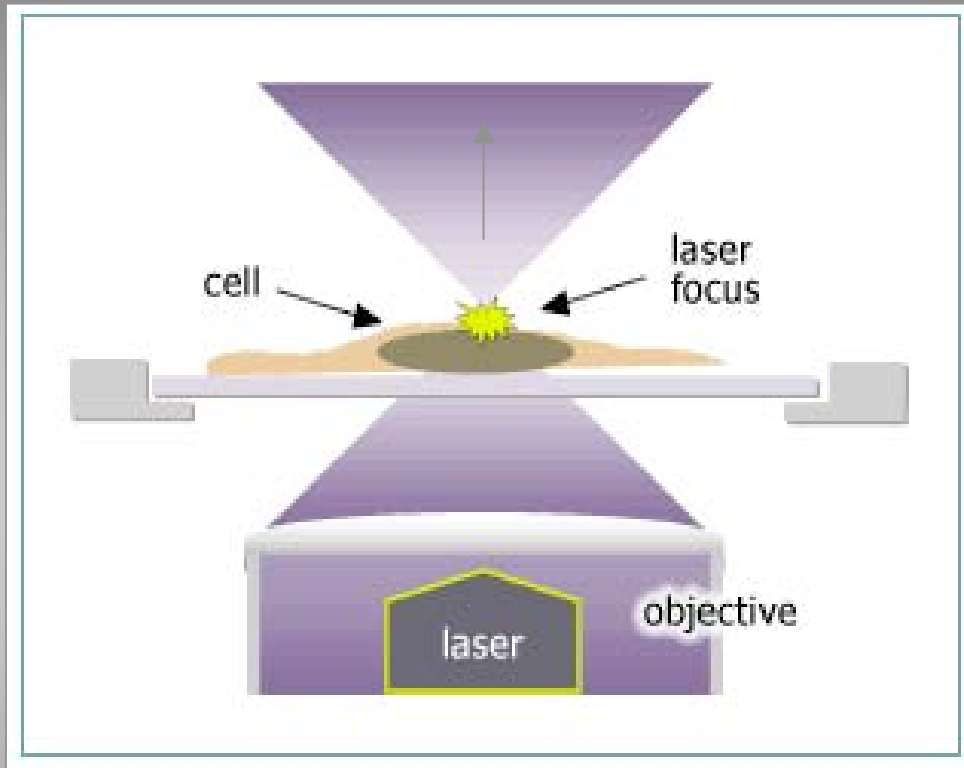


PALM: Photo Ablation and Laser Microdissection



337 nm Nitrogen (UV-A) Laser

Laser Microdissection und Katapulting einzelner Neuronen

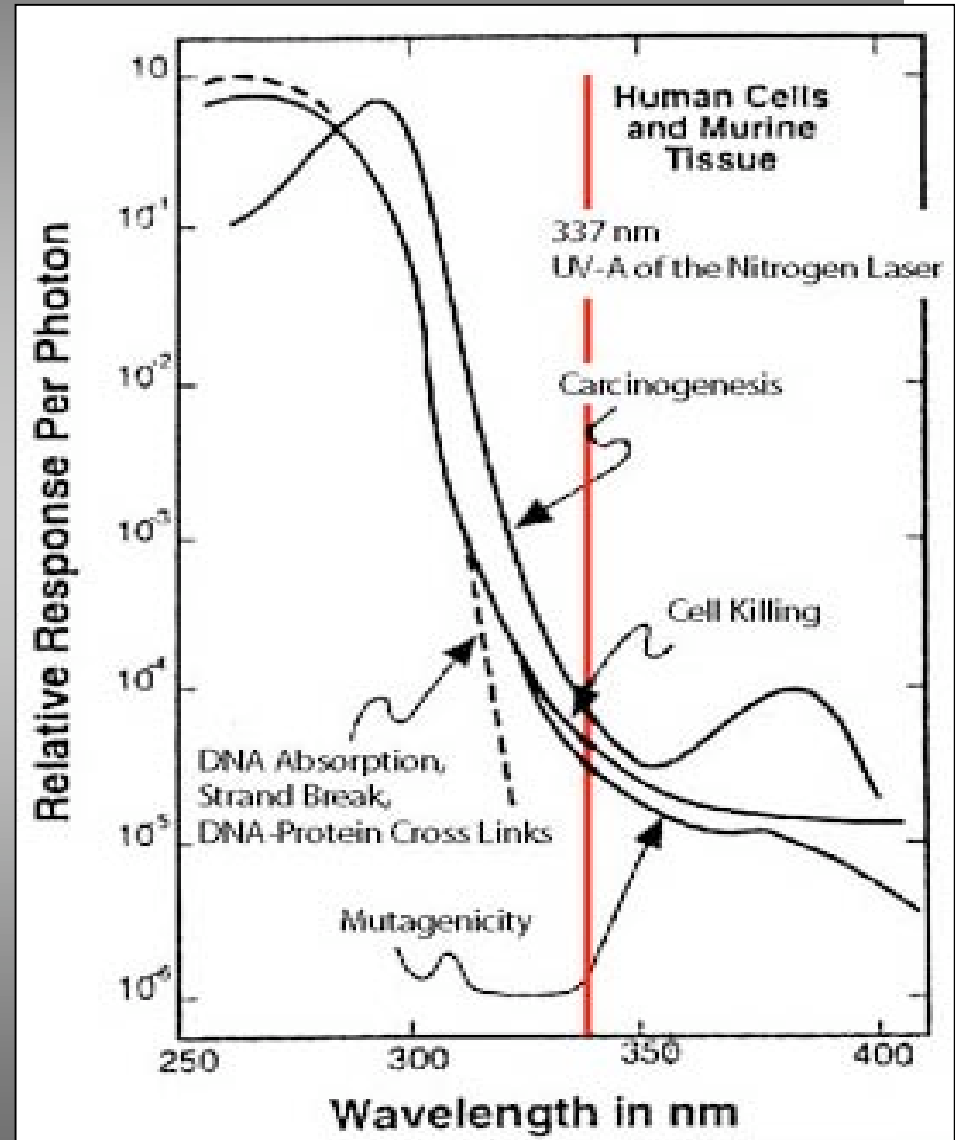
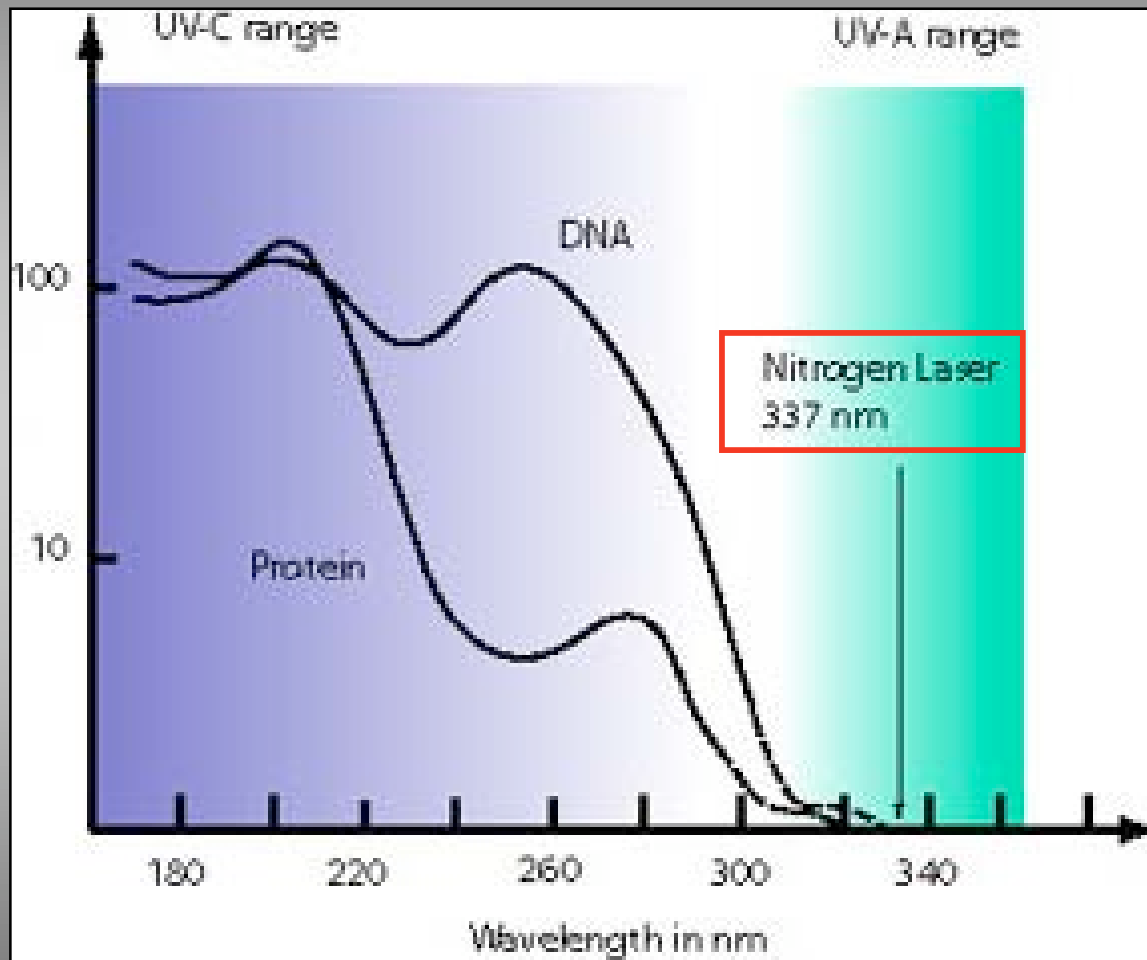


Ablative Photo-Decompensation (APD):

Hohe Photondichte, Energie höher als Molekülbindungsenergie,
Microplasma-Formation/Collapsing (nanosec), Expansion, keine Hitze

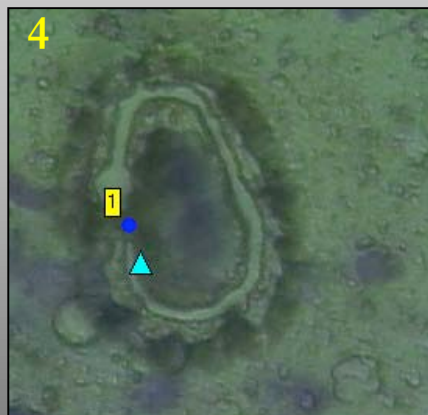
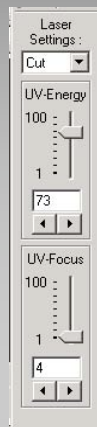
“where the laser hits, nothing is left that could be analyzed as biomolecule”

PALM: Photo Ablation and Laser Microdissection



337 nm: keine DNA/RNA/Proteinschädigung

Laser-Microdissection eines dopaminergen Neurons



adhesive cap control

Gen-Expressionsanalyse nach Laser-Microdissection

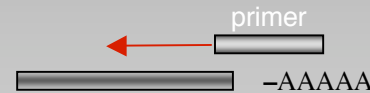
stained mouse brain section (10 μ m)



after laser dissection and catapulting



RT-PCR amplification of single-cell mRNA



Lysis of captured neurons

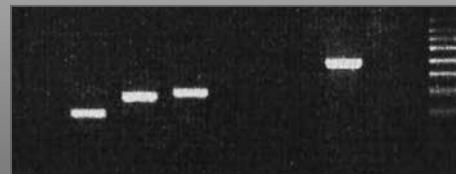
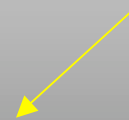


cDNA synthesis

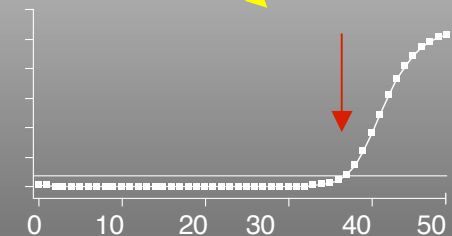
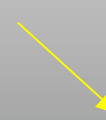


PCR amplification of target cDNAs

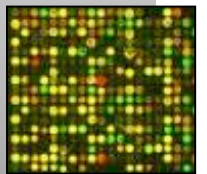
detection of gene expression



qualitative multiplex PCR



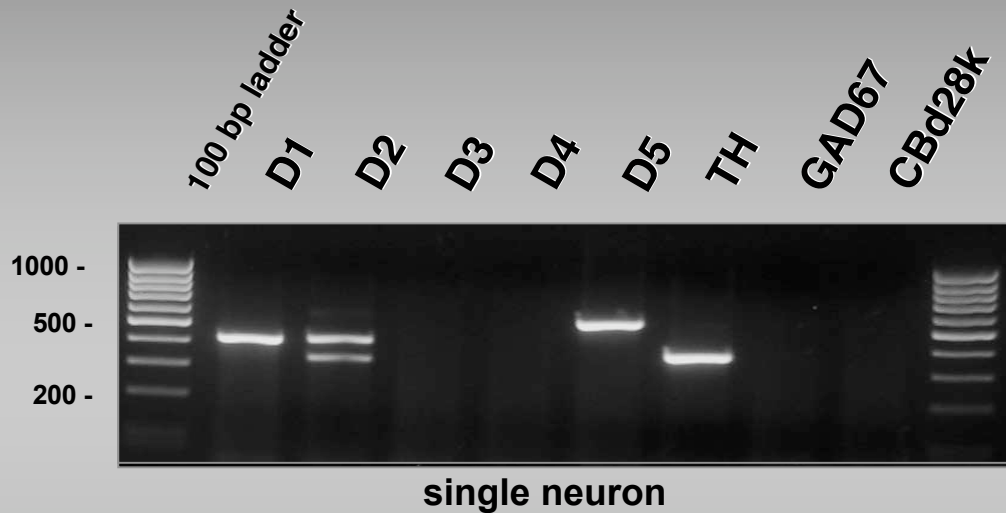
quantitative real-time PCR



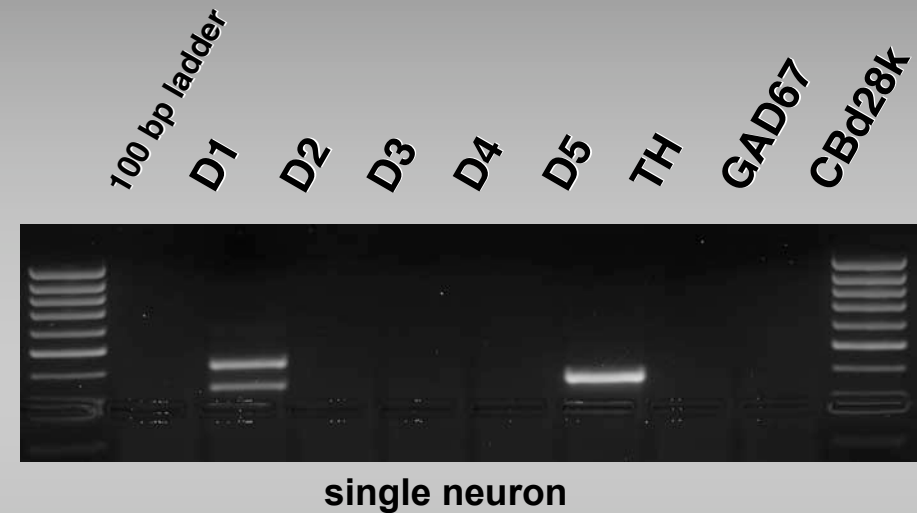
Sensitivität der Einzelzell RT-PCR Protokolls: Detektion der genomischen DNA (Zellkern)



Single cell RT-PCR without DNase digest



Single cell DNase digest prior RT-PCR



single neuron: RT minus control



single neuron: RT minus control

D1 and D5 primers: Intronless PCR product

Protocol robustly detects two copies of genomic DNA from single nucleus

Gen-Expressionsanalyse einzelner Zellen mittels Datenbank-Recherche und RT-PCR

A) Praktischer Teil im Labor: Methode

Laser Microdissektion und Isolation einzelner dopaminerge Neuronen

Jan Gründemann

B) Praktische Teil am Computer: PCR-Theorie

Wie suche ich geeignete Primer für ein RT-PCR Experiment aus?

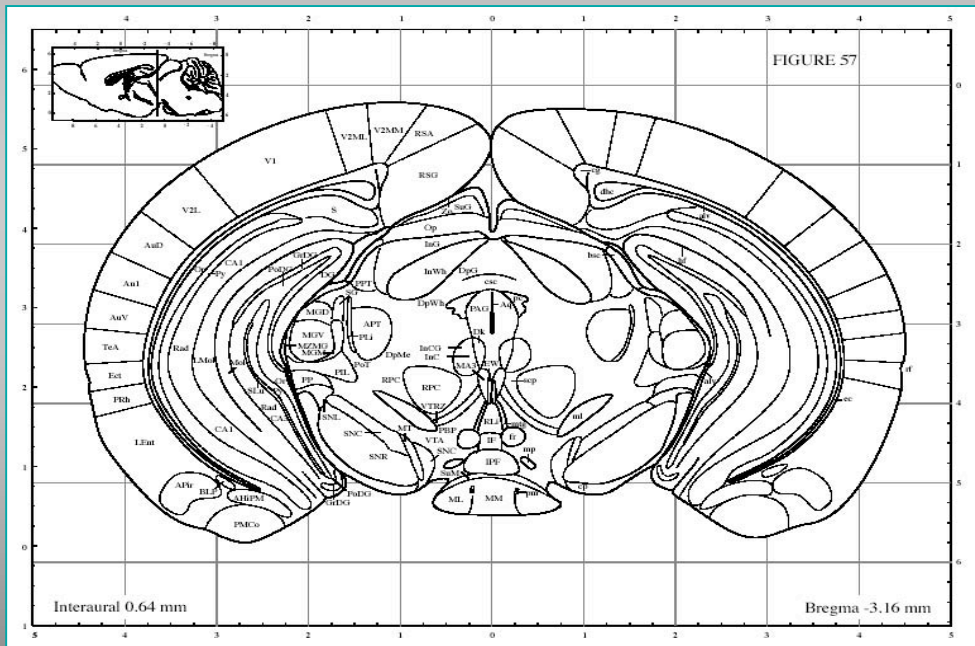
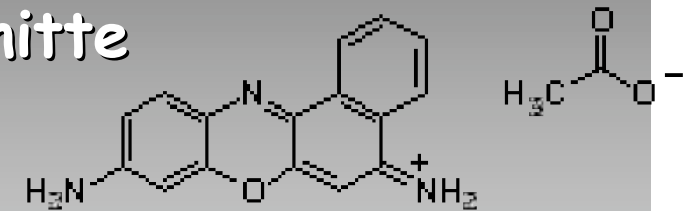
Anca Alexandru

C) Hintergrund (soviel wie gewünscht):

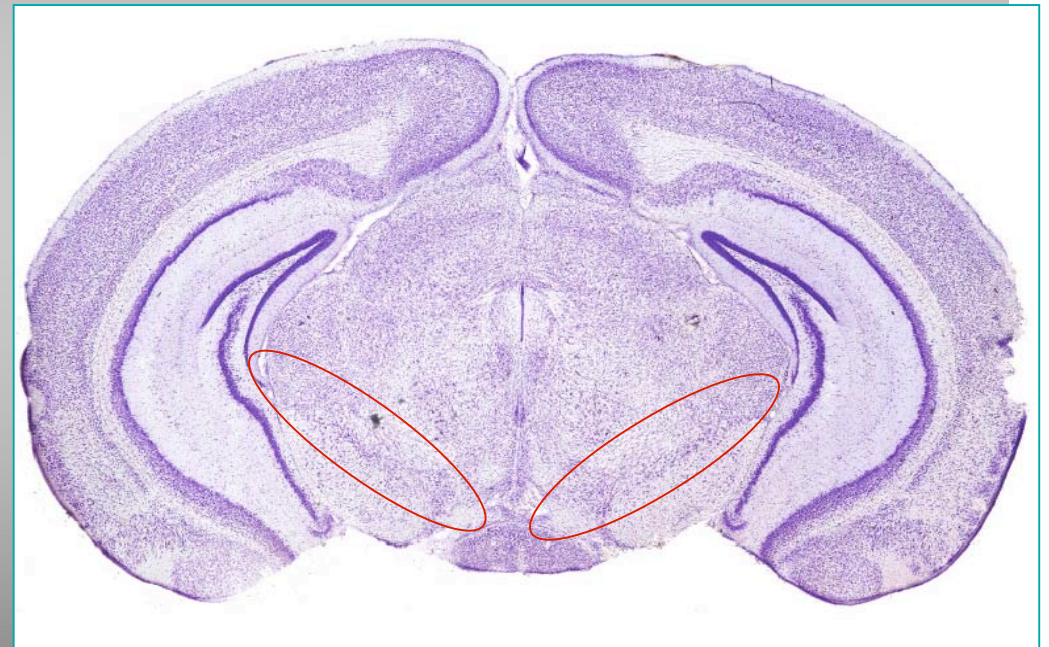
Warum Genexpressions-Analyse von dopaminerge Neuronen?

A) Auffinden der dopaminergen Mittelhirn-Neurone

Maus: koronale Mittelhirnschnitte

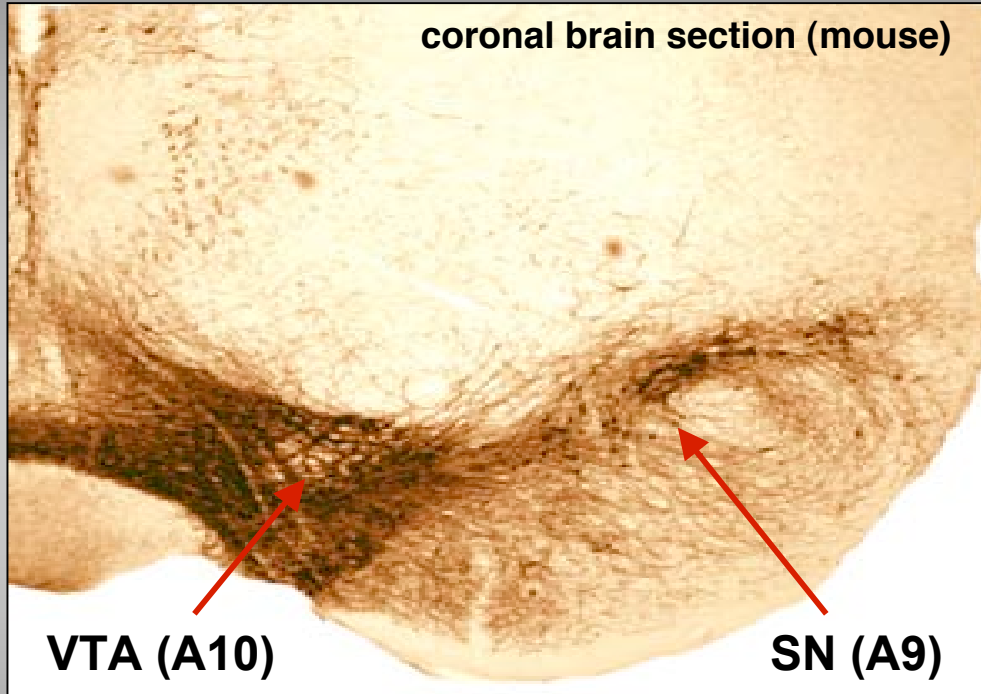


mouse brain atlas

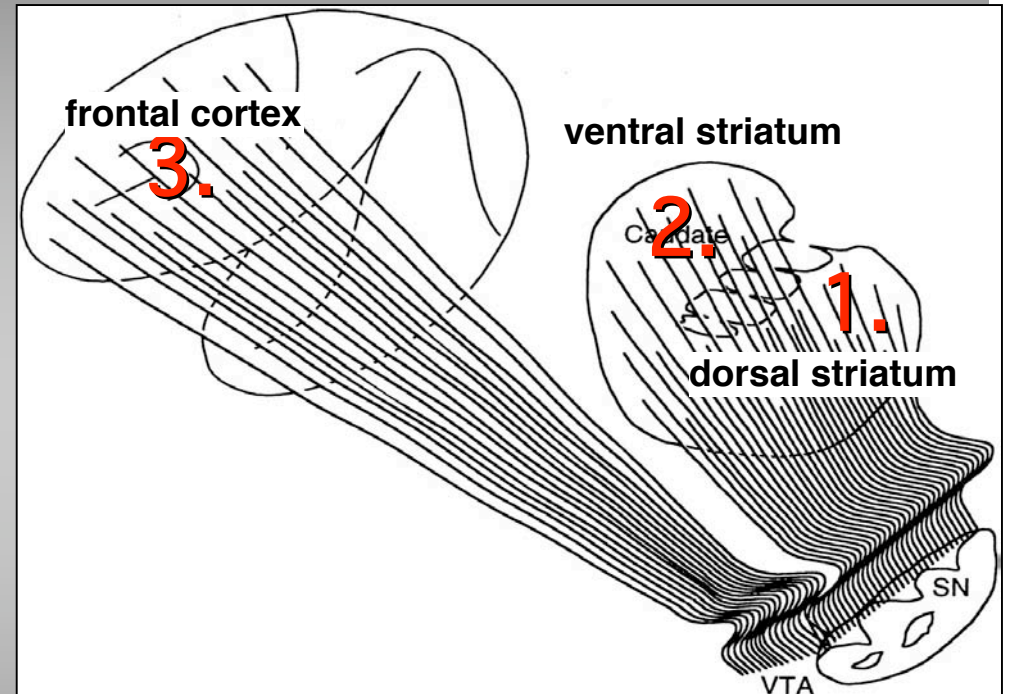


Zellfärbung (Nissl / Cresylviolet)

Drei verschiedene Typen von dopaminergen Neuronen: Drei verschiedene Projektionsgebiete



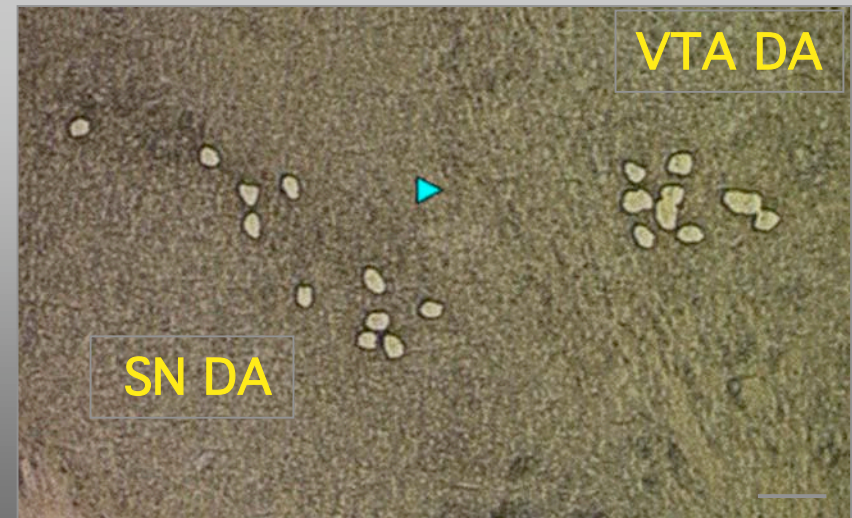
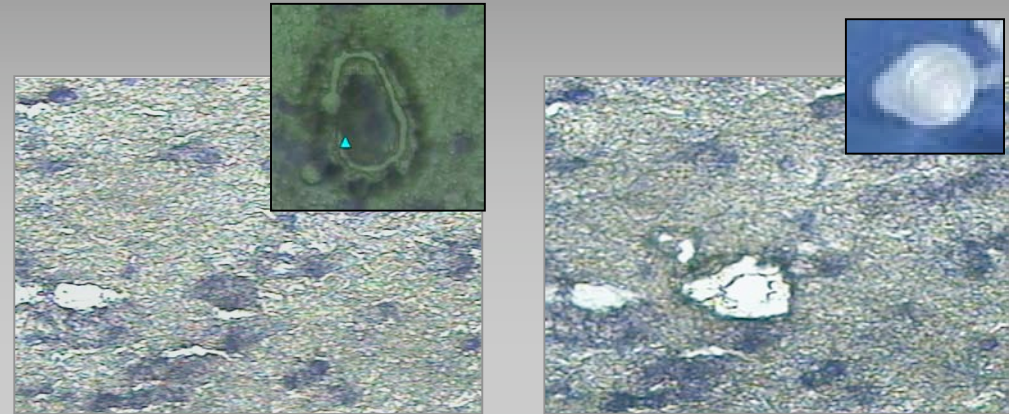
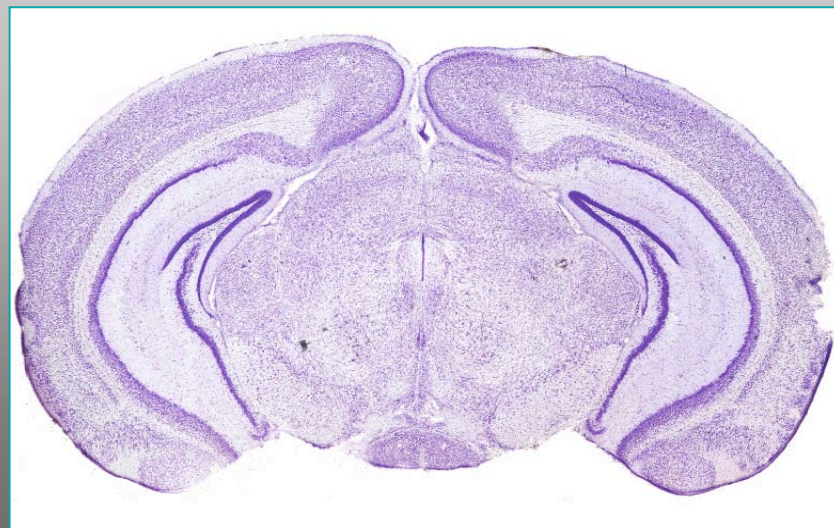
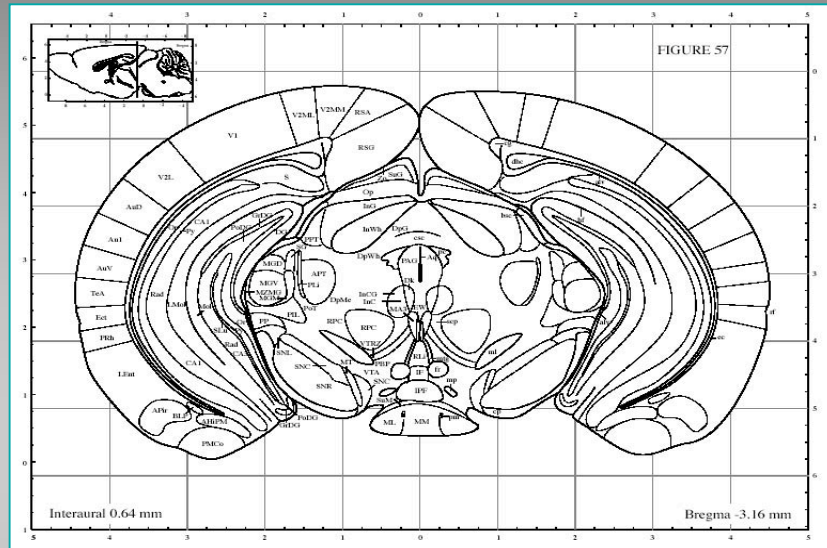
Tyrosine-hydroxylase (TH) immunostain



dopaminergic projections

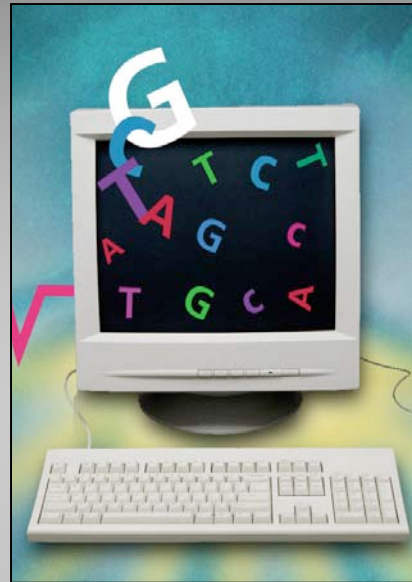
- | | | | | |
|----|------|--------------|----------------------------|------------------------------------|
| 1. | SN: | mesostriatal | voluntary movement | Parkinson's disease |
| 2. | VTA: | mesolimbic | motivation (reward) | Schizophrenia (+) / drug addiction |
| 3. | VTA: | mesocortical | cognition (working memory) | Schizophrenia (-) |

A) Laser-Mikrodissection einzelner Mittelhirn-Neurone



B) Praktische Teil am Computer: PCR-Theorie:

Wie suche ich Primer für ein RT-PCR Experiment aus ?



- Der Weg ist das (Praktikums) Ziel ! -

Aktualisierte Praktikumsunterlagen / Einführungsdias
auf meiner Homepage (unter Lehre Info):

<http://www.med.uni-marburg.de/d-einrichtungen/molneurophys/lehreinfo/1259/>

Design geeigneter Primer für RT-PCR Genexpressions-Analysen

1. Was weiss man über das zu untersuchende Gen bzw die Funktion des Genprodukt?

- Gibt es weitere Gen-Familienmitglieder oder Homologe oder Pseudogene ? (Primer-Spezifität)
- Datenbanken-Search: *NCBI-OMIM, Gencards, Pubmed* (zB Rewievs)

2. Auffinden der cDNA Sequenz (*NCBI Nucleotide*):

- Entscheidung für eine Sequenz, da meistens mehrere Genbankeinträge für ein Gen/Genprodukt:
- Aufschreiben der **Accession number** (einzigartige Identifikationsnummer "Personalausweis")
- Ausdrucken der cDNA Sequenz im "*Genbank-Format*" und im "*Fasta-Format*"

3. Auffinden der Genstruktur bzw der Intron/Exon Übergänge (*Ensemble*):

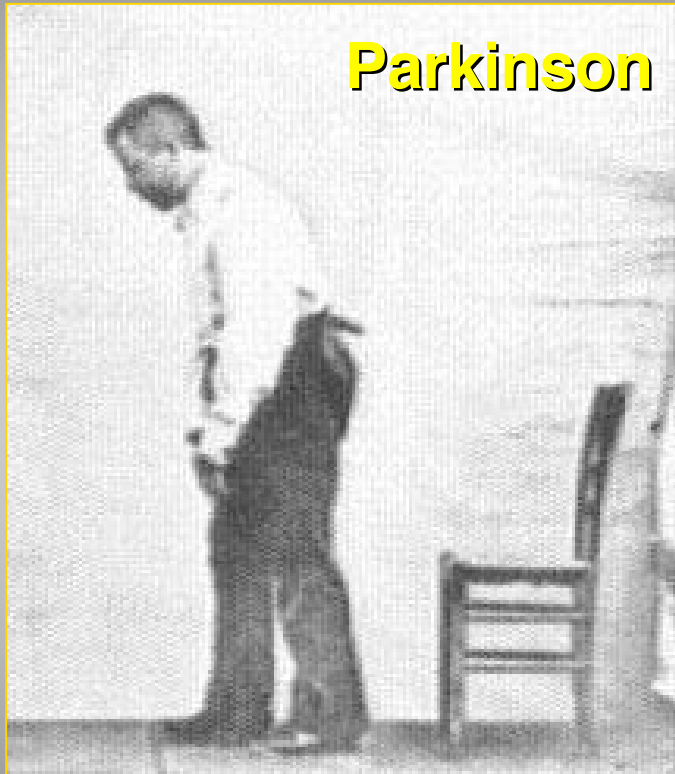
- Primer / PCR-Produkt sollte mindestens ein Intron-Exon Übergang überspannen (warum?)
- Datenbank "*Ensemble*" ist hierfür am besten geeignet (Achtung: andere Gen-Einträge als NCBI!)

4. Auswahl "geeigneter", spezifischer Primer:

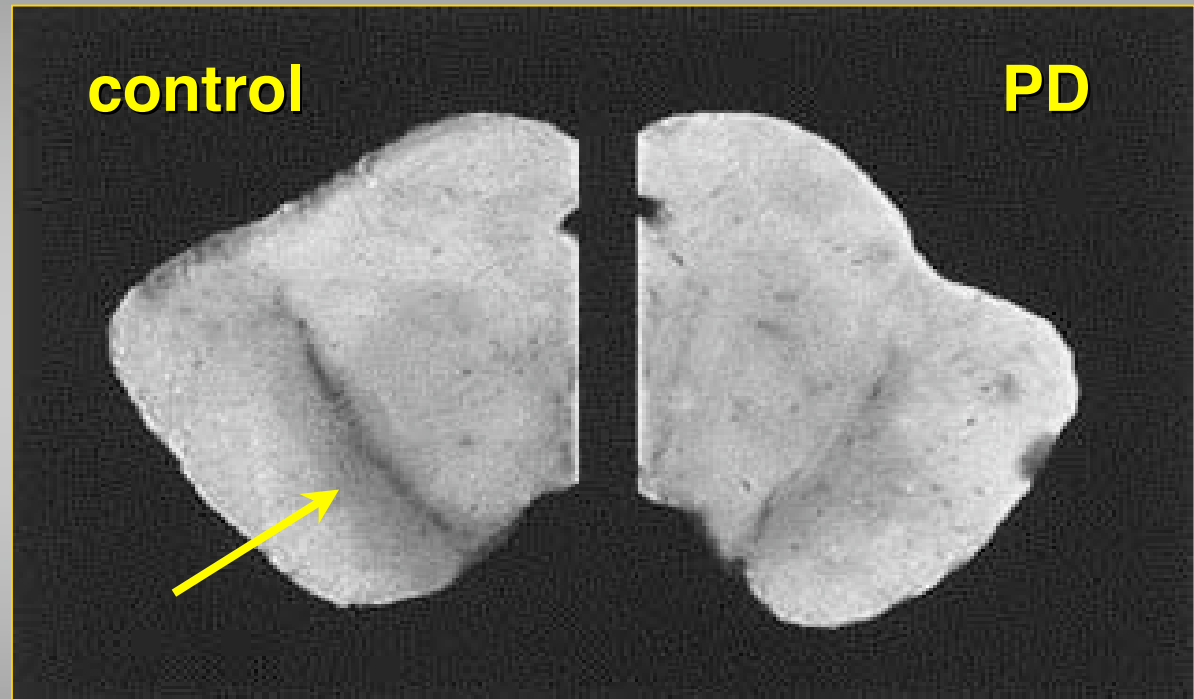
- Einlesen der cDNA Sequenz in ein *Primer-Design Programm* (Länge, Annealing-Temp, GC-Gehalt)
- Auswahl je eines geeigneten Forward (sense) und Reverse (antisense) Primers:
Primer sollen nur Zielgen binden, mind. ein Intron überspannen, PCR Fragment ca 150- max.1000 bp
- Auffinden/Eintragen der Primersequenzen in der jeweiligen cDNA *Genbank-Sequenz*:
Beachte: Reverse Primer idR in 5' - 3' angegeben, müssen also reverse complement gelesen werden
- Überprüfen der Spezifität der ausgewählten Primer ("*BLAST short sequence search*")



Selective loss of dopaminergic neurons in Parkinson's disease

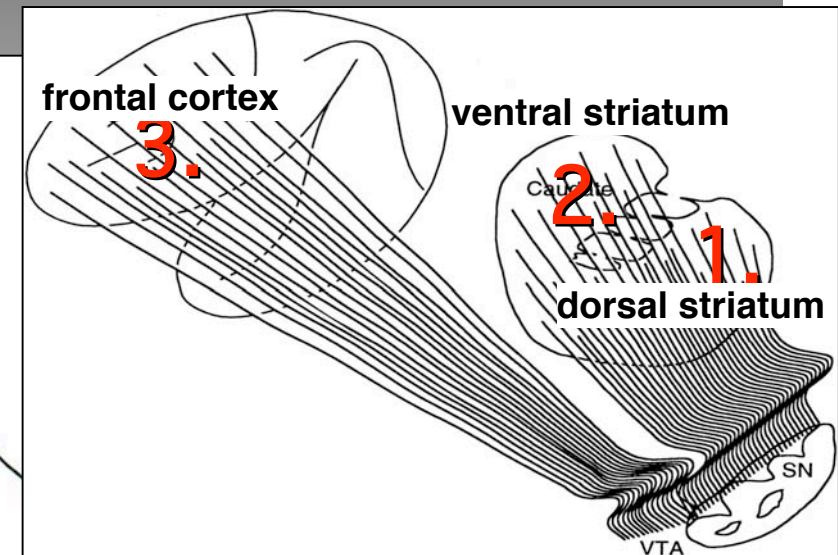
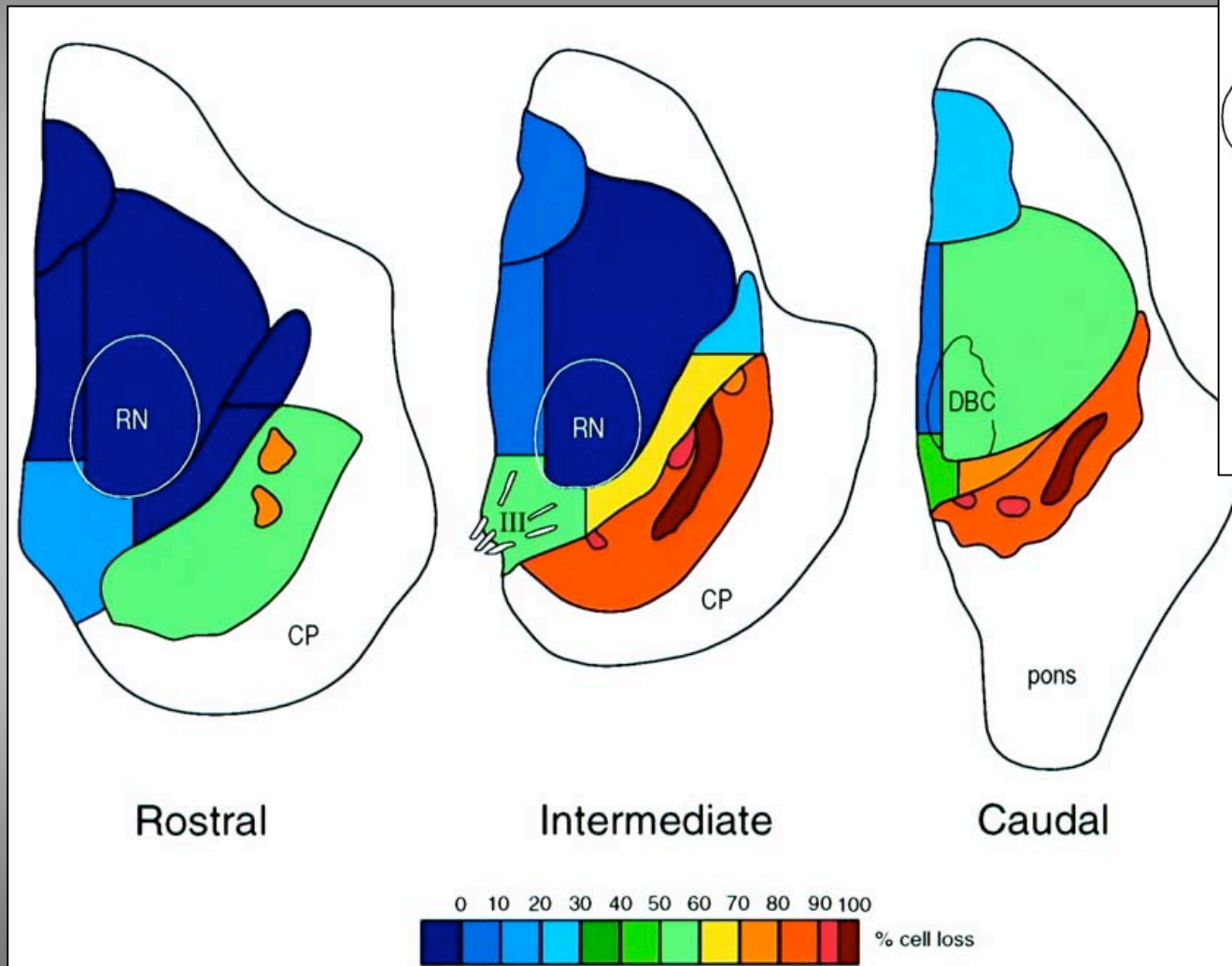


**rigor, tremor, akinesia
posturale instability**



cause : unknown
(mitochondrial (CX I) and proteasome dysfunction
as candidate mechanism for PD)

Differential vulnerability of DA neurons in Parkinson's Disease



dopaminergic projections

SN (A9) >> VTA (A10)

caudal >> rostral
nigrosomes >> matrix
CB neg >> CB pos

Damier et al. (1999a/b)

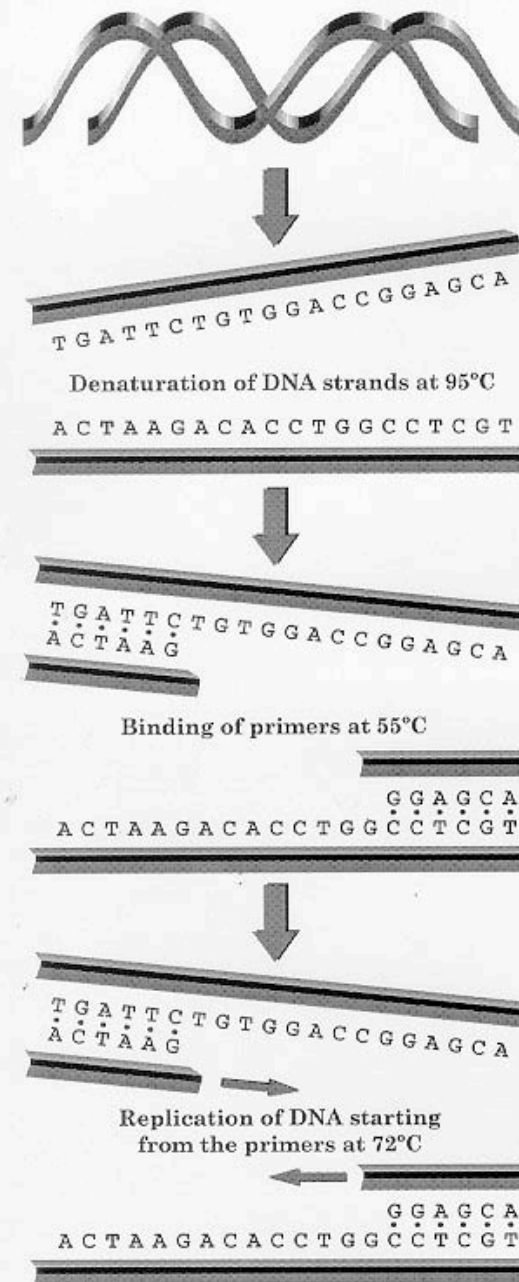
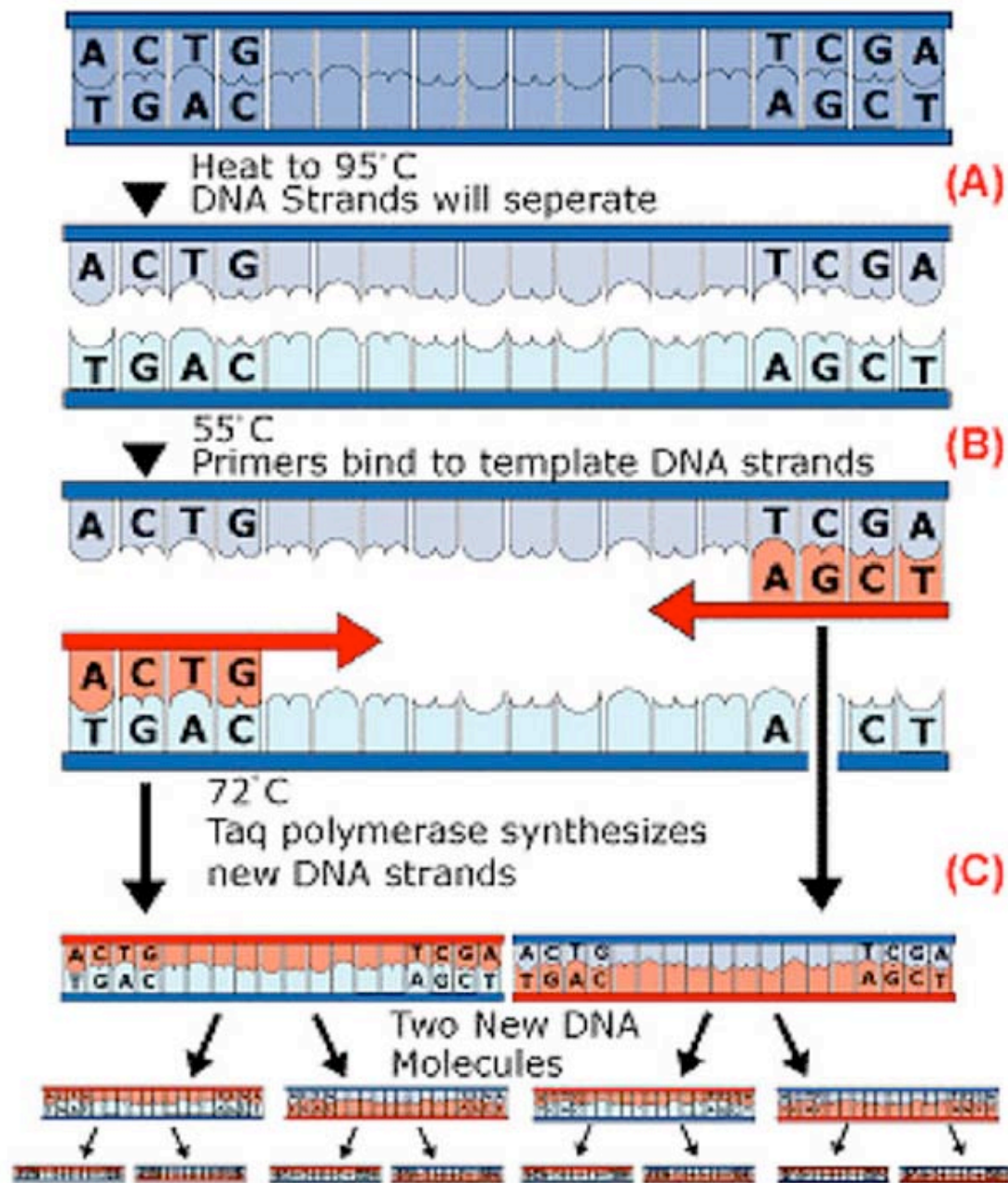
Forschungsschwerpunkt AG Liss:

Molekulare Mechanismen der differentiellen Vulnerabilität ?

1. Isolation der Einzelzell mRNA (Funktionsanalyse)
(Laser Mikrodissektion, Patch-clamp Technik)
2. cDNA Synthese (Reverse Transkription der mRNA)
(Reverse Transkription -RT)
3. PCR Amplifikation der zu untersuchenden Gene/cDNAs
(qualitative PCR, quantitative real-time PCR)

Vergleich der Genexpression
der hoch-empfindlichen und der resistenten DA Neuronen

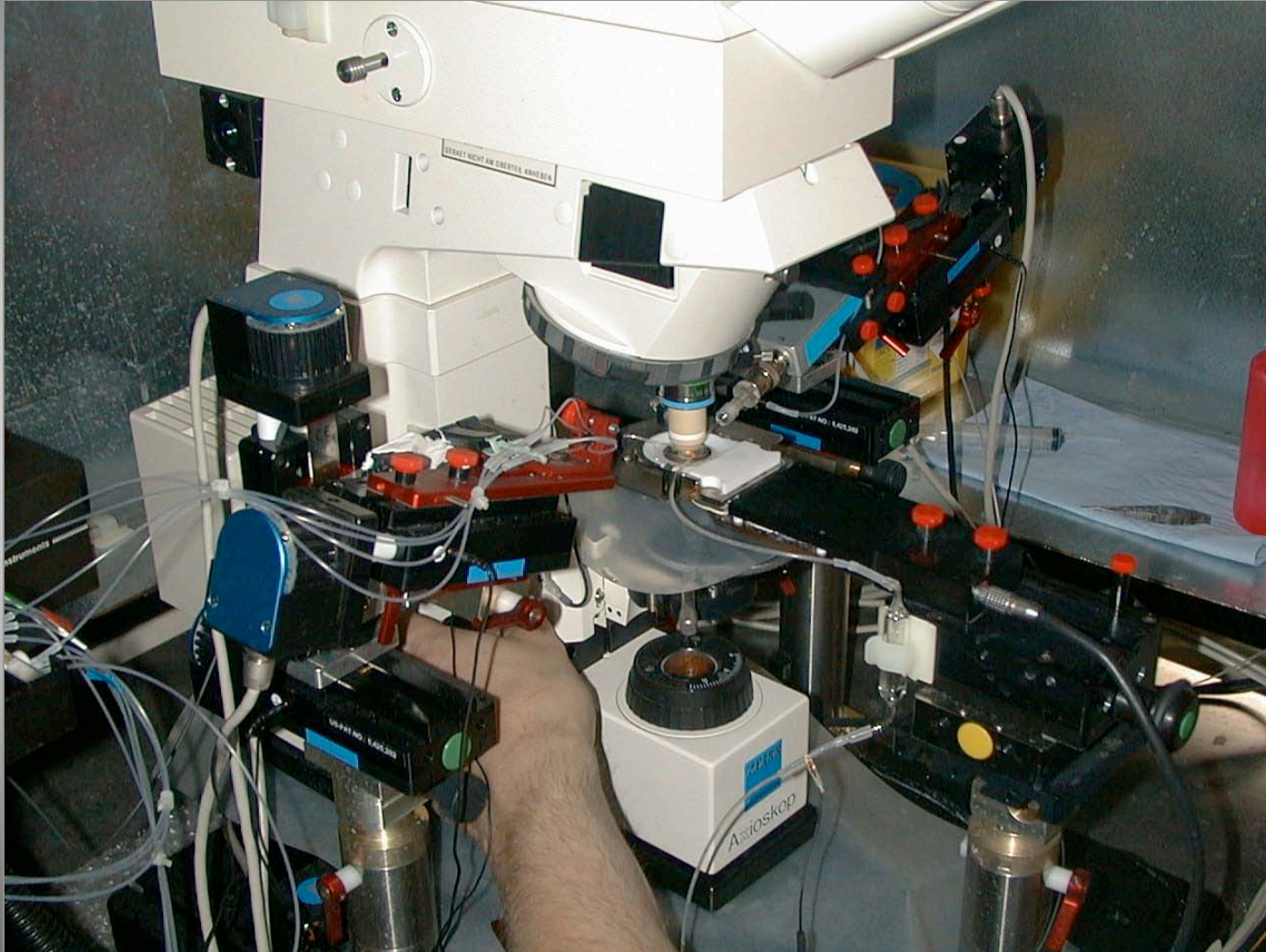




After each cycle heating and cooling the number of copies of the specified DNA sequence is doubled

After 1-2 hours millions of copies have been produced

Hirnschnitt Patch-clamp Technik



Patch-clamp technique - principle

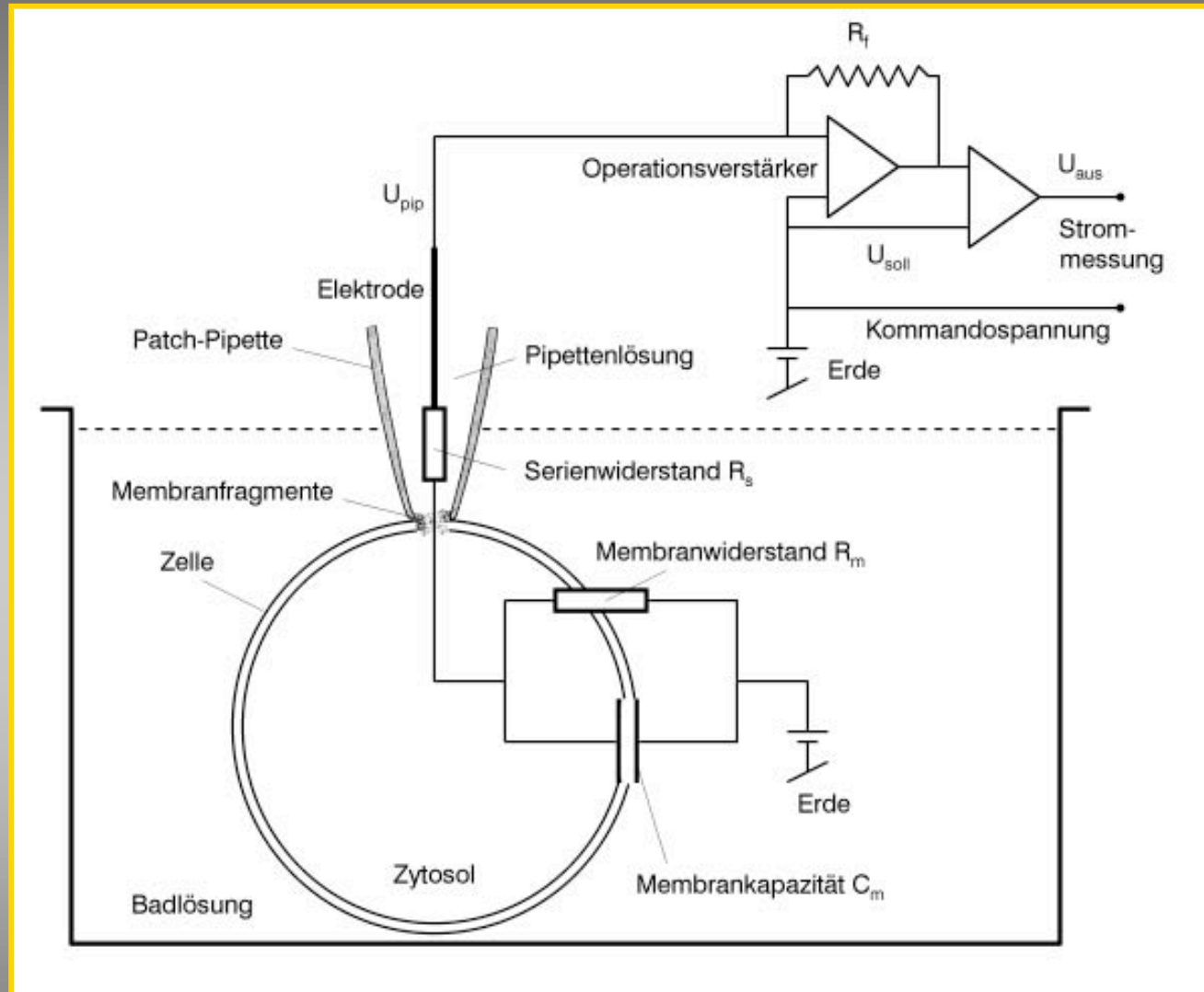
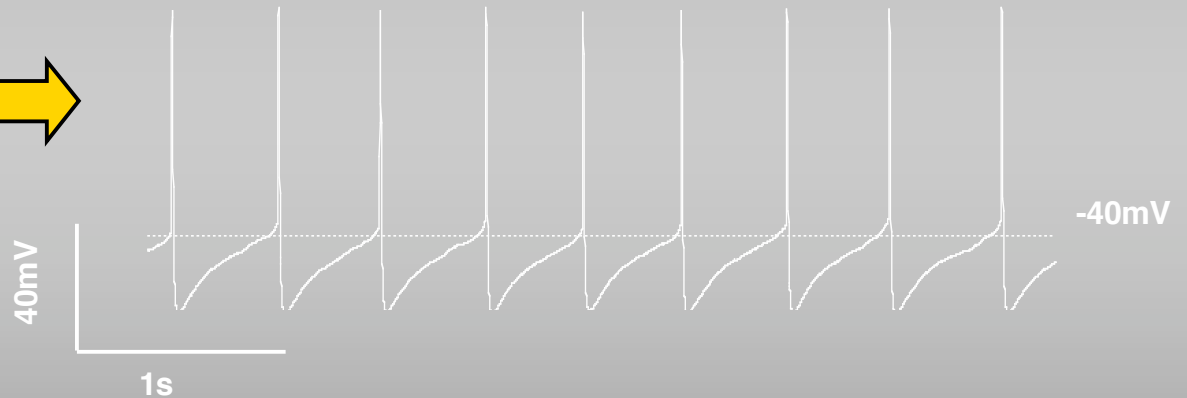
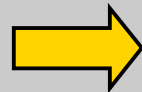


Fig. 1: Simplified circuit of a patch-clamp amplifier and substitute-circuit of the whole-cell-configuration: R_f : Feedback resistance, U_{soll} : Nominal – or debit-potential, U_{pip} : Pipette-potential, U_{aus} : Exit-potential proportionally to the current. The circuit of the amplifier, that represents a so-called current-potential transformer, is accommodated near the pipette in a small box (the preamplifier). After breaking through the membrane, the so called whole-cell configuration is reached. Membrane fragments and other cell-material are sucked near the tip of the patch-pipette increasing the series-resistance R_s .

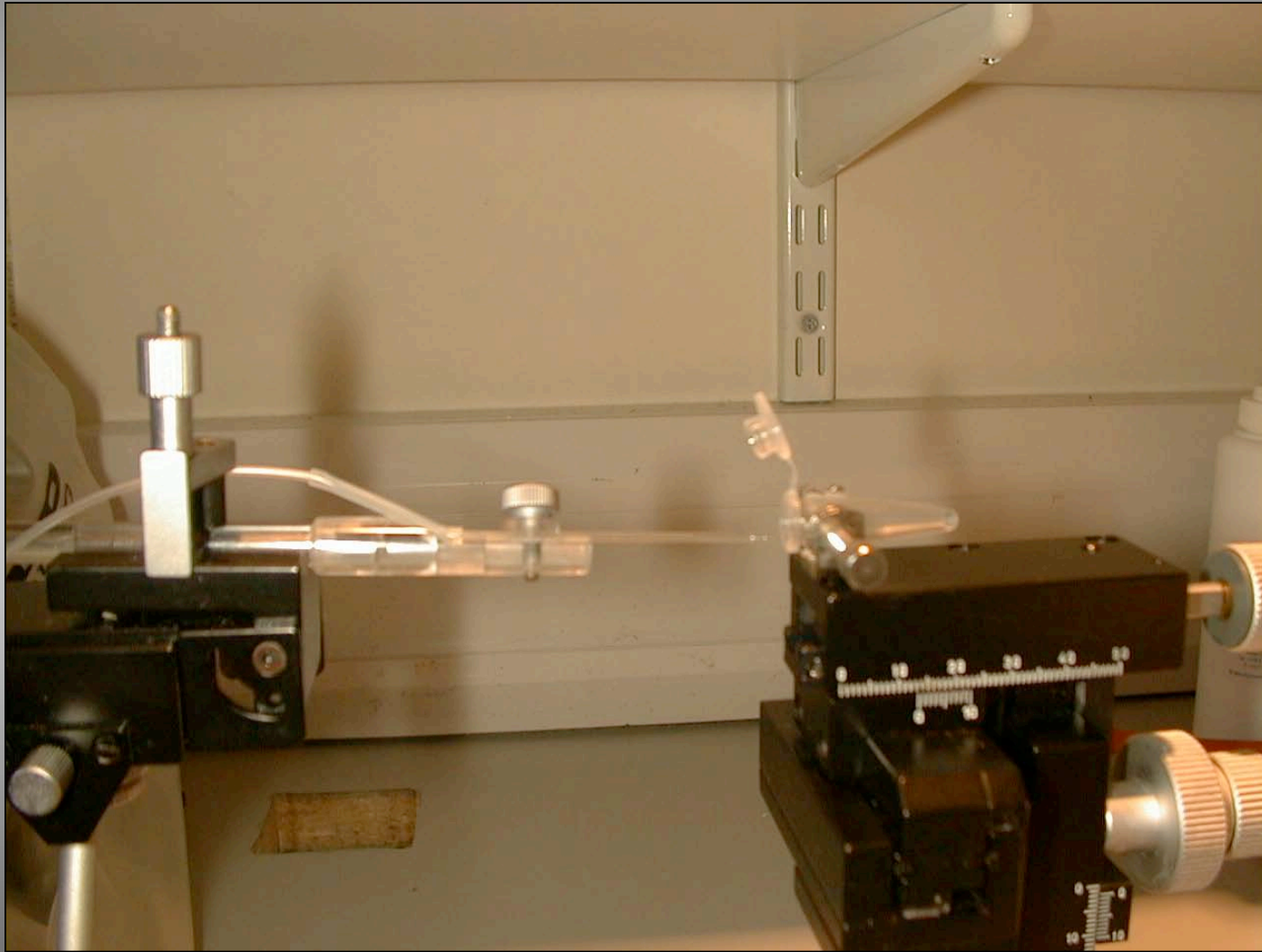
Untersuchung der spontanen Aktivitaet

brain-slice whole cell current clamp recording



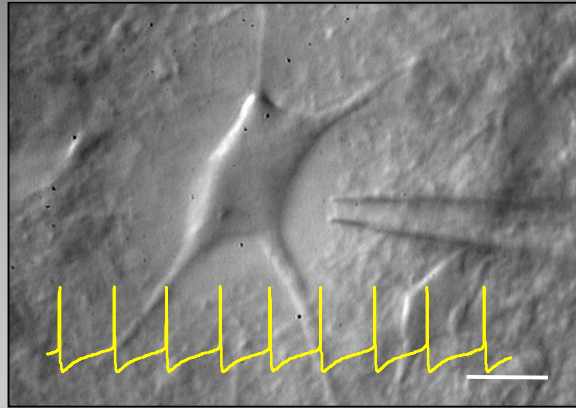
$1.3 \pm 1.1 \text{ Hz}$

Ueberfuehren des Einzelzell Zytoplasmas in ein PCR Tube



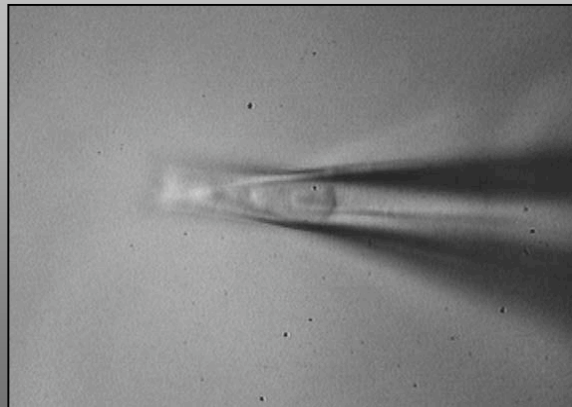
Einzelzell RT-PCR nach Patch-clamp Analyse

electrophysiological characterisation

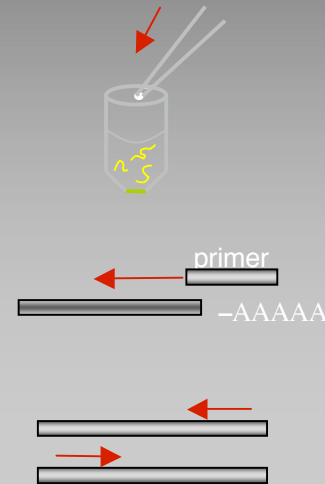


brain-slice patch-clamp technique

harvesting of single-cell cytoplasm



RT-PCR amplification of single-cell mRNA



expelling of pipette contents

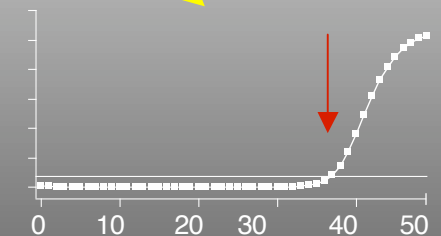
cDNA synthesis

PCR amplification of target cDNAs

detection of gene expression

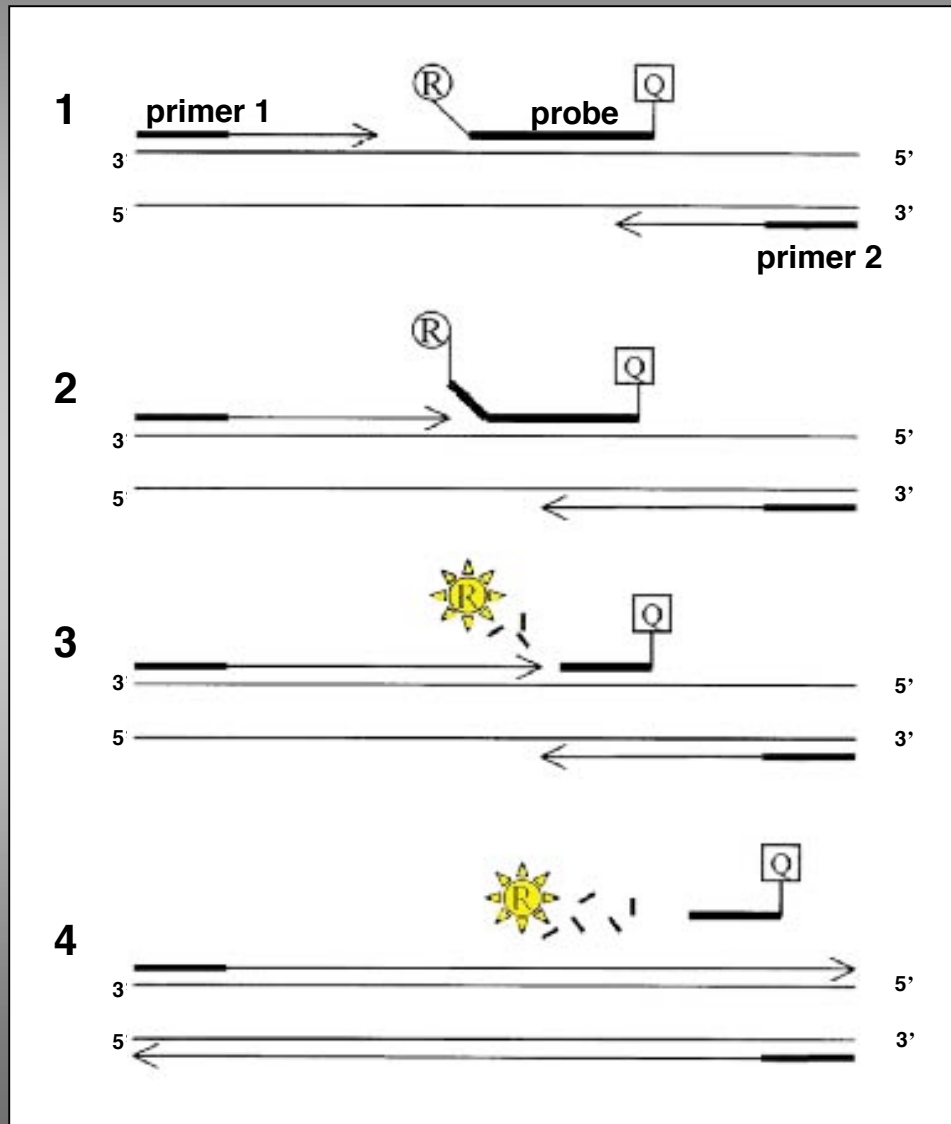


qualitative multiplex PCR



quantitative real-time PC

Quantitative real-time PCR: TaqMan primer / hybridisation-probe PCR assay



annealing of primer and hybridisation-probe
(5'-reporter / 3'-quencher)

primer-extension

5' exonuclease activity of Taq Polymerase:
Hydrolysis of the probe, release of fluorescence

detection of fluorescence increase