Ingestion of current environmental levels of microplastics is associated with changes in the gut microbiome of two wild seabird species

Microplastic pollution is an increasing problem that society faces today. The lack of action to combat these pollutants likely stems from an absence of evidence that they do harm at environmentally-relevant concentrations and mixtures numerous relevant species, and potentially accumulate across the trophic train. We provided evidence that microplastic ingestion changes gut microbial communities throughout the gastrointestinal tract in two different wild seabird species feeding on marine mollusks, crustacea, and fish across both hemispheres, and therefore represent relevant marine plastic bioindicators. Moreover, we looked at not only one microbial community within the gastrointestinal tract, but at several such communities located anteriorly (in the proventriculus) and posteriorly (in the cloaca) and found that all these communities are changed by the ingestion of microplastics. Microbiomes showed signs of dysbiosis, with decreases in commensal microbes and increases in pathogens (even zoonotic pathogens), antibiotic resistant bacteria, and plastic-degrading microbes.

Our interdisciplinary approach differs from previous laboratory studies employing high concentrations of microplastics which often are not representative of the concentrations observed in the natural world. In our study, however, we show that changes to the microbiome are already occurring at current microplastic concentrations in the environment. The two seabird species we focused on (Northern fulmars and Cory's shearwaters) are highly relevant because they already chronically ingest microplastics and are not limited to small foraging areas, but instead migrate for thousands of kilometers across both hemispheres, putting the scale of these findings into global proportions.


TB epidemiology: The effects of immune gene diversity, gut microbiota on disease susceptibility in meerkats

Infections with tuberculosis (TB)-causing agents of the Mycobacterium tuberculosis complex threaten human, livestock and wildlife health globally due to the high capacity to cross trans-species boundaries. Tuberculosis is a cryptic disease characterized by prolonged, sometimes lifelong subclinical infections, which complicates disease monitoring. Using wild meerkats (Suricata suricatta) inhabiting the Kalahari Desert in South Africa as a model, we aim at disentangling the complex interplay between host immune genetics, microbiota composition, climatic change and TB. We utilize an extensive dataset compiled over 20 years by the Kalahari Research Project, providing detailed information on climate change, individual life history and behaviour, and apply high throughput sequencing techniques to identify shifts in meerkats’ microbiota, Major Histocompatibility Complex (MHC) and olfactory receptor genes (TAARs) constitution to link them to individual life histories and TB outcomes.

First, we developed a PCR-based screening tool to detect Mycobacteria non-invasively in faeces and could demonstrate that infections often occur more than one year prior to the onset of clinical signs, indicating prolonged subclinical infections (Donadio et al. 2022). Quantification of TB prevalence and progression allowed us to discover an increase in TB exposure and prevalence since the early 2000s. Based on life history data of over 3000 individuals, we detected that exposure to TB is the norm rather than the exception in the population, but only a quarter of exposed individuals progress to clinical stages. While individuals can survive exposure and clinical disease for years, eventually most individuals progress to clinical TB stages within 1.5 years of exposure and die within 6 months of developing the first clinical signs. Our results suggest that TB prevalence and mortality is higher than previously reported. These findings shed light on TB epidemiological dynamics in species with complex social system and is relevant for conservation management of animal populations with TB (Müller-Klein et al. 2022).

With respect to the gut microbiome, we first investigated the main drivers of natural variation in meerkat gut microbial diversity. It turned out that meerkat gut microbiomes are highly variable. Variation throughout the day is particularly strong when compared to variation across an meerkats’ lifetimes. This implies circadian rhythms to play an acute role on faecal microbiome composition (Risely et al. 2021). Accordingly, microbiome repeatability within individuals is high over short scales, but identity does not explain microbiomes over longer timescales, with seasonal and circadian effects contributing the most (Risely et al. 2022). Moreover, linking information on TB and the microbiome, we could demonstrate that both exposure and clinical disease are linked to shifts in the microbiome. These are probably not adaptive changes, since they do not increase individual survival, but are consequences of infection related shifts and/ or individual stress. Similar shifts are observed with increasing changes in the climate, with droughts having the most serious effect (Risely et al. submitted).

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Harnessing the microbiome for turtle conservation: investigating the protective effects of the eggshell microbiome against the emerging fungal disease fusariosis in an Amazonian freshwater turtle

The emergence of fungal pathogens threatening wildlife has increased exponentially, with well-documented catastrophic declines in amphibians and bats due to emerging fungal pathogens. However, high mortality in sea turtles due to the emerging fungal disease Fusariosis has received less attention. Fusariosis is caused by fungal pathogens within the Fusarium solani species complex (FSSC), representing important human, plant and animal pathogens, which has led to mass mortality in all seven sea turtle species by infecting turtle eggs during incubation, resulting in hatching failure. Alarmingly, we recently detected this pathogen in the Yellow-spotted Amazonian river turtle (Podocnemis unifilis) in the remote Ecuadorian Amazon (Carranco et al., 2022a), suggesting that this pathogen may also pose a threat to the 356 species of freshwater turtles and tortoises. Despite the imminent threat of this emerging pathogen towards turtle biodiversity, we have very little understanding of this fungal disease, including its transmission dynamics, pathogenicity, and long-term effects on the fitness of host turtles.

Our first aim has been to address this gap by investigating Fusarium disease dynamics using the Yellow-spotted Amazonian river turtle as a model system. We investigated fusariosis in unhatched eggs showing symptoms of Fusarium infection by using Fusarium-specific primers and by applying high throughput sequencing to detect FSSC pathogens. We could confirm the presence of three main FSSC pathogens causing egg fusariosis in turtle species. Moreover, we observed that a high proportion of eggs showing symptoms of fusariosis infection among nests was significantly associated with egg hatching success. However, since not all the eggs showing symptoms of fusariosis tested positive using a PCR FSSC test, we suggest the possibility of multiple causes of bacterial/fungal infection in turtle eggs (Carranco et al., 2022a).

Since the host-associated microbiome is one of the key host traits for conferring resistance against pathogens, our next aim was to unveil the eggshell microbiome and microbial assemblages during turtle development. We investigated the internal eggshell microbiome and the cloaca microbiome of hatchlings and juveniles during the first month of development, together with the surrounding nest and river environment, using specific primers to amplify the V4 region of bacterial 16S rRNA on an Illumina platform. We first observed that the turtle eggs harbour an internal microbiome which is the seed for hatchlings’ gut microbiome. We demonstrated that the nest environment has an influence on the egg and hatchling cloaca microbiome, which is the base of development for the cloaca microbiome of juveniles. During the first 20 days of juvenile turtle development, the gut microbiome changes and matures by the force of host selection processes and the interaction with the river environment. With these findings we could quantify for the first time the relative role of the environment and host-selection on gut microbial assemblages of oviparous reptiles, and expanded our understanding of Fusarium as an emerging infectious disease and the role of host-microbiome interactions in turtles (Carranco et al., 2022b). Our next project will allow us to investigate Fusarium epidemiology and the role of the host-associated micro and mycobiome in conferring resistance and resilience to fusariosis infection in turtle eggs in detail.


Predicting the impact of co-infections on host gut microbiomes

Co-infections are the norm in nature. Almost 50% of bank voles, for example, carry the tick-borne bacterium ‘Candidatus Neoehrlichia mikurensis’ and the disease agent Borrelia afyelii, associated with Lyme disease in humans, and as many as 40% of bats are infected with more than one virus. Yet, we know very little about how co-infections, particularly with pathogens replicating enterically, impact the abundance and diversity of commensal gut microbiota. Since commensal bacteria take up important roles in hosts, including those related to nutrient breakdown and host immune defences, a better understanding of how pathogens shift their fragile balance was needed. In this work, we summarised findings that suggest that co-infecting pathogens can accentuate, undermine and disregard changes to the microbiome observed from single infections. We constructed a predictive framework that outlines how to test for precisely those synergistic, antagonistic or neutral effects of co-infecting pathogens and showcase our approach by re-analysing published microbiome data from a study on Malagasy mouse lemurs co-infected with an Adenovirus and helminths (Montero et al. 2021). It seems that co-infections may become more common in human encroached environments. In turn, this could raise the possibility of sicker wildlife populations owing to the disturbance of the gut microbiota and their functions, and risk the emergence and persistence of novel disease agents that jump the species barrier to infect domesticated animals or humans.


Disentangling effects of habitat disturbance on biodiversity

Changes in habitat characteristics following human disturbances are one of the most important factors influencing ecological communities and species abundance patterns. Yet, comparably little attention has been paid to the interconnectedness of biological communities at various levels of biodiversity, and whether their response differs. This study documents the nature and strength of both direct and indirect, and, at times, parallel effects of habitat disturbance on biodiversity: habitat characteristics associated with a gradient of human disturbance were the strongest predictors for small mammal species richness, and the genetic diversity and microbial heterogeneity of a dominant generalist. However, we also found clear evidence for indirect effects of human disturbance on host-associated species assemblages via small mammal species richness and host genetic diversity. Changes to host genetic diversity and the host species assemblage in connection with a reshuffling of the host associated micro- and macroorganisms may also explain why anthropogenically altered habitats become sources of diseases. Taken together, our study illustrates how human disturbance impacts multiple components of biodiversity in parallel, and causes cascading effects among them.

Interaction between MHC diversity and constitution, gut microbiota and Astrovirus infections in a neotropical bat

Astroviruses (AstVs) infect numerous mammalian species including reservoirs such as bats. Peptides encoded by the genes of the highly polymorphic Major Histocompatibility Complex (MHC) form the first line of host defence against pathogens. Aside from direct involvement in mounting adaptive immune responses, MHC class II genes are hypothesized to regulate gut commensal diversity and shape the production of immune-modulatory substances by microbes, indirectly affecting host susceptibility. Despite initial empirical evidence for the link between host MHC and the microbiota, associations among these factors remain largely unknown. To fill this gap, we examined MHC allelic/supertype diversity and constitution, the gut bacterial community and abundance pattern of a wild population of the neotropical bat (Artibeus jamaicensis) challenged by AstV infections. First, we show an age-dependent relationship between the host MHC class II diversity and constitution and the gut microbiota in AstV uninfected bats. Crucially, these associations changed in AstV infected bats. Additionally, we identify changes in abundance of specific bacterial taxa linked to the presence of certain MHC supertypes and AstV infection. We suggest changes in the microbiota to be either a result of AstV infection or the MHC-mediated modulation of microbial communities. The latter could subsequently affect microbe-mediated immunity and resistance against AstV infection. Our results underscore that the reciprocal nature of host immune genetics, gut microbial diversity and pathogen infection requires attention, which is particularly important given its repercussions for disease susceptibility and severity in wild animal populations with a history of zoonotic spillover and frequent human contact.


Effects of human encroachment into wildlife microbiomes

The gut microbiome is a symbiont consisting of a vast community of microbes and their genes, which play a key role in host health. Disturbances of this gut microbial community has been linked to many autoimmune diseases and infections, highlighting just how important a healthy gut microbiome is for the health of its host. Changes to wildlife gut microbiomes due to anthropogenic disturbances, such as habitat fragmentation, can disrupt natural gut microbiota homeostasis and make animals vulnerable to infections that may become zoonotic. In this study, we used the gut microbiome as an indicator for health in order to understand how zoonotic diseases can be transferred from wildlife to humans in the Anthropocene. Although anthropogenic disturbances are known to negatively impact the gut microbiome in a process known as dysbiosis, it is still unclear which anthropogenic factors drive these changes. Our study design in the Panama Canal area allows for the differentiation between habitat fragmentation per se and living in close proximity to humans, their domesticated animals, and agriculture. We found that 384 spiny rats (Proechimys semispinosus) living in fragmented habitats close to humans had greatly altered gut microbiomes, whereas spiny rats living in protected, fragmented habitats did not. This means habitat fragmentation per se did not drive the change in microbiome. Instead, the additional contact with humans shaped gut microbial composition and diversity. These findings are significant because spiny rats are generalist species expected to be more resilient to environmental changes. Yet, their microbiomes displayed two patterns of change: The structural composition of the microbes that make up the gut microbiome became more irregular (heterogeneous) and shifted away from its natural composition. They also harbored more bacterial species that are found in domesticated animals and are known to cause disease in livestock. Our work suggests that gut microbiomes of spiny rats react to changes in their environment and offer the host a pathway to adapt or cope with anthropogenic alterations. Whether microbiome-mediated adaptations are fast and substantial enough to keep up with the rate and degree of change in the Anthropocene remains to be seen. Understanding that wild animals are struggling to cope with changes in the Anthropocene should serve as a warning that we may see more zoonosis-driven pandemics in the future.

Meerkat gut microbiomes exhibit strong daily oscillations that do not decay with age

Gut bacterial communities are often incredibly dynamic over time, which makes them challenging to study. Microbes can fluctuate over the scale of hours, weeks, months and years, in response to feeding, seasonal climate, and host ageing, yet so far, we have not been able to disentangle these processes. We investigated gut microbiome dynamics in wild meerkats (*Suricata suricatta*) over a 20-year period to compare diurnal, seasonal, and lifetime processes in concert. We found that almost all common genera exhibited diurnal oscillations, and daily dynamics in bacterial load and diversity were stronger than seasonal and lifetime effects. Diurnal oscillations were driven by changes to the genus *Clostridium*, were better explained by light-dark cycles than foraging schedule, and did not decay with age. Across life, specific genera were associated with ageing and condition, and were more tightly linked to biological rather than chronological senescence. Our findings highlight that daily oscillations in the microbiome are probably more common than previously thought, and demonstrate that accounting for circadian rhythms is crucial for future gut microbiome research.


Foraging in monocultures with heavy pesticide input decreases the gut microbiota diversity of nectar-feeding bats

The health and physiology of native species can be negatively affected by agriculture, both due to habitat alteration and to pesticide use, and it is possible that gut microbiome disruption plays a role in these negative effects. We examined the association between management intensity of banana plantations and both the gut microbiota composition and body condition of one species of nectar-feeding bat (*Glossophaga soricina*, Phyllostomidae) in the Costa Rican Caribbean lowlands. We discovered that gut microbiota from bats foraging in conventional monocultures were overall less phylogenetically diverse than those from bats foraging in organic plantations or natural forests, both of which were characterized by diverse bacterial assemblages and individualized microbiota. Nonetheless, co-occurrence network complexity was higher in conventional monocultures, possibly indicating altered microbial interactions in agricultural landscapes. Bats from both banana plantations were larger and heavier than their forest counterparts, reflecting the higher and year-long food supply. With our study, we discovered that even if both banana plantations (conventional and organic) do provide a reliable food source for bats, conventional monocultures are associated with less diverse and potentially dysbiotic microbiota, while organic plantations promote diverse and individualized gut microbiota akin to their natural forest foraging counterparts. Whilst the long-term negative effects of anthropogenically-altered microbiota are unclear, our study provides evidence that organic agricultural practices are indeed more sustainable for wildlife health.

Impact of habitat disturbance and fungicides on homeostasis of amphibian skin microbiomes and abundance of protective bacteria against chydrid fungus infections - using endangered and rediscovered Costa Rican amphibians as natural model organisms

Some amphibians that were thought to be “extinct” have been found decades after population declines in the late 1980s. These species appear to have evolved resistance to the pathogen *Batrachochytrium dendrobatidis* (Bd), the likely causative agent of their declines, and raises the question how selection and diversity of protective skin microbiomes enabled these endangered neotropical amphibians to survive. Our study showed that the skin microbiome of surviving species (*Lithobates vibicarius, Craugastor escoces, Isthmohyla rivularis* and *I. pseudopuma*) possess putatively Bd-inhibitory activity across species (e.g., by the presence of *Pseudomonas veronii* and *Acinetobacter johnsonii*), which could be involved in resistance against Bd. We also studied the variation of the skin microbiome across the life stages of *L. vibicarius* and observed that life stage is a strong predictor of the diversity of the skin microbiome, suggesting a dynamic skin microbiome through development. Unfortunately, these amphibian populations are still not “safe” due to ongoing anthropogenic threats, caused by habitat disturbance and overuse of pesticides. Alteration of the microbiome associated with environmental changes produced by anthropogenic activities may make the host more susceptible to pathogens. Our research showed that bacterial diversity of tadpoles of *L. vibicarius* from the disturbed habitats was lower than in those from the undisturbed habitats. Adults of *L. vibicarius* from disturbed habitats exhibited greater community dispersion than those from undisturbed habitats. These observed patterns could be associated to the presence of environmental stressors in our study sites, which can perturb a stable state leaving animals more susceptible to pathogen infections even beyond Bd. We could demonstrate that exposure to a widely used fungicide, chlorothalonil, changes the skin bacterial communities of tadpoles of *L. vibicarius*, potentially disrupting this protective trait against pathogens. With this research, we gained important information of the functional importance of the skin microbiomes, and the negative impacts of anthropogenic activities, especially agrochemicals, which can be used to develop management strategies that protect the health of these endangered amphibian populations.


Anthropogenic disturbance, Adenovirus and Astrovirus infections perturb the gut microbial community of non-human primates and bats

Non-human primates represent an excellent natural model system for studying host-pathogen-microbiome interactions, since they harbour an important source of several crucial viruses of humans and livestock, including Adenoviruses AdV - a major cause of diarrhoea in children. As in humans, a remarkably high AdV-prevalence, and inferred zoonotic potential has been reported for mouse lemur. We examined the influence of anthropogenic disturbance and AdV on the gut microbiome of naturally infected Malagasy mouse lemur (*Microcebus griseorufus*). Our study shows that habitat fragmentation and enteric AdV indeed disturbed the gut microbiome community composition. Beneficial genera decreased, whereas pathogen-containing genera increased after AdV-infection leading to pathobiont-like shifts in the gut microbiome. This shows that in apparently healthy lemur, AdV infections disturb the gut bacterial homeostasis, which can increase previously suppressed health risks by promoting co-infections.
Interestingly, we observed similar effects also in a neotropical bat species (*Artibeus jamaicensis*) infected with Astroviruses, also causing diarrhoea in young and immune-compromised humans, supporting the idea that virus-microbiome interactions have important, largely neglected implications for host health. If similar processes are present in humans, the effects of enterovirus infections would cause longer-term impacts and go beyond the directly observed symptoms.


Microplastics and the gut microbiome: how chronically exposed species may suffer from gut dysbiosis

Since the mass production of plastics began in the 1950s, we have become inundated in plastic. Although this development brought with it advantages, it also started a massive and uncontrolled experiment since it takes centuries for plastics to degrade. In this review, we identified three mechanisms through which microplastics could harm gut microbial symbionts. Given the gut microbiomes’ vital standing in host health, microplastics could harm wildlife health in several ways: 1) by mechanically damaging the gut lining, an essential component in maintaining homeostasis between host and the gut microbiome; 2) by acting as a pathogen vector, allowing potential pathogens to hitchhike on the surface of microplastics and thus disturbing the gut microbial community; and 3) by leaching chemicals that can interfere with normal host hormone signaling, another major regulator of the gut microbiome.


Gut microbiome as indicator of heavy metal pollution in soil

Gut microbiomes are becoming recognized as important players in organism health, with comprehension of their perturbations in the polluted environment offering new insights into the nature and extent of heavy metal effects on the health of soil biota. Earthworms are essential species in soil, which carry out the soil biological regulation by mixing organic matter and mineral particles inside their gut. Through their intimate contact with soil, earthworms are exposed to the variety of pollutants and are important indicators of soil quality. In this research we investigated the effect of heavy metal cadmium (Cd) on the earthworm (*Lumbricus terrestris*) gut microbiota. Cd exposure led to perturbations of earthworm gut microbiota with an increase in bacteria previously described as heavy metal resistant or able to bind heavy metals, revealing the potential of the earthworm-gut microbiota system in overcoming heavy metal pollution. Furthermore, we defined several bacterial genera as bacterial indicators of exposure to Cd which could serve as biomarkers in soil biomonitoring practices. The results of this study help to understand the impact of anthropogenic disturbance on soil fauna health and will have implications for environmental monitoring and protection of soil resources.

Human-induced landscape change disturb animal communities and species abundance pattern with significant effects on virus prevalence in wildlife

Anthropogenic environmental change can impact community traits such as species richness and population densities which have been shown to increase the prevalence of viral infections in wildlife reservoirs. It has been postulated that especially host species that are more resilient to changes of their natural habitat may increase in numbers which in turn may affect the prevalence of directly transmitted viruses. We have carried out an ecological survey of small mammal communities in three tropical landscapes differing in their degree of environmental change in Central Panama and investigated the effects of community changes on Hepacivirus prevalence, a virus closely related to human hepatitis C virus. The modification of continuous habitat into partly connected or isolated habitat patches during the past century was linked to changes in species richness and species assemblages, which in turn was associated with shifts in the abundance of generalist marsupial and rodent species. In particular, the spiny rat Proechimys semispinosus has become dominant in isolated habitat patches. Landscape-specific host density represents the most important ecological driver influencing local Hepacivirus prevalence. Our study provides important empirical data on how human-induced landscape changes may affect virus prevalence in wildlife and emphasizes the importance of a landscape scale approach considering the complex interactions between ecological factors driving host-virus interactions.


The best smellers make the best choosers: MHC-dependent mate choice is affected by female chemosensory receptor gene diversity

Sexual selection involving genetically disassortative mate choice is one of several evolutionary processes that can enhance population genetic variability and pathogen resistance. In many vertebrate species, females select mates depending on their major histocompatibility complex (MHC) genes. The products of the MHC genes are known to be drivers of pathogen resistance and sexual selection enhancing offspring genetic diversity. MHC further influences individual odor types and social communication. However, little is known about the receptors and their volatile ligands that are involved in this type of chemical communication. We investigated the effect of two groups of chemosensory receptor genes (trace-amine associated receptors (TAARs) and olfactory receptors (ORs)) on MHC-dependent mate choice. Our study is amongst the first to show a genetic link between behavior and chemosensory receptor genes. These results contribute to understanding the link between genetics, olfaction and associated life history decisions.


Extreme MHC diversity in bats with wide geographic ranges

Different hypotheses may explain extreme MHC variability. One is that such variability is attributable to adaptation to a wide geographic range and a diverse array of habitats, as found in passerine birds. Here we show that MHC class I diversity in Seba’s short-tailed bats (Carollia perspicillata), a widely distributed, generalist, neotropical species, shows a remarkable individual and population-level diversity and length-polymorphism comparable with passerine birds. Investigation of the details of the underlying adaptive processes and the role of the high MHC diversity in pathogen resistance are important next steps for a better understanding of the role of bats in viral evolution and as carriers of several deadly zoonotic viruses.


How low is functional MHC diversity in cheetahs?

Species that experienced a population bottleneck, a sharp reduction in the size of a population, often show a low genetic variability, also in adaptive immune relevant genes required to defend against parasites and pathogens. Free-ranging Namibian cheetahs (Acinonyx jubatus), however, despite being a textbook example of depleted genetic variability due to habitat loss, human-wildlife conflicts and historic bottlenecks, show no signs of impaired immunocompetence or health - thus contradicting the theoretical assumptions. How are cheetahs able to defend themselves against diseases? This study revealed that free-ranging Namibian cheetahs indeed possess only a low number of immune gene alleles at the major histocompatibility complex (MHC) comparable to other bottlenecked (endangered) cat species. However, these remaining alleles are functionally very divergent and might (currently) be sufficient in terms of pathogen recognition and initiation of the immune defence. Moreover, the allelic composition of cheetahs influences the level of MHC expression (so far only known from human research) which could add to functional variation and, thus, might play a role in the ability to defend against pathogens. This offers an explanation as to how bottlenecked cat species might have avoided impaired immuno-competence, despite showing low MHC allelic diversity.


Corridor effects on the genetic diversity of mouse lemurs

Corridors rank top among the recommendations to counteract the effects of habitat modification and reduce the vulnerability of small populations to environmental variation and stochastic processes, such as genetic drift. Our study focuses on the effect of the establishment of corridors on neutral (microsatellites) and adaptive (MHC) genetic diversity in Microcebus ganzhorni, a recently discovered primate species restricted to a few patches of littoral forest in south-eastern Madagascar. While we find similar patterns of neutral and adaptive individual diversity as suggested from estimates of heterozygosity and allelic richness between sampling periods, we provide evidence that after the establishment of corridors a larger number of shared private alleles are found at microsatellite loci. Furthermore, our results confirm the role of selection as a main driver of MHC II diversity in M. ganzhorni. We argue that corridors, even for animals that appear to be robust to fragmentation, might play an important role in population dynamics. This work highlights the relevance of long-term genetic monitoring providing insights into the evolutionary history and patterns of gene flow of wild
populations and the possibility to tease apart the effects of drift from selection maintaining MHC adaptive variability.


### Method development for microbiome and MHC research

#### A novel workflow to improve multi-locus genotyping of wildlife species

Genotyping novel complex multigene systems is particularly challenging in non-model organisms. Target primers frequently amplify simultaneously multiple loci leading to high PCR and sequencing artefacts such as chimeras and allele amplification bias. Most genotyping pipelines have been validated in non-model systems whereby the real genotype is unknown and the generation of artefacts may be highly repeatable. Further hindering accurate genotyping, the relationship between artefacts and copy number variation (CNV) within a PCR remains poorly described. We developed a novel open-source genotyping pipeline (ACACIA) to the data, and compared its performance with another, previously published, pipeline. ACACIA yielded very high allele calling accuracy (>98%). We discuss in detail the pitfalls researchers should avoid in order to reliably genotype complex multigene systems.


#### Can the estimation of core gut microbiomes be standardized across studies?

The filtering of faecal microbial datasets to retain high prevalence taxa is often performed to identify a common core microbiome that may be important for host biological functions. The common core gut microbiome is the subset of gut microbes that are particularly common across the host population, and are often identified based on prevalence (e.g. microbes that occur in over 50% of sampled individuals). However, these thresholds - though frequently used to compare results across studies and species - vary substantially across studies, and it is unclear whether gut microbial communities sourced from different host species demonstrate universal or species-specific responses to increasing prevalence thresholds. This has consequences for how comparable studies of core microbiomes are. We compared macroecological patterns in prevalence and abundance of eight gut microbial datasets from different mammal and bird species, and tested the effect of increasing prevalence thresholds on eight measures of alpha and beta diversity. Our results highlight some critical differences in macroecological patterns of rare taxa across the different datasets that can guide future sample collection and sample size, yet show that despite these differences, all microbial communities demonstrated similar responses in alpha diversity and beta dissimilarity to increasing prevalence thresholds. This suggests that most gut microbial communities exhibit similar patterns in prevalence and abundance distributions, especially of dominant taxa. Based on our results, we recommend methods that increase comparability of studies that identify a common core.

Jumping the green wall: the use of PNA-DNA clamps to enhance microbiome sampling depth in wildlife microbiome research

As microbiome research moves away from model organisms to wildlife, new challenges for microbiome high throughput sequencing arise caused by the variety of wildlife diets. Normally, high levels of contamination from the host (mitochondria) or diet (chloroplast) are commonly observed in wildlife samples. This contamination “hijacks” reads thus decreasing the overall sequencing depth of wildlife samples and reducing statistical power in downstream analysis. We developed an amplification protocol utilizing PNA-DNA clamps to maximize the use of resources and to increase the sampling depth of true microbiome sequences in samples with high levels of plastid contamination. The PNA-DNA clamps are DNA mimicking molecules that bind selectively to a target sequence. We used these clamps to block the sequences of chloroplast and mitochondria in fecal samples of birds and bats. Our protocol successfully blocks the signal organelle signals and provides a 13-fold increase in bacterial sequence amplification in comparison with the Earth Microbiome Protocol.


Home-made cost-effective preservation buffer stands the test against commercial preservation methods for microbiome research

The investigation of wildlife gastrointestinal microbiomes is a growing field in microbial ecology and conservation. Such studies often face difficulties in sample preservation if neither freezing facilities nor liquid nitrogen (LQN) are readily available. Thus, in order to prevent microbial community changes because of bacterial growth after sampling, preservation buffers need to be applied to samples. However, the amount of microbial community variation attributable to the different preservation treatments and potentially affecting biological interpretation is hardly known. Using sheep feces, we analyzed the effect of air-drying, an inexpensive self-made nucleic acid preservation buffer (NAP), DNA/RNA Shield™, and RNAlater®, each together with freezing or storing at room temperature prior to 16S rRNA gene high-throughput sequencing to determine bacterial communities. Overall, NAP had better preservation qualities than RNAlater® and DNA/RNA Shield™ making this self-made buffer a valuable solution in wildlife microbiome studies.