SK-Ca2 Small Conductance Calcium Activated Potassium Channel

Stephan Grissmer Universitat Ulm, Ulm, Germany

© 2007 Elsevier Inc. All rights reserved.

Introduction

The SK2 small conductance calcium-activated potassium channel belongs to the family of potassium channels that consist of one pore region (1P) with 6 putative transmembrane segments (6T) per alpha subunit (1P6T). It is a potassium selective ion channel that is opened by an increase in [Ca⁺⁺]_i. The opening of the channel is independent from the applied voltage, and the single channel conductance is of small size compared to calcium-activated potassium channels with intermediate and large single channel conductances Hille (2001). This channel type is usually thought to underlie the slow after hyperpolarization seen in neuronal cells. In addition to the different single channel conductance, these channels do also have a specific pharmacology, i.e., they are blocked by apamin, a peptide toxin isolated from bee venom, as well as blocked by Scyllatoxin (Leiurustoxin I), a peptide toxin isolated from scorpion venom. In addition, the channels are also blocked selectively by several bis-quinolinium cyclophanes (UCL 1530, UCL 1684, UCL 1848, UCL 2079). The channels can be activated by 1-EBIO (1-ethyl-benzimidazolinone), similar to the calcium-activated potassium channels with intermediate conductance.

Nomenclature

Superfamily 1P6T potassium channels

Family Voltage independent Ca⁺⁺-activated potassium

channels

Type SK2

Subtypes

Classification Numbers KCNN2
Alternate or Previous SK2, SKCa2

Names Comments

Target Structure

Protein Information

SK2 is a pore forming subunit. The functional channel consists of four identical subunits (homotetramer), each with one pore region (1P) and 6 putative transmembrane segments (6T). The Ca⁺⁺ sensor seems to be calmodulin bound to each subunit at a region between the S6 segment and the C-terminal end of the channel.

Protein Sequence Information

	Number or Name	Comments
Subunit Name	hKCNN2	
Organism Name	Human	
Gene Accession #	NM_021614	AF239613 Desai et al (2000), AF397175, XM_011226
SwissProt Accession #	Q9H2S1	
# of Amino Acid Residues	579	
Protein Sequence Motifs		hKCNN2 contains 3 consensus sites for N-glycosylation site, 3 consensus sites for cAMP- and cGMP-dependent protein kinase phosphorylation, 12 consensus sites for Protein kinase C phosphorylation, 7 consensus sites for Casein kinase II phosphorylation, 20 consensus sites for N-myristoylation site, 1 glycine-rich and 1 serine-rich region.
Chromosomal Localization	5q23.1-23.2	Ganshani et al (2000) NT_006899 chromosome 5 reference genomic contig.

	Number or Name	Comments
Subunit Name	rKCNN2	Kohler et al (1996)
Organism Name	Rat	Kohler et al (1996)
Gene Accession #	NM_019314	
SwissProt Accession #	P70604	
# of Amino Acid Residues	581	
Protein Sequence Motifs		rKCNN2 contains 3 consensus sites for N-glycosylation, 3 consensus sites for cyclic AMP- and cyclic GMP-dependent protein kinase phosphorylation, 13 consensus sites for Protein kinase C phosphorylation site, 8 consensus sites for casein kinase II phosphorylation, 24 consensus sites for N-myristoylation site, 1 glycine-rich and 1 serine-rich region.

	Number or Name	Comments
Subunit Name	mKCNN2	
Organism Name	Mouse	
Gene Accession #	NM_080465	AF357240, AY123778
SwissProt Accession #	P58390	
# of Amino Acid Residues	575	
Protein Sequence Motifs		mKCNN2 contains 3 consensus sites for N-glycosylation, 3 consensus sites for cyclic AMP- and cyvlic GMP-dependent protein kinase phosphorylation, 13 consesnus sites for Protein kinase C phosphorylation, 8 consensus sites for Casein kinase II phosphorylation, 18 consensus sites for N-myristoylation, 1 glycine-rich and 1 serine-rich region.
Chromosomal Localization	18	NW_000134 mus musculus WGS supercontig Mm18_WIFeb01_311

	Number or Name	Comments
Subunit Name Organism Name Gene Accession # SwissProt Accession # # of Amino Acid Residues Protein Sequence Motifs	cKCNN2 Gallus gallus (chicken) AF079372 554	cKCNN2 contains 3 consensus sites for N-glycosylation site, 3 consensus sites for cAMP- and cGMP-dependent protein kinase phosphorylation, 10 consensus sites for Protein kinase C phosphorylation, 5 consensus sites for Casein kinase II phosphorylation, and 5 consensus
Chromosomal Localization		sites for N-myristoylation.

Localization

Protein

Neurons Hille (2001); Jurkat T lymphocytes Grissmer et al (1992), Jager et al (2000).

mRNA

Hippocampus (CA3), dentate gyrus, subiculum, anterior olfactory nucleus, olfactory tubercle, cerebellum, and cortex Stocker et al (2000).

Ligands, Substrates, Ions

Ligands

Ca⁺⁺, calmodulin

Substrates

Name	Km value	Km units	Reference	Remarks
Ca++ Calmodulin	300	nM	Kohler et al (1996), Grissmer et al (1992) Xia et al (1998), Schuhmacher et al (2001)	

lons

	Value	Units	Reference	Remarks
Potassium				
Conductance	4-20	pS	Hille (2001)	The higher conductance is measured with elevated external potassium concentrations.
Voltage				Dependence
none				

Effectors, Products

Establishing a link between Ca⁺⁺-based second messenger systems and the electrical activity of cells.

Endogenous Regulation

Protein Partners

calmodulin

Pharmacological Regulation

Selective peptide blockers are apamin and scyllatoxin Hille (2001). Highly selective non-peptide blockers are different bis-quinolinium cyclophanes Stroebaek et al (2000), Shah et al (2000). Non-selective, more unspecific blockers are d-tubocurarine, verapamil, diltiazem, and tetraethylammonium. Openers are 1-ethyl-benzimidazolinone (1-EBIO, (Pedarzani et al., 2001)); EBIO activates also IK channels Devor et al (1996) but not BK channels.

Agonist / Activator / Substrate

	Value	e Units	s Organism	Organ Tissue	Cell Line/ Type	Reference	Comments
Age	nt: 1-e	thyl-be	nzimidazolino	one (1-EBIC	O)		
Ki:	650	μM	Rat KCNN2		Xenopus oocytes	Pedarzani et al (2001)	1-EBIO activates directly the channel and requires the presence of intracellular Ca ⁺⁺ method: electrophysiology (whole cell).

Antagonist / Inhibitor

	Value	Units	Organism	Organ Tissue	Cell Line/ Type	Reference	Comments			
Agent:	Agent: UCL1684									
IC50:	0.4	nM	Rat KCNN2		HEK- 293	Stroebaek et al (2000)	Method used: electrophysiology (whole cell).			

Antagonist / Inhibitor

	Value	Units	Organism	Organ Tissue	Cell Line/ Type	Reference	Comments
Agent:	: Apam 0.3	in nM	Human	T-cell carcinoma; endogeneously expressed	Jurkat	Grissmer et al (1992)	Method used: electrophysiology (whole cell).

Antagonist / Inhibitor

	Value	Units	Organism	Organ Tissue	Cell Line/ Type	Reference	Comments
Agent	: Apamin	ı					
IC50:	0.1	nM	Rat KCNN2		HEK-293	Stroebaek et al (2000)	Method used: electrophysiology (whole cell).

Antagonist / Inhibitor

	Value	Units	Organism	Organ Tissue	Cell Line/ Type	Reference	Comments
Agent:	: Leiurut	oxin / sc	yllatoxin				
IC50:	0.3	nM	Human	T-cell carninoma; endogeneously expressed	Jurkat	Hanselmann and Grissmer (1996)	Method used: electro- physiology (whole cell).

Antagonist / Inhibitor

	Value	Units	Organism	Organ Tissue	Cell Line/ Type	Reference	Comments
Agent:	Leiurut	oxin/s	cyllatoxin				
IC50:	0.3	nM	Rat KCNN2		HEK-293	Stroebaek et al (2000)	Method used: electrophysiology (whole cell).

Antagonist / Inhibitor

Value	Units	Organism	Organ Tissue	Cell Line/Type	Reference	Comments
Agent: Lei-Da IC50: 4	ab7 nM	Rat KCNN2		COS-7	Shakkotai et al (2001)	

Antagonist / Inhibitor

Value	Units	Organism	Organ Tissue	Cell Line/ Type	Reference	Comments
Agent: Dequali IC50: 160	nium nM	Rat KCNN2		HEK-293	Stroebaek et al (2000	Method used: electrophysiology (whole cell).

Antagonist / Inhibitor

Value U	Jnits	Organism	Organ Tissue	Cell Line/ Type	Reference	Comments
Agent: Tubocu	ırarine	•	•			
IC50: 2 μ	ιM	Rat KCNN2	2	Xenopus oocytes; HEK-293	Kohler et al (1996), Jager et al (1997), Stroebaek et al (2000)	,

Antagonist / Inhibitor

Value	Units	Organism	Organ Tissue	Cell Line/ Type	Reference	Comments
Agent: Bicuculli IC50: 1	ne μM	Rat KCNN2		Xenopus oocytes	Khawaled et al (1999)	Method used: electrophysiology (whole cell).

Journal Citations

- Desai, R., Peretz, A., Idelson, H., Lazarovici, P., Attali, B., 2000. Ca²⁺ activated K⁺ channels in human leukemic Jurkat T cells. Molecular cloning, biochemical and functional characterization. *J. Biol. Chem.*, 275, 39954–39963.
- Devor, D.C., Singh, A.K., Frizell, R.A., Bridges, R.J., 1996. Modulation of Cl⁻ secretion by benzimidazolones. I. Direct activation of a Ca(2+)-dependent K+ channel. *Am. J. Physiol.*, 271, 775–L784.
- Ganshani, S., Wulff, H., Miller, M.J., Rohm, H., Neben, A., Gutman, G.A., Cahalan, M.D., Chandy, K.G., 2000. Up-regulation of the IKCa1 potassium channel during T-cell activation. *J. Biol. Chem.*, 275, 37137–37149.
- Grissmer, S., Lewis, R.S., Cahalan, M.D., 1992. Ca(2+) activated K+ channels in human leukemic T cells. *J. Gen. Physiol.*, 99, 63–84.
- Hanselmann, C., Grissmer, S. et al., 1996. Characterization of apamin-sensitive Ca(2+)-activated potassium channels in human leukemic T lymphocytes. *J. Physiol. (Lond)*, 496, 627–637.
- Jager, H., Grissmer, S., et al. 1997. Small Ca2+-activated potassium channels in the human leukemic T cells and activated human peripheral blood lymphocytes. Cell Physiol. Biochem., 7, 179–187.
- Jager, H., Adelman, J.P., Grissmer, S., 2000. SK2 encodes the apamin-sensitive Ca2+-activated K+ channels in the human leukemic T cell line, Jurkat. *FEBS Lett.*, 469, 196–202.
- Khawaled, R., Brueniung-Wright, A., Adelman, J.P., Maylie, J., 1999. Bicuculline block of small-conductance calcium-activated potassium channels. *Pflugers Arch.*, 438, 314–321.

- Kohler, M., Hirschberg, B., Bond, C.T., Kinzie, J.M., Marrion, N.V., Maylie, J., Adelman, J.P., 1996. Small-conductance, calcium-activated potassium channels from mammalian brain. *Science*, 273, 1709–1714.
- Pedarzani, P., Mosbacher, J., Rivard, A., Cingolani, L.A., Oliver, D., Stocker, M., Adelman, J.P., Fakler, B., 2001. Control of electrical activity in central neurons by modulating the gating of small conductance Ca2 +-activated K+ channels. *J. Biol. Chem.*, 276(13), 9762–9769.
- Schuhmacher, M.A., Rivard, A., Bachinger, H.P., Adelman, J.P., 2001. Mechanism of calcium gating in small-conductance Ca2+-activated K+ channel complexed with Ca2+/calmodulin. *Nature*, 410, 1120–1124.
- Shah, M., Haylett, D.G. et al., 2000. The pharmacology of hSK1 Ca2+-activated K+ channels expressed in mammalian cell lines. *Br. J. Pharmacol.*, 129, 627–630.
- Shakkotai, V.G., Regaya, I., Wulff, H., Fajloun, Z., Tomita, H., Fathallah, M., Cahalan, M.D., Gargus, J.J., Sabatier, J.M., Chandy, K.G., 2001. Design and characterization of a highly selective peptide inhibitor of the small conductance calcium-activated K+ channel, SkCa2. *J. Biol. Chem.*, 276, 43145–43151.
- Stocker, M., Pedarzani, P. et al., 2000. Differential distribution of three Ca2+-activated K+ channel subunits, SK1, SK2, and SK3, in the adult rat central nerveous system. *Mol. Cell Neurosci.*, 15, 476–493.
- Stroebaek, D., Jorgensen, T.D., Christophersen, P., Ahring, P.K., Olesen, S.P., 2000. Pharmacological characterization of small-conductance Ca(2+)-activated K(+) channels stably expressed in HEK 293 cells. *Br. J. Pharmacol.*, 129, 991–999.
- Xia, X.M., Fakler, B., Rivard, A., Wayman, G., Johnson-Pais, T., Keen, J.E., Ishii, T., Hirschberg, B., Bond, C.T., Lutsenko, S., Maylie, J., Adelman, J.P., 1998. Mechanism of calcium gating in small conductance calcium-activated potassium channels. *Nature*, 395, 503–507.

Book Citations

Hille, B., 2001. Ion channels of excitable membranes, Edition 3. Sinauer Associates, Sunderland, MA.