- 2 Nadal, A. *et al.* (2001) The estrogen trinity: membrane, cytosolic and nuclear effects. *News Physiol. Sci.* 16, 251–255
- 3 Valverde, M.A. et al. (1999) Acute activation of Maxi-K channels (hSlo) by estradiol binding to the β subunit. Science 285, 1929–1931
- 4 Falkenstein, E. *et al.* (2000) Multiple actions of steroid hormones a focus on rapid, nongenomic effects. *Pharmacol. Rev.* 52, 513–556
- 5 Kelly, M.J. and Levin, E.R. (2001) Rapid actions of plasma membrane estrogen receptors. *Trends Endocrinol. Metab.* 12, 152–156
- 6 Pappas, T.C. *et al.* (1995) Membrane estrogen receptors identified by multiple antibody labeling and impeded-ligand binding. *FASEB J.* 9, 404–410
- 7 Razandi, M. et al. (1999) Cell membrane and nuclear estrogen receptors (ERs) originate from a single transcript: studies of ERα and ERβ expressed in Chinese hamster ovary cells. Mol. Endocrinol. 13, 307–319
- 8 Wade, C.B. *et al.* (2001) Estrogen receptor (ER) α and ER β exhibit unique pharmacologic properties when coupled to activation of the mitogen-activated protein kinase pathway. *Endocrinology* 142, 2336–2342
- 9 Sonnhampmer, E.L. et al. (1998) A hidden Markov model for predicting transmembrane helices in protein sequences. Proc. Int. Conf. Intell. Syst. Mol. Biol. 6, 175–182
- 10 Filardo, E.J. et al. (2000) Estrogen induced activation of Erk-1 and Erk-2 requires the G protein-coupled receptor homolog, GPR30, and occurs via trans-activation of the epidermal growth factor receptor through release of HB-EGF. Mol. Endocrinol. 14, 1649–1660
- 11 Wyckoff, M.H. *et al.* (2001) Plasma membrane estrogen receptors are coupled to eNOS through Gai. *J. Biol. Chem.* 276, 27071–27076
- 12 Nadal, A. *et al.* (1998) Rapid insulinotropic effect of 17β -estradiol via a plasma

membrane receptor. FASEB J. 12, 1341-1348

- 13 Luconi, M. et al. (1999) Identification and characterization of a novel functional estrogen receptor on human sperm membrane that interferes with progesterone effects. J. Clin. Endocrinol. Metab. 84, 1670–1678
- 14 Monje, P. and Boland, R. (1999) Characterization of membrane estrogen binding proteins from rabbit uterus. *Mol. Cell Endocrinol.* 147, 75–84.
- 15 Zheng, J. and Ramirez, V.D. (1999) Purification and identification of an estrogen binding protein from rat brain: oligomycin sensitivity-conferring protein (OSCP), a subunit of mitochondrial F0F1-ATP synthase/ATPase. J. Steroid Biochem. Mol. Biol. 68, 65–75
- 16 Powell, C.E. et al. (2001) Identification and characterization of membrane estrogen receptor from MCF7 estrogen-target cells. J. Steroid Biochem. Mol. Biol. 77, 97–108
- 17 Gu, Q. et al. (1999) Rapid action of 17β-estradiol on kainate-induced currents in hippocampal neurons lacking intracellular estrogen receptors. *Endocrinology* 140, 660–666
- 18 Toran-Allerand, C.D. (2000) Novel sites and mechanisms of oestrogen action in the brain. *Novartis Found. Symp.* 230, 56–69; Discussion 69–73
- 19 Das, S.K. et al. (1997) Estrogenic responses in estrogen receptor-α deficient mice reveal a distinct estrogen signaling pathway. Proc. Natl. Acad. Sci. U. S. A. 94, 12786–12791
- 20 Nadal, A. *et al.* (2000) Nongenomic actions of estrogens and xenoestrogens by binding at a plasma membrane receptor unrelated to estrogen receptor α and estrogen receptor β . *Proc. Natl. Acad. Sci. U. S. A.* 97, 11603–11608
- 21 Ropero, A.B. *et al.* (1999) Non-genomic actions of 17β-oestradiol in mouse pancreatic β-cells are mediated by a cGMP-dependent

protein kinase. J. Physiol. 521, 397–407

- 22 Benten, W.P. et al. (2001) Estradiol signaling via sequestrable surface receptors. Endocrinology 142, 1669–1677
- 23 Condliffe, S.B. et al. (2001) 17β-Oestradiol acutely regulates Cl⁻ secretion in rat distal colonic epithelium. J. Physiol. 530, 47–54
- 24 Luconi, M. *et al.* (2001) Effetcs of estrogenic compounds on human spermatozoa: evidence for interaction with a nongenomic receptor for estrogen on human sperm membrane. *Mol. Cell Endocrinol.* 178, 39–45

Angel Nadal*

Ana B. Ropero

Bernat Soria

Institute of Bioengineering, Dept of Physiology, Miguel Hernández University, Campus of San Juan, Alicante 03550, Spain. *e-mail: nadal@umh.es

Esther Fuentes

Institute of Bioengineering, Division of Nutrition, Dept of Applied Biology, Miguel Hernández University, Campus of San Juan, Alicante 03550, Spain.

Chemical names

A role for neuronal K_{ATP} channels in metabolic control of the seizure gate

Birgit Liss and Jochen Roeper

ATP-sensitive K⁺ (K_{ATP}) channels are expressed in many different tissues including the brain, where they couple energy metabolism to cellular excitability. Although their classical role in insulin secretion in pancreatic β-cells is well understood, their neuronal function remains unclear. Now, an important study using knockout mice provides clear evidence that neuronal K_{ATP} channels are crucial players in counteracting seizure induction and propagation.

K⁺-selective ion channels belong to the superfamily of transmembrane proteins that couple a large diversity of physicochemical stimuli to the activity of their ion-conducting pores. These stimuli are predominantly transmembrane voltage gradients or the binding of a variety of extracellular and intracellular ligands. A large number of different genes encoding the principal pore-containing α -subunits and the regulatory β -subunits of K⁺ channels have been identified¹, consistent with the functional diversity of K⁺ channels. Indeed, two very different types of subunit team up to build ATP-sensitive $K^+(K_{ATP})$ channels: four pore-forming subunits of the inwardly rectifying K⁺ channel class, named Kir6.1 or Kir6.2, join together with four large β -subunits, the sulfonylurea receptors (SUR1 or SUR2A/B), which as members of the ABC (ATP binding cassette) transporter family possess two highly conserved nucleotide binding sites. Different combinations of these $K_{\mbox{\tiny ATP}}$ channel subunits give rise to functional channels with distinct biophysical, pharmacological and metabolic properties²⁻⁴.

The unique makeup of K_{ATP} channels allows them to convert the energy state of a cell (mainly expressed by the ATP:ADP ratio) directly into altered K⁺ fluxes through the plasma membrane: the open probability of these channels is increased with decreasing energy levels, thus adapting excitability to energy metabolism. The close coupling between cellular activity and metabolic state might be of particular significance in neurones because they expend the majority of their energy on ion pumps such as the Na⁺–K⁺-ATPase to maintain the electrochemical gradients that are perturbed by electrical signalling.

Physiological function of neuronal K_{ATP} channels

The functional role of K_{ATP} channels is best understood for pancreatic β -cells, where they couple blood glucose concentrations to insulin secretion⁵. A similar role in sensing central glucose levels and triggering pancreatic glucagon secretion has recently been demonstrated for $K_{\mbox{\tiny ATP}}$ channels in hypothalamic glucose responsive neurones⁶. However, K_{ATP} channels are expressed in many regions of the brain that are not involved in specific neuroendocrine functions such as glucose sensing. Therefore, a second, more general role for these channels has been proposed, in particular for excitable tissues such as muscle and neurones: the opening of $\mathbf{K}_{\! \mathrm{ATP}}$ channels in response to a decreased ATP:ADP ratio hyperpolarizes the neurone, thus reducing or even completely abolishing electrical activity. In this sense, $\mathbf{K}_{\mathrm{ATP}}$ channels could act as metabolic gatekeepers of electrical activity, allowing the cells to recover their energy balance by adapting electrical activity of neurones and synapses to their intrinsic energy levels7.

Initial evidence for the distribution of K_{ATP} channels in the brain came from binding studies using labelled sulfonylureas. Not only was there evidence for K_{ATP} channel expression in pyramidal and other cortical neurones, but the highest density of binding sites was found in the basal ganglia, in particular in the GABAcontaining output nuclei: the globus pallidus and the substantia nigra pars reticulata (SNr)^{8,9}. The strategic placement of K_{ATP} channels on excitatory or inhibitory neurones could result in the control of network activity, which under conditions of impaired neuronal energy metabolism drives seizure generation and propagation. The GABA-containing neurones in the SNr form a key structure controlling the propagation of seizures¹⁰ because these neurones receive several inputs from basal ganglia nuclei, including a massive GABAergic projection from striatonigral neurones, and in turn project to thalamic targets¹¹. Pharmacological inhibition or selective lesion of these SNr neurones reduces the spread of epileptic activity in the brain¹⁰. Postsynaptic K_{ATP} channels that interfere with somatodendritic action potential generation, and presynaptic $K_{\!\scriptscriptstyle ATP}$ channels that limit neurotransmitter release could contribute to this network control. Indeed, functional and lesion studies indicated that postsynaptic $K_{\mbox{\tiny ATP}}$ channels on GABA-containing projection neurones, in addition to presynaptic $K_{\mbox{\tiny ATP}}$ channels on GABAergic terminals of

striatal projection neurones, contribute to this 'hot spot' of $\rm K_{ATP}$ channel expression in the SNr (Refs 12–14).

Involvement of \mathbf{K}_{ATP} channels in seizure propagation

In 1990, Amoroso and colleagues showed that KATP channel opening reduced GABA release in the SNr and they were the first to speculate that $K_{\mbox{\tiny ATP}}$ channels in the substantia nigra might be involved in the control of seizure propagation¹⁵. An elegant study by Yamada and colleagues now provides direct experimental evidence for the concept of K_{ATP} -channel-mediated seizure control¹⁶. The research teams of Nobuya Inagaki and Susumu Seino analysed a K_{ATP} channel knockout mouse, where the gene encoding Kir6.2 was inactivated17, to address the key question of whether ${\rm K}_{\rm ATP}$ channels are involved in hypoxia-induced seizures. Their behavioural data, combined with electroencephalogram (EEG) and electromyogram (EMG) recordings, clearly demonstrate that K_{ATP} knockout mice possess a dramatically reduced threshold for hypoxia-induced seizure generation. Additional support for a role of neuronal K_{ATP} channels in the dynamic control of the seizure threshold comes from a recent complementary study by Hernandez-Sanchez and co-workers¹⁸. These authors generated transgenic mice that selectively overexpressed the K_{ATP} channel β-subunit SUR1 in forebrain structures, under the control of a Ca2+-calmodulin kinase promoter, and found that not only was there a significant increase in the threshold of kainateinduced seizures, but also a resistance to excitotoxic neuronal damage in these SUR1-overexpressing animals. These studies examine both ends of a spectrum of K_{ATP} channel expression – complete loss and tenfold overexpression - and provide strong support that the number of active neuronal K_{ATP} channels dynamically controls the seizure threshold in the brain. In this context, it will be relevant to understand regulatory mechanisms such as ischaemic preconditioning that might tune the expression level of neuronal $K_{\mbox{\tiny ATP}}$ channels in vivo and thus determine the level of protection¹⁹.

Yamada and colleagues¹⁶ also present electrophysiological evidence at the cellular level that postsynaptic K_{ATP} channels alone are sufficient to reduce the activity of the GABA-containing SNr neurones in response to hypoxia. In SNr neurones from knockout mice, the reduction of spontaneous discharge in response to hypoxia is not only completely lost but is converted into a hypoxiamediated excitation. This indicates that K_{ATP} channel activation is an important counterbalancing mechanism that offsets increased excitability induced by impaired metabolism. En passant, their data also demonstrate that the pore-forming subunit Kir6.2 is an essential component of K_{ATP} channels in SNr neurones, consistent with previous biophysical and single-cell reverse-transcriptase polymerase chain reaction (RT-PCR) studies^{14,20}. This implies that K_{ATP} channels in the pancreatic β -cell and in metabolically sensitive neurones all share the same molecular makeup of Kir6.2 and SUR1 subunits^{4,6,17,20,21}. Consequently, the selective pharmacological targeting of the SNr K_{ATP} channels as suggested by Yamada and coworkers might be difficult. However, it will be important to investigate whether clinically used K_{ATP} channel openers such as diazoxide, which are used to treat hypertension, or K_{ATP} channel blockers such as sulfonylureas, which are used in the treatment of diabetes mellitus type II, also have effects on neuronal K_{ATP} channels in clinically relevant concentrations and thus on ischaemic tolerance and seizure threshold.

Yamada and colleagues convincingly demonstrate a special role of K_{ATP} channels in the SNr as a crucial seizure gate. However, to provide direct proof, a new generation of transgenic mouse models with specific alteration of K_{ATP} channel expression in SNr neurones might be necessary. This important study has provided a large step towards a better understanding of the physiological role of these metabolic gatekeeper channels in the brain.

References

- 1 Coetzee, W.A. *et al.* (1999) Molecular diversity of K⁺ channels. *Ann. New York Acad. Sci.* 868, 233–285
- 2 Seino, S. (1999) ATP-sensitive potassium channels: a model of heteromultimeric potassium channel/receptor assemblies. *Annu. Rev. Physiol.* 61, 337–362
- $\begin{array}{l} 3 \mbox{ Ashcroft, F.M. and Gribble, F.M. (2000) New} \\ \mbox{windows on the mechanism of action of K_{ATP} channel openers. Trends Pharmacol. Sci. 21, $439–445 \end{array}$
- 4 Aguilar-Bryan, L. *et al.* (2001) Of mice and men: K_{ATP} channels and insulin secretion. *Recent Prog. Horm. Res.* 56, 47–68
- 5 Ashcroft, S.J. (2000) The β -cell K_{ATP} channel. J. Membr. Biol. 176, 187–206

- 6 Miki, T. et al. (2001) ATP-sensitive K+ channels in the hypothalamus are essential for the maintenance of glucose homeostasis. Nat. Neurosci. 4, 507–512
- 7 Liss, B. and Roeper, J. (2001) Molecular physiology of neuronal K-ATP channels. *Mol. Membr. Biol.* 18, 117–127
- 8 Treherne, J.M. and Ashford, M.L. (1991) The regional distribution of sulphonylurea binding sites in rat brain. *Neuroscience* 40, 523–531
- 9 Mourre, C. et al. (1990) Sulfonylurea binding sites associated with ATP-regulated K⁺ channels in the central nervous system: autoradiographic analysis of their distribution and ontogenesis, and of their localization in mutant mice cerebellum. Brain Res. 519, 29–43
- 10 Depaulis, A. et al. (1994) Endogenous control of epilepsy: the nigral inhibitory system. Prog. Neurobiol. 42, 33–52
- 11 Bolam, J.P. et al. (2000) Synaptic organisation of the basal ganglia. J. Anat. 196, 527–542
- 12 Watts, A.E. *et al.* (1995) Putative pre- and postsynaptic ATP-sensitive potassium channels

in the rat substantia nigra *in vitro*. J. Neurosci. 15, 3065–3074

- 13 Stanford, I.M. and Lacey, M.G. (1996) Electrophysiological investigation of adenosine trisphosphate-sensitive potassium channels in the rat substantia nigra pars reticulata. *Neuroscience* 74, 499–509
- Schwanstecher, C. and Panten, U. (1993) Tolbutamide- and diazoxide-sensitive K⁺ channel in neurons of substantia nigra pars reticulata. *Naunyn-Schmiedeberg's Arch. Pharmacol.* 348, 113–117
- 15 Amoroso, S. et al. (1990) Glucose, sulfonylureas, and neurotransmitter release: role of ATP-sensitive K⁺ channels. Science 247, 852–854
- 16 Yamada, K. et al. (2001) Protective role of ATPsensitive potassium channels in hypoxia-induced generalized seizure. Science 292, 1543–1546
- $17 \ \ {\rm Seino, S.} \ et al. \ (2000) \ {\rm Diverse \ roles \ of \ } K_{\rm ATP} \\ {\rm channels \ learned \ from \ Kir6.2 \ genetically} \\ {\rm engineered \ mice.} \ Diabetes \ 49, \ 311-318 \\ \end{array}$
- 18 Hernandez-Sanchez, C. *et al.* (2001) Mice transgenically overexpressing sulfonylurea receptor 1 in forebrain resist seizure induction

and excitotoxic neuron death. Proc. Natl. Acad. Sci. U. S. A. 98, 3549–3554

- 20 Liss, B. et al. (1999) Alternative sulfonylurea receptor expression defines metabolic sensitivity of K-ATP channels in dopaminergic midbrain neurons. EMBO J. 18, 833–846
- 21 Seghers, V. et al. (2000) Sur1 knockout mice. A model for K_{ATP} channel-independent regulation of insulin secretion. J. Biol. Chem. 275, 9270–9277

Birgit Liss

e-mail: birgit.liss@physiol.ox.ac.uk Jochen Roeper

MRC Anatomical Neuropharmacology Unit, University of Oxford, Mansfield Road, Oxford, UK OX1 3TH.

e-mail: jochen.roeper@pharm.ox.ac.uk

A role for neuronal K_{ATP} channels in metabolic control of the seizure gate

Response from Yamada and Inagaki

As noted by Birgit Liss and Jochen Roeper, the subunit combinations of the ATP-sensitive $K^+(K_{ATP})$ channels are important in the analysis of the function of these channels in the CNS. Indeed, Roeper's group has shown by a combination of patch-clamp and single-cell reverse-transcriptase polymerase chain reaction (RT-PCR) that there are three populations of dopamine-containing substantia nigra pars compacta (SNc) neurons with different K_{ATP} channel subunit expression profiles and distinct metabolic and drug sensitivities, and that GABA-containing SNc interneurons do not express any of the K_{ATP} channel subunits¹. This clearly indicates that pharmacological studies should carefully evaluate the functional roles of K_{ATP} channels in brain nuclei comprising heterogeneous neuronal populations that express different subunit combinations. In contrast to the SNc, the substantia nigra pars reticulata (SNr) expresses a homogenous K_{ATP} channel subunit profile with SUR1 and Kir6.2 coexpression in all GABA-containing neurons¹.

The SNr, the major output nucleus of the basal ganglia, is considered to be a key structure that controls the propagation of generalized seizures². A striking feature of the SNr is that almost all of the neurons are spontaneously very active. Their firing rate of up to 100 spikes per second is among the highest in the CNS and is a crucial determinant of basal ganglia functions³.

Given that spike generation, maintenance of membrane potential, and neurotransmitter synthesis all depend on glucose and oxygen, insufficiency of either glucose or oxygen might lead to an earlier decrease in the cytosolic ATP concentration in this nucleus than in other 'relatively silent' brain areas such as the cerebral cortex and hippocampus. Our data using mice that lack the Kir6.2 subunit of the K_{ATP} channel seemed to support this hypothesis, and we proposed that neurons in the SNr act as a sensor for hypoxia⁴.

 $\rm K_{ATP}$ channels are expressed in varying abundance in the brain. For example, a recent study found that $\rm K_{ATP}$ channels comprising Kir6.2 and SUR1 were expressed in 58% of hippocampal CA1 interneurons⁵. These channels are also activated by energy depletion⁶. What is the specific role of $\rm K_{ATP}$ channels in the SNr? SNr neurons innervate diverse motorrelated target nuclei at long distances, including not only the ventral thalamic

nuclei but also the midbrain superior colliculus and the pedunculopontine nucleus in the brain stem⁷. These nuclei are involved in a broad spectrum of motor control⁸. Thus, abrupt silence in the SNr GABA-containing projection neurons in synchrony during the early phase of brain metabolic deficiency might well convey a signal to all these target nuclei as massive disinhibition. Further study is required to clarify the mechanisms of altered function at the target regions in the control of seizure during hypoxia².

There is a larger increase in the cerebral metabolic rate of oxygen $(CMRO_2)$ during seizure than under any other circumstance⁹. This rapid increase in energy demand associated with the onset of seizure produces a rapid fall in brain energy metabolites. Hence, nigral suppression of seizure propagation might protect from brain damage by reducing the energy consumption rate in the whole brain.

Quantification of the dynamic changes in intracellular ATP concentrations in different cells and regions during normal and pathophysiological conditions is necessary to clarify the various activities of the K_{ATP} channels sufficiently to contemplate