

HOST-PATHOGEN INTERACTIONS

Carbohydrate-dependent B cell activation by fucose-binding bacterial lectins

Isabel Wilhelm^{1,2,3*}, Ella Levit-Zerdoun^{4,5,6*}, Johanna Jakob⁷, Sarah Villringer^{1,3}, Marco Frensch^{1,3,5}, Rudolf Übelhart⁷, Alessia Landi^{1,3}, Peter Müller^{1,3}, Anne Imberty⁸, Roland Thuenauer^{1,3}, Julie Claudinon^{1,3}, Hassan Jumaa⁷, Michael Reth^{1,3,4}, Hermann Eibel^{9,10}, Elias Hobeika^{7†‡}, Winfried Römer^{1,2,3†‡}

Bacterial lectins are typically multivalent and bind noncovalently to specific carbohydrates on host tissues to facilitate bacterial adhesion. Here, we analyzed the effects of two fucose-binding lectins, BamBL from *Burkholderia ambifaria* and LecB from *Pseudomonas aeruginosa*, on specific signaling pathways in B cells. We found that these bacterial lectins induced B cell activation, which, in vitro, was dependent on the cell surface expression of the B cell antigen receptor (BCR) and its co-receptor CD19, as well as on spleen tyrosine kinase (Syk) activity. The resulting release of intracellular Ca²⁺ was followed by an increase in the cell surface abundance of the activation marker CD86, augmented cytokine secretion, and subsequent cell death, replicating all of the events that are observed in vitro upon canonical and antigen-mediated B cell activation. Moreover, injection of BamBL in mice resulted in a substantial, BCR-independent loss of B cells in the bone marrow with simultaneous, transient enlargement of the spleen (splenomegaly), as well as an increase in the numbers of splenic B cells and myeloid cells. Together, these data suggest that bacterial lectins can initiate polyclonal activation of B cells through their sole capacity to bind to fucose.

INTRODUCTION

Pseudomonas and *Burkholderia* species are proteobacteria, which release various lectins and form biofilms (1, 2). These bacteria cause opportunistic infections and exhibit multidrug antibiotic resistance (2–4). *Burkholderia ambifaria* belongs to the *Burkholderia cepacia* complex, a group of closely related bacterial strains, which cause so-called cepacia syndrome (4). The fatal combination of necrotizing pneumonia, respiratory failure, and bacteremia has been observed in the lung infections of patients suffering from cystic fibrosis (CF), as well as in nosocomial infections of other immunocompromised hosts (5–7). Furthermore, *B. ambifaria* species were identified in a polyclonal bacteremia outbreak caused by contaminated ultrasound gel in Brazil in 2015, but the epidemiology of *B. ambifaria* remains poorly characterized to date (7).

In addition to many other virulence factors, these bacteria produce soluble carbohydrate-binding proteins, so-called lectins. *Pseudomonas aeruginosa* expresses two tetravalent lectins, LecA and LecB (also called PA-IL and PA-IIL), with carbohydrate-binding specificities for galactose and fucose, respectively (8). The *B. ambifaria* lectin BamBL

(gene annotation across databases: *bamb_5415*; UniProt-ID: Q0B4G1_BURCM; NCBI-ProteinID: ABI90962; KEGG-ID: bam:Bamb_5415) contains six carbohydrate-binding sites (9). Both LecB and BamBL display exceptionally high affinities for fucose, with the respective K_d (dissociation constant) values in the micromolar range, which is rather unusual for a lectin-monosaccharide interaction (9). Several studies point to an underestimated, highly complex role of bacterial lectins in affecting host cellular processes, going far beyond an adhesive function (10–14). In particular, there is very limited knowledge about the influence of soluble bacterial lectins on cells of the immune system (15, 16).

However, many receptors of immune cells, including the B cell antigen receptor (BCR), are highly glycosylated and thus present potential targets for lectins (17–19). The BCR, which is expressed by B lymphocytes, plays a pivotal role in the humoral immune response. In mammals, B lymphocyte development follows a strict, multistep maturation program starting in the bone marrow from hematopoietic stem cells through progenitor (pro-) to precursor (pre-) cell stages. The pre-B cells are defined by their expression of the pre-BCR and further develop into immature B cells, which mainly express a BCR of the immunoglobulin M (IgM) isotype (20, 21). Subsequently, these immature B cells migrate to the periphery and secondary lymphoid organs such as the spleen and the lymph nodes, where they undergo further differentiation, eventually coexpressing IgM- and IgD-BCR isotypes of the same antigen specificity. IgD expression increases with progressing differentiation with mature B cells expressing more IgD than IgM (20, 22).

The BCR comprises two heavy chains (HCs), two light chains (LCs), and the Ig α /Ig β (CD79a/CD79b) heterodimer, which functions as the signaling component of the BCR complex (23–25). Mature B cells also carry a co-receptor complex comprising the molecules CD19, CD81, and CD21 (26). After B cell activation, CD19 is found in close proximity to the IgM-BCR and signals through the phosphoinositide 3-kinase (PI3K) pathway (27). B cell survival and differentiation are also

¹Faculty of Biology, University of Freiburg, 79104 Freiburg, Germany. ²Spemann Graduate School of Biology and Medicine (SGBM), University of Freiburg, 79104 Freiburg, Germany. ³Signalling Research Centres BIOS and CIBSS, University of Freiburg, 79104 Freiburg, Germany. ⁴Max Planck Institute of Immunology and Epigenetics Freiburg, 79108 Freiburg, Germany. ⁵International Max Planck Research School (IMPRS), Max Planck Institute of Immunobiology and Epigenetics Freiburg, 79108 Freiburg, Germany. ⁶German Cancer Consortium (DKTK) partner site Freiburg, German Cancer Center (DKFZ), Heidelberg, Institute of Molecular Medicine and Cell Research, 79104 Freiburg, Germany. ⁷Institute for Immunology, University Medical Centre Ulm, 89081 Ulm, Germany. ⁸Univ. Grenoble Alpes, CNRS, CERMAV, 38000 Grenoble, France. ⁹CCI-Center for Chronic Immunodeficiency (CCI), University Medical Centre, 79106 Freiburg, Germany. ¹⁰Faculty of Medicine, University of Freiburg, 79106 Freiburg, Germany.

*These authors contributed equally to this work.

†These senior authors contributed equally to this work.

‡Corresponding author. Email: elias.hobeika@uni-ulm.de (E.H.); winfried.roemer@bios.uni-freiburg.de (W.R.)

promoted by the B cell-activating factor of the tumor necrosis factor (TNF) family (BAFF), which is also known as B-Lys (28, 29). The BCR can be activated by monovalent and polyvalent antigens (30, 31). IgD-BCRs are specialized in the binding of multivalent antigens, whereas IgM-BCRs can bind to both types of antigen (30, 32, 33).

The constant region of both IgM- and IgD-BCRs is N-glycosylated with glycans of the complex type, whereas the pre-BCR is highly mannoseylated (18). Fucosylated epitopes can be part of N-linked or O-linked sugar chains. However, because of the challenges in resolving the glycosylation patterns of the BCR with spectrometric techniques, the exact amounts and sites, particularly of O-linked glycans, have not been elucidated to date (34–36). Furthermore, the number and position of N-linked glycosylation sites may vary between murine and human BCRs (34, 36). However, there are indications that glycosylation events can affect B cell development in mice, wherein a specific glycosylation site (Asn⁴⁶, N46) on IgM is required for efficient pre-BCR function and further B cell maturation (17). Furthermore, the glycosylation pattern of the BCR changes during carcinogenesis: Follicular lymphoma cells acquire carbohydrates of the high mannose type in the variable domain of the BCR, generating binding sites for mannose-binding lectins (19, 37). Moreover, the Ig α and whole BCR molecules carry altered glycans in chronic lymphocytic leukemia, influencing BCR signaling (38).

In this study, we found that the fucose-specific bacterial lectins BambL and LecB bound to murine B cells and induced an intracellular signaling cascade that changed the flux of Ca²⁺. In vitro, activation of this cascade by BambL required expression of the BCR, as well as of its co-receptor CD19. Stimulation by BambL induced canonical BCR-mediated signaling events that resulted in a polyclonal B cell death, which was partially reversed by BAFF-induced signaling. In vivo, intraperitoneal injection of BambL resulted in lymphopenia in the bone marrow combined with a transient splenomegaly, as well as accumulation of B cells and myeloid cells in the spleen. These events were independent of BCR signaling because they were recapitulated in BCR-deficient mice. We propose that these bacterial lectins act as superstimulatory antigens, activating B cells in a BCR-dependent manner in vitro by binding to fucosylated epitopes and eliciting a strong, but transient, BCR-independent immune response in vivo.

RESULTS

BambL induces Ca²⁺ release that is dependent on BCR expression and Syk activity

Upon antigen binding, the immunoreceptor tyrosine-based activation motifs of the Ig α / β subunits are phosphorylated, activating an intracellular signaling cascade (23, 39). As a part of the second messenger cascade, the intracellular Ca²⁺ concentration changes by store-operated calcium entry through the Ca²⁺ release-activated Ca²⁺ channels (40, 41). To assess whether the two fucose-binding lectins, LecB and BambL, activated primary murine B cells, we performed intracellular Ca²⁺ flux measurements in vitro (Fig. 1). The hexavalent BambL (10 μ g/ml) induced a Ca²⁺ release similar to that induced by BCR stimulation with anti- κ (hereafter anti-kappa) antibodies (10 μ g/ml) in mature follicular (MFO) B cells (Fig. 1A). Saturating the BambL carbohydrate binding sites with soluble L-fucose (25 mg/ml) prevented BambL binding to B cells (fig. S1A) and Ca²⁺ flux (Fig. 1A), indicating that this activation process was fucose specific. The mitogenic potential of LecB on murine splenocytes was described previously (15). Here, we demonstrated that binding of the tetravalent LecB (10 μ g/ml)

to B cells also induced a Ca²⁺ signal but to a somewhat lesser extent than that induced by the hexavalent BambL (Fig. 1A).

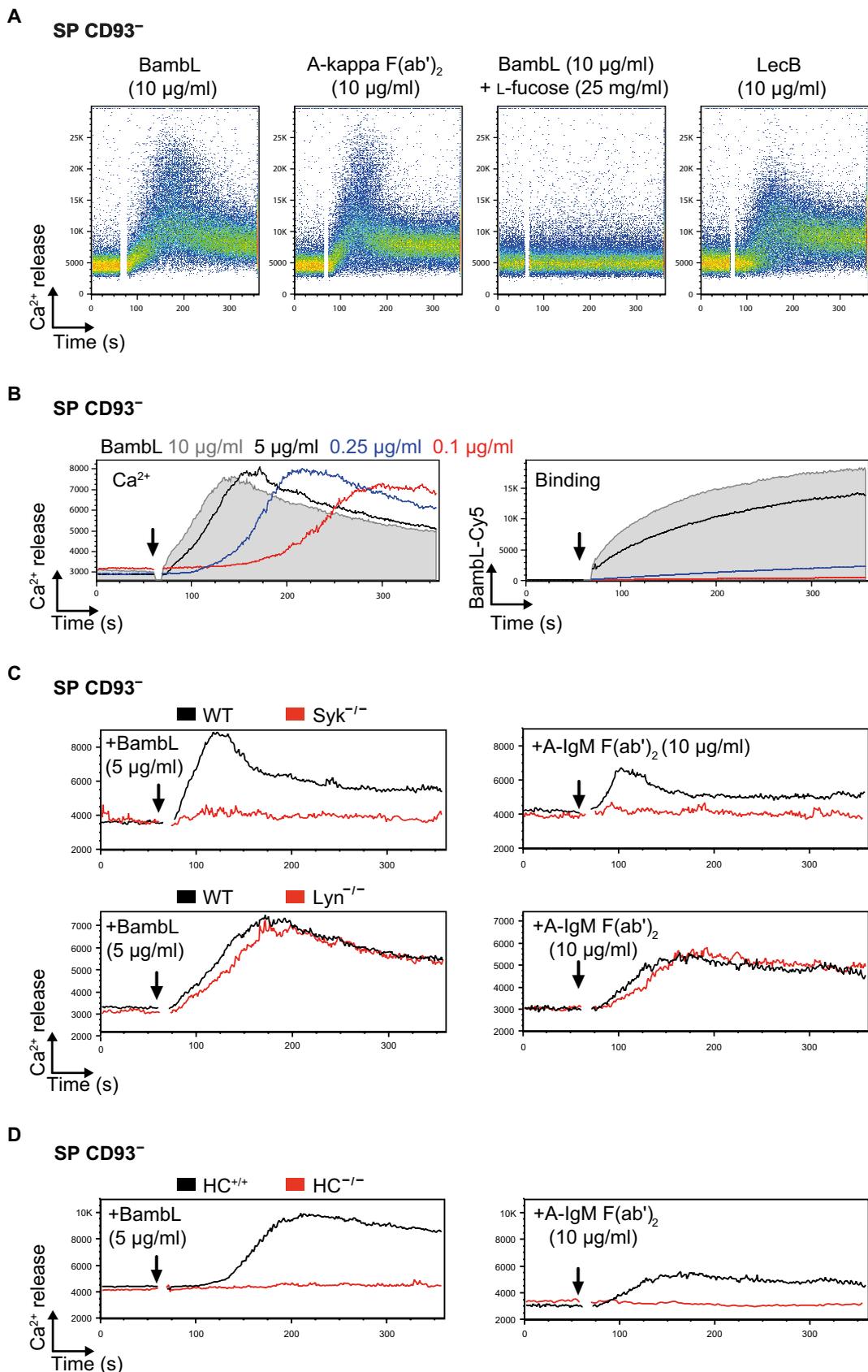
To assess whether the strength of lectin binding correlated with the extent of Ca²⁺ flux, we analyzed the binding of both lectins by flow cytometry and found that both lectins bound to B cells. LecB is tetravalent with a molecular mass of 46.924 kDa, whereas BambL is hexavalent with a molecular mass of 28.143 kDa. Because of its hexavalency, the avidity of BambL is increased, which resulted in higher binding capacity at similar concentrations in comparison to LecB (fig. S1A). The binding capacity may be further influenced by the differences in BambL lectin structure in comparison to LecB, as well as the affinity of BambL for different host cell receptors. Furthermore, the lower MW of BambL supposedly leads to the binding of more molecules per cell (fig. S1A). With BambL and LecB showing similar effects on B cells, we focused on the characterization of BambL binding. Additional binding assays demonstrated that BambL bound to all CD19-positive splenic B cells, as well as T cells (fig. S1B). Analyzing Ca²⁺ mobilization in response to BambL binding in terms of dose dependency, we found a clear correlation in the range between 0.1 and 10 μ g/ml of the lectin (Fig. 1B). To dissect the role of individual components of the BCR-associated signaling cascade in the response to BambL, we analyzed Ca²⁺ mobilization using B cells from mice lacking spleen tyrosine kinase (Syk), the Src-family kinase Lyn, or the BCR HC. Syk and Lyn are the most proximal kinases to the BCR and are involved in canonical B cell activation (25, 42). Similar to the Ca²⁺ release upon BCR stimulation by anti-IgM F(ab')₂, the BambL-mediated Ca²⁺ release was independent of Lyn but required the presence of the kinase Syk (Fig. 1C). Note that BambL efficiently bound to Syk-deficient B cells (fig. S1C). Stimulation with anti-IgM served as a control for Ca²⁺ release in these experiments; however, because of normalization of the y axis to the BambL-mediated Ca²⁺ release experiments, we cannot directly compare our data to those from previous studies (42, 43).

In a similar approach, we monitored Ca²⁺ release in B cells lacking surface BCR expression using mice carrying a B1-8 HC allele flanked by loxP sites (B1-8^{fl}) on an inducible mb1-CreERT^{T2} “deleter” background (44, 45). These mice express a B cell-specific Cre recombinase from the Ig α -encoding *mb-1* locus, which, upon tamoxifen treatment, induces the deletion of the B1-8^{fl} allele, resulting in the generation of HC-deficient B cells. The HC-deficient B cells were obtained at day 7 after tamoxifen treatment when an abundant number of these cells were still present in mb1-CreERT^{T2}; B1-8^{fl} mice before they were lost at later time points (44–46). These cells (denoted as HC^{-/-} cells), identified by gating on IgM- and IgD-negative populations (fig. S1D), displayed no detectable Ca²⁺ flux when stimulated with BambL despite effectively binding BambL (Fig. 1D and fig. S1E). The efficiency of tamoxifen-induced Cre-mediated recombination is not 100% in the mb1-CreERT^{T2}; B1-8^{fl} mice. Only about 15 to 20% of B cells lost HC 7 days after tamoxifen treatment. Therefore, IgM- and IgD-positive B cells were still detected in these animals and served as an internal HC-sufficient control (HC^{+/+} cells; Fig. 1D and fig. S1C). These data indicate that BambL-dependent activation of B cells requires the presence of the BCR and Syk to mobilize intracellular Ca²⁺ and is abrogated in the presence of L-fucose.

BambL-induced B cell activation is restricted to IgD-expressing B cells

We next examined whether activation of B cells by BambL differed between pre- and mature B cells. To this end, we compared

Fig. 1. BCR and Syk are essential for BamBL-induced Ca²⁺ release in primary B cells. (A) Flow cytometric analysis of Ca²⁺ release in splenic (SP) CD93⁻ MFO B cells from wild-type (WT) mice upon stimulation with BamBL (10 μg/ml), LecB (10 μg/ml), or anti-kappa F(ab')₂ [10 μg/ml]; hereafter, a-kappa F(ab')₂] antibody (Ab) as a control. BamBL preincubated with L-fucose served as a control for BamBL fucose-binding specificity. Data are representative of four independent experiments. (B) Dose-dependent stimulation of MFO B cells with BamBL was monitored by flow cytometric analysis of Ca²⁺ release (left) and binding of BamBL (right) to these cells. For the binding analysis, the relative fluorescence of BamBL-Cy5 over time is shown. Data are representative of three independent experiments. (C) Flow cytometric analysis of Ca²⁺ release in WT (black) and Syk^{-/-} (red, top) or Lyn^{-/-} (red, bottom) MFO B cells, stimulated with BamBL (10 μg/ml; left) or anti-IgM F(ab')₂ [10 μg/ml; hereafter, a-IgM F(ab')₂] Ab (right). Data are representative of three independent experiments. (D) Flow cytometric analysis of Ca²⁺ release in BCR-negative B cells (HC^{-/-}, red) as well as in BCR-sufficient B cells from the same mouse (HC^{+/+}, black). Cells were stimulated with BamBL (10 μg/ml; left) or anti-IgM F(ab')₂ (10 μg/ml; right). Data are representative of four independent experiments. The gating strategy for HC^{-/-} cells and the HC^{+/+} control is depicted in fig. S1C.



Downloaded from <http://stke.sciencemag.org/> on March 5, 2019

BambL-triggered Ca^{2+} release in bone marrow–derived and splenic B cells. We stained bone marrow–derived and splenic B cells with antibodies to detect the expression of typical cell surface markers to distinguish among the different B cell maturation stages (fig. S2). In contrast to incubation of the cells with anti-IgM F(ab')₂, incubation with BambL did not result in Ca^{2+} flux in pre-B cells or immature B cells (Fig. 2A), although BambL bound to both subsets of cells (fig. S2B). Similarly, transitional 1 (T1) B cells from the spleen did not release Ca^{2+} , whereas splenic T2/T3, marginal zone (MZ), and MFO B cells displayed changes in intracellular Ca^{2+} concentrations upon treatment with BambL (Fig. 2A). Immature and T1 B cells express little to no IgD (fig. S2D) (22); therefore, we suspected that the activation of mature B cells by BambL was IgD dependent. To test this hypothesis, we analyzed BambL-induced Ca^{2+} flux using bone marrow–derived B cells from IgM^{-/-} mice (47). In these mice, the DNA sequences encoding the constant region of the μHC have been removed, enforcing the expression of IgD starting from early maturation stages. We observed that BambL increased intracellular Ca^{2+} concentrations in these B cells (Fig. 2B). These data demonstrate that the IgM-BCR was not required for BambL-induced Ca^{2+} release and suggest that the IgD, but not the IgM, isotype of the BCR is required for BambL-induced Ca^{2+} release in mature B cells.

B cell blasts treated with the Toll-like receptor 4 (TLR4) agonist lipopolysaccharide (LPS) down-regulate cell surface IgD and differentiate into IgM^{high} plasmablasts (48). We therefore compared Ca^{2+} release in LPS-treated B cells isolated from WT and IgM^{-/-} mice, which had been crossed with Blimp-1^{GFP} mice. In the latter, a green fluorescent protein (GFP) reporter is expressed from the *Prdm1* gene locus, which encodes the transcription factor Blimp-1, a master regulator of plasma cell development expressed at early stages of plasma cell differentiation (49, 50). Plasmablasts were identified on the basis of the expression of GFP and the plasma cell marker Syndecan-1 (CD138). WT plasmablasts displayed only a moderate Ca^{2+} release upon exposure to BambL, whereas IgM^{-/-} plasmablasts, which expressed only IgD, exhibited a greater intracellular Ca^{2+} response (Fig. 2C). These results further support our hypothesis that only IgD-bearing B cells are activated by BambL.

BambL-induced murine B cell activation also requires CD19

To corroborate our findings that BambL mobilized Ca^{2+} only in IgD⁺ B cells, we compared changes in Ca^{2+} concentration in IgM- and in IgD-deficient B cells after incubation with different concentrations of BambL. As expected, we observed that the IgD^{-/-} B cells expressing only IgM (fig. S3) were able to bind to BambL but failed to mobilize Ca^{2+} , which was in contrast to the responses of WT B cells and B cells expressing IgD alone (Fig. 3, A and B). Because the IgD-BCRs of resting mature B lymphocytes are reportedly organized in “protein islands” with the co-receptor CD19 in close proximity (51, 52), we assumed that the IgD-dependent activation of B cells by BambL might also involve CD19. Although CD19-deficient B cells bound to BambL to a similar extent as did WT B cells, they released much less Ca^{2+} than did WT B cells upon exposure to BambL (Fig. 3, C and D). B cells from mice lacking both IgD and CD19 but expressing IgM (identified by CD19 and IgD staining; fig. S3) were still able to bind to BambL but did not mobilize the intracellular Ca^{2+} flux even at a very high BambL concentration (10 $\mu\text{g}/\text{ml}$) (Fig. 3, C and D). Together, these data suggest that the surface expression of both IgD and CD19 is indispensable for the BambL-induced activation of primary mature B cells.

CD19, IgM, and IgD are internalized upon BambL binding

Upon binding to the BCR of a cognate antigen, the receptor complex becomes internalized by endocytosis (53). Because CD19 and IgD were essential for BambL-mediated B cell activation, we tested whether BambL induced the endocytosis of these surface molecules. As another control, we also monitored the cell surface expression of IgM. After 16 hours of incubation with BambL, murine B cells expressed substantially reduced amounts of CD19, IgD, and, unexpectedly, also of IgM on the surface compared to those of unstimulated cells or B cells stimulated with anti-IgM (Fig. 4, A and B). Moreover, the effects were inhibited in the presence of L-fucose (Fig. 4, A and B). We assumed that binding to and subsequent internalization of CD19, IgD, and IgM were specific, because other heavily glycosylated molecules, such as B220 (CD45R), showed no decrease in cell surface abundance after 16 hours of incubation with BambL (fig. S4). Together, these observations demonstrate that BambL leads to the internalization of CD19, IgD, and, to a lesser degree, IgM.

BambL induces an increase in CD86 abundance and TNF- α secretion

To further elucidate the physiological effects of BambL-mediated activation, we sought to determine whether early B cell activation markers, such as CD86 (B7.2), were increased in response to BambL (54). CD86 is a highly glycosylated molecule and was internalized upon BambL incubation (fig. S5A). Thus, we established a “BambL wash-out” protocol in which B cells were first incubated with BambL at 37°C for 15 min, then washed to remove unbound lectin, and left overnight to recover. We found that the increase in CD86 abundance in response to BambL was similar to that induced after activation with the TLR ligands cytosine-phosphate-guanine (CpG) oligonucleotides and LPS, although not to the same extent as that in response to anti-IgM F(ab')₂ (Fig. 5, A and B). The addition of L-fucose resulted in BambL inducing only a moderate increase in CD86 cell surface expression.

To confirm that the observed effect was dependent on the presence of the BCR rather than on an artifact caused by pattern recognition receptor stimulation, we assessed the response of BCR-deficient B cells to BambL. We did not observe an increase in CD86 abundance in these cells after BambL stimulation (Fig. 5C). Initial signaling events, such as Ca^{2+} flux, induce the differentiation of lymphocytes into effector cells. For B lymphocytes, the spectrum of effector cell function includes the secretion of cytokines (55). Analyzing the supernatants of BambL-treated B cells, we detected a marked increase in TNF- α production (fig. S5B) and secretion (Fig. 5D). TNF- α secretion by B cells is reported to be an outcome of canonical nuclear factor κB (NF- κB) signaling (56), which suggests that BambL activates the NF- κB pathway. In summary, treatment of B cells with BambL results in a sustained increase in CD86 abundance.

BambL leads to B cell death, which is partially reversible by BAFF

Strong intracellular signaling can drive B cells into activation-induced cell death (57–59). Because BambL binding to B cells resulted in the IgD- and CD19-dependent activation of intracellular signaling cascades, we tested the survival of splenic B cells after stimulation. After 24 hours of incubation with BambL, B cell viability decreased in a concentration-dependent manner (Fig. 6A). To characterize the type of BambL-induced cell death, we performed further viability assays using various inhibitors, which targeted pathways related to apoptosis, pyroptosis, and necroptosis. Pyroptosis can be induced by

Fig. 2. IgD-expressing B cell subsets release Ca^{2+} upon treatment with BambL. (A) Flow cytometric analysis of Ca^{2+} release in WT B cells at different developmental stages: bone marrow (BM)-derived pre-B cells (gray), immature B cells (red), and mature B cells (black). Cells were stimulated with BambL (5 μ g/ml; left) or anti-IgM F(ab')₂ (10 μ g/ml; right). The BM gating strategy is shown in fig. S2A. The splenic (SP) subsets are displayed in the middle and bottom panels. Middle: CD93⁺; transitional 1 (T1, red) and T2/3 (black). Bottom, CD93⁺; MFO (red) B cells and MZ (black) B cells. The SP gating strategy is depicted in fig. S2C. The relative cell surface IgD expression of the respective B cell subtypes is shown in fig. S2D. Data are representative of five independent experiments. (B) Flow cytometric analysis of Ca^{2+} release in IgM^{-/-} BM-derived B cells at different developmental stages. Cells were stimulated with BambL (5 μ g/ml; left) or anti-kappa F(ab')₂ (10 μ g/ml; right). BM subsets are displayed as pre-B cells (gray), immature B cells (red), and mature B cells (black). Data are representative of three independent experiments. (C) Flow cytometric analysis of Ca^{2+} release in WT (black) and IgM^{-/-} (red) plasmablasts after 4 days of incubation with LPS. Top: Representative analysis of the cell surface expression of CD19, IgD, and IgM. Cells were selected on the basis of Blimp-1-GFP and Syndecan-1 (CD138) expression. The cells were stimulated with BambL (5 μ g/ml; bottom left) or anti-kappa F(ab')₂ (10 μ g/ml; bottom right). Data are representative of three independent experiments.

infection with Gram-negative bacteria (including *Burkholderia*) but has rarely been observed in B cells (60–62). The initiation of pyroptosis is thought to depend on caspase-1 activity (63); however, treatment of primary B cells with the caspase-1 inhibitor did not alter BambL-mediated cytotoxicity (fig. S6A). In contrast, the survival of BambL-treated B cells increased upon their treatment with the apoptosis inhibitor QVD, and treatment with the necroptosis inhibitor Nec-1 resulted in detectable rescue effects (fig. S6A). Thus, B cells exposed to BambL most likely succumb to a combination of apoptosis and necroptosis.

Binding of BAFF to its receptor BAFF-R activates essential survival functions in transitional and mature B cells (29). We, therefore, tested whether BAFF could reverse or prevent BambL-induced cell death. However, coinubation with BAFF overnight only restored the viability of BambL-treated B cells by about 20% (Fig. 6B). In a 4-day time course experiment with a sublethal concentration of BambL, we observed

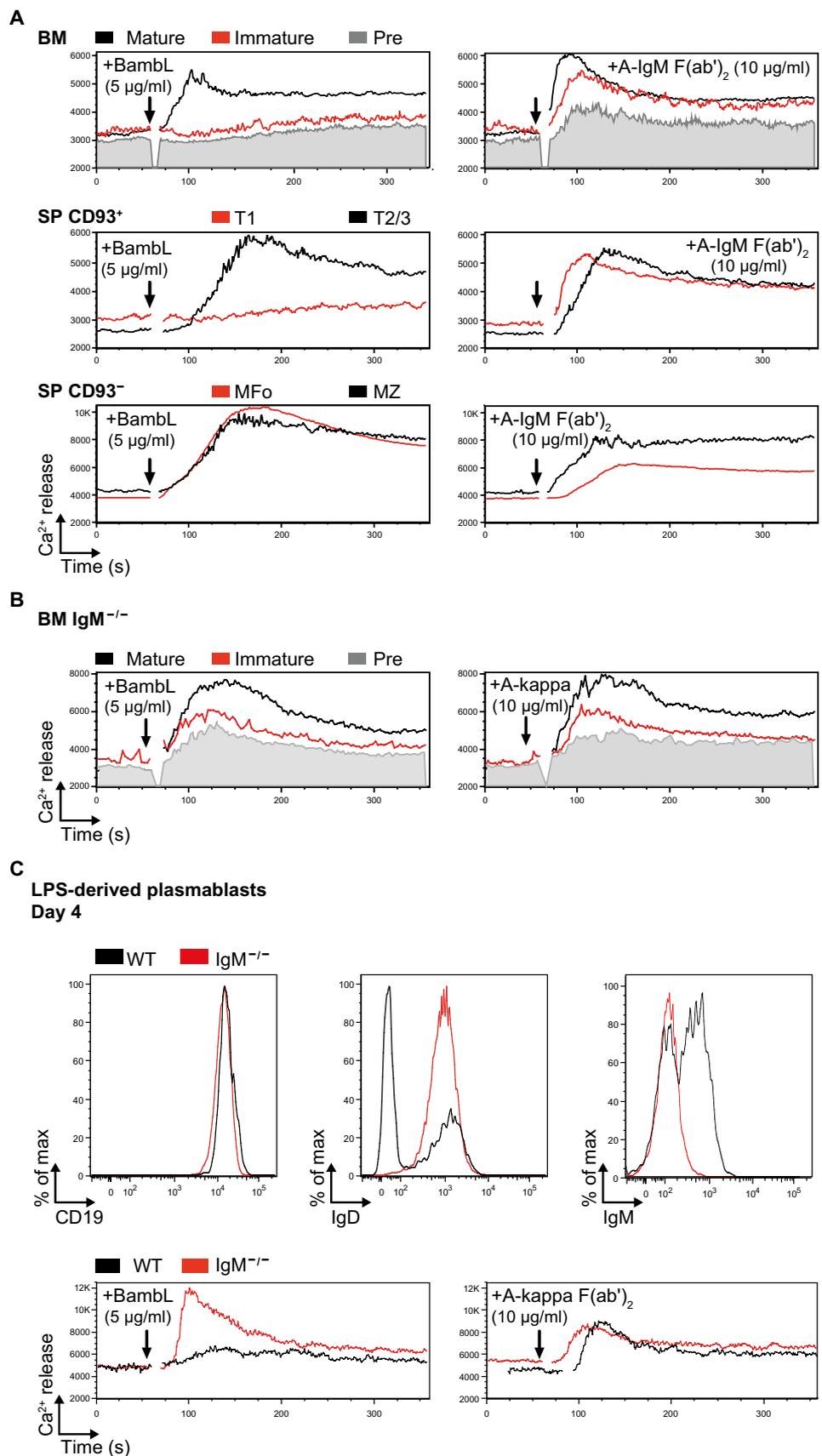


Fig. 3. IgD and CD19 play a major role in BamBL-induced signaling.

(A) Flow cytometric analysis of Ca^{2+} release in SP-derived MFO B cells (SP CD93⁻) from WT (black) and $IgM^{-/-}$ (red; top, $n = 3$) and $IgD^{-/-}$ (red; bottom, $n = 4$) mice. Cells were stimulated with BamBL (250 ng/ml; left), anti-IgD F(ab')₂ (10 μg/ml; top right), or anti-IgM F(ab')₂ (10 μg/ml; bottom right). (B) Flow cytometric analysis of BamBL-Cy5 binding to the corresponding B cells: WT (left), $IgM^{-/-}$ (middle), and $IgD^{-/-}$ (right). Data are representative of at least three independent experiments. (C) Flow cytometric analysis of Ca^{2+} release in WT (black), $CD19^{-/-}$ (red; top, $n = 3$), and $CD19^{-/-}IgD^{-/-}$ (red; bottom, $n = 3$) B cells. Cells were stimulated with BamBL (5 or 10 μg/ml; left) or anti-IgM F(ab')₂ (10 μg/ml; right). (D) Flow cytometric analysis of BamBL-Cy5 binding to the corresponding B cells: WT (left), $CD19^{-/-}$ (middle), and $CD19^{-/-}IgD^{-/-}$ (right). Data are representative of at least three independent experiments.

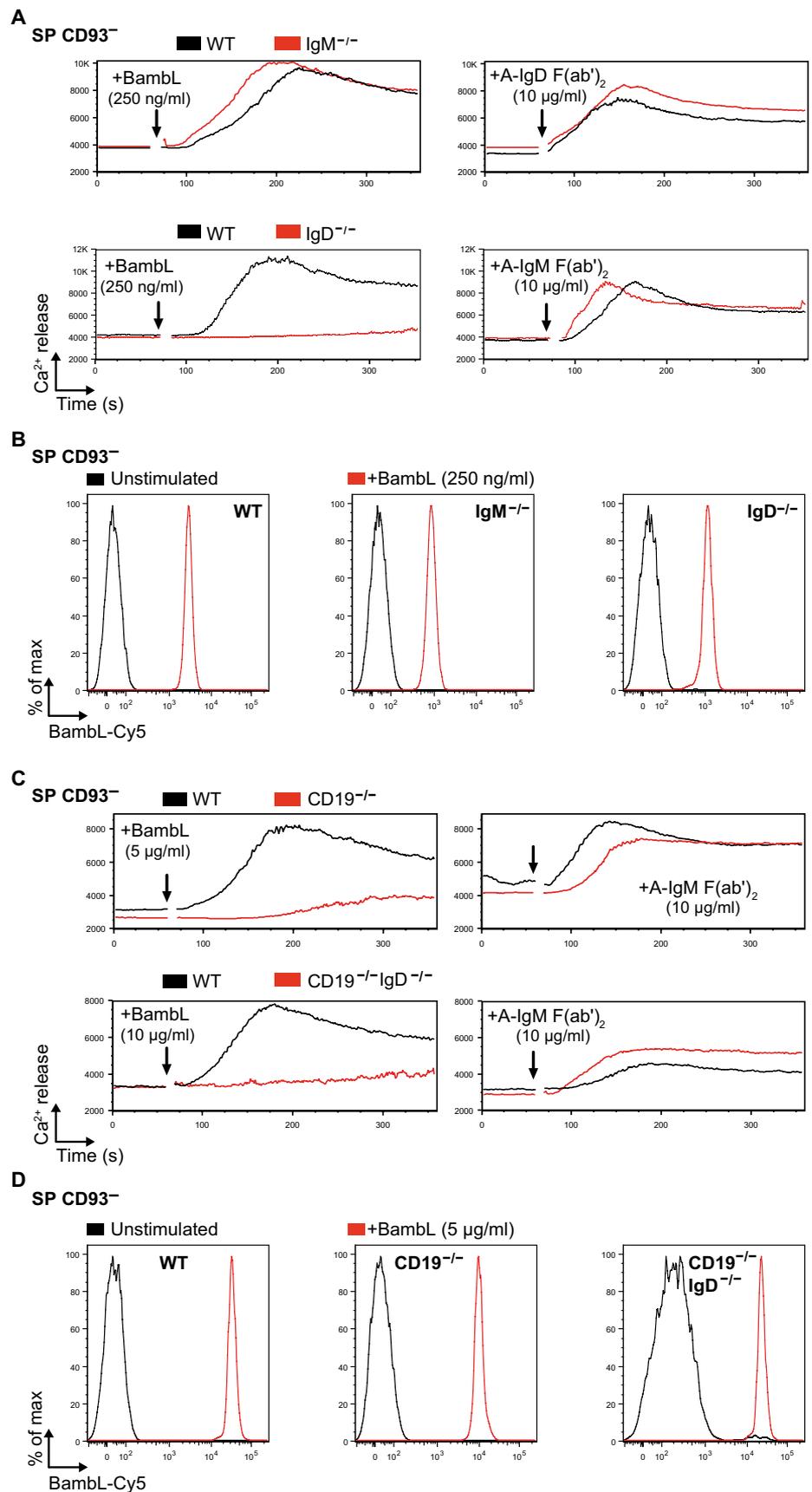
an increase in the viability of BamBL-treated B cells in the presence of BAFF (fig. S6B). This increased survival was not due to BAFF-dependent changes in BamBL binding (Fig. 6C). Together, these results suggest that BamBL induces B cell death, which can be partially rescued by BAFF.

In vivo administration of BamBL leads to splenomegaly and the loss of B cells from the bone marrow

To address the in vivo relevance of our in vitro findings, we injected BamBL (15 mg/kg) intraperitoneally into WT mice and assessed the effects of BamBL at days 3 and 7. We decided to use this concentration based on our observation that BamBL is bound by soluble Igs: Coincubation with Igs neutralized BamBL, reducing its binding to B cells and subsequently diminishing the intracellular Ca^{2+} release (fig. S7).

The mice were healthy before and after injection; the application of BamBL did not lead to weight loss or any other macroscopically detectable side effects. At day 3 after BamBL application, the mice exhibited splenomegaly with the spleens being on average 2.7-fold heavier compared to those of phosphate-buffered saline (PBS)-injected control mice (Fig. 7, A and B). On day 7, the spleen weights were statistically significantly reduced compared to those on day 3 but were still about twofold heavier than the spleens from the control mice (Fig. 7, A and B).

The total B cell numbers in the spleens of BamBL-treated mice were markedly increased, mainly due to the changes in the numbers of MFO, T1, and T2/3 populations (Fig. 7, C and D; for gating strategy, see fig. S8A), whereas the T cell numbers were not



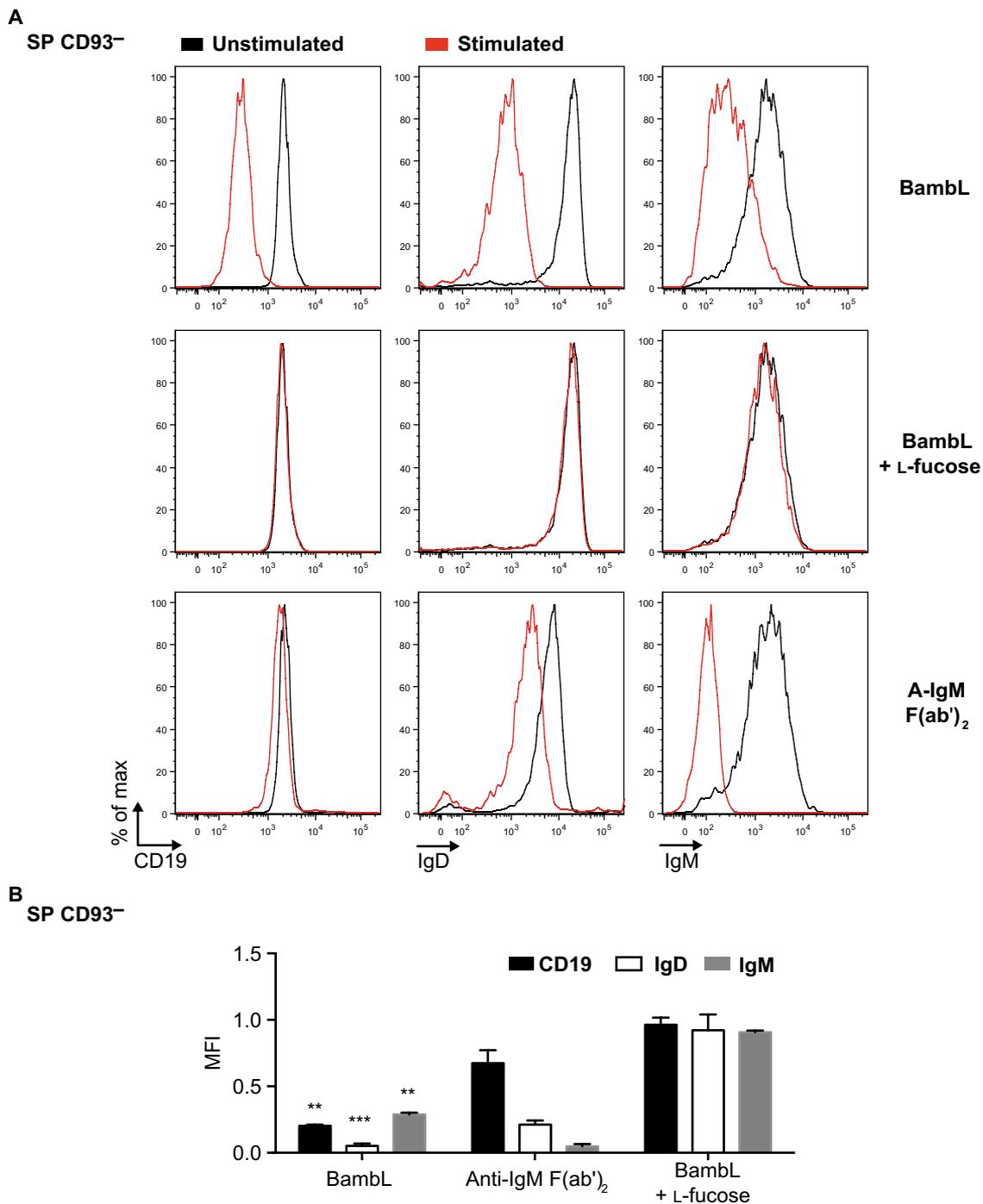


Fig. 4. Binding of BamBL leads to the internalization of CD19, IgD, and IgM and can be blocked by soluble L-fucose. (A) Purified splenic MFO B cells from WT mice were stimulated ex vivo in the presence of BamBL (2.5 μ g/ml), BamBL (2.5 μ g/ml) blocked with L-fucose (25 mg/ml), or anti-IgM F(ab')₂ and analyzed by flow cytometry after 16 hours of incubation for the cell surface expression of CD19, IgD, and IgM. Unstimulated cells served as a negative control for receptor expression. The combination of BamBL and L-fucose served as a control for the specificity of BamBL. Anti-IgM F(ab')₂ served as a positive control for BCR activation. Viable cells were selected using a fixable viability dye. Data are representative of six independent experiments. (B) Statistical analysis of data shown in (A). Mean values of one representative analysis with at least three technical replicates \pm SD are shown. For statistical analysis, a Student's *t* test was performed. ***P* < 0.01; ****P* < 0.001. MFI, mean fluorescence intensity.

affected (fig. S8B). The absolute cell counts of these populations had declined by day 7 (Fig. 7, C and D; for gating strategy, see fig. S8A). MZ B cell numbers were not affected in BamBL-treated mice (Fig. 7D; for gating strategy, see fig. S8A). Because of the successful

inhibition of cell death by QVD in vitro (fig. S6A) and its reported applicability in vivo (64, 65), we decided to coinject QVD with BamBL into WT mice to investigate the influence of caspases on BamBL-mediated effects in vivo. Caspase-6 and caspase-8 as well as

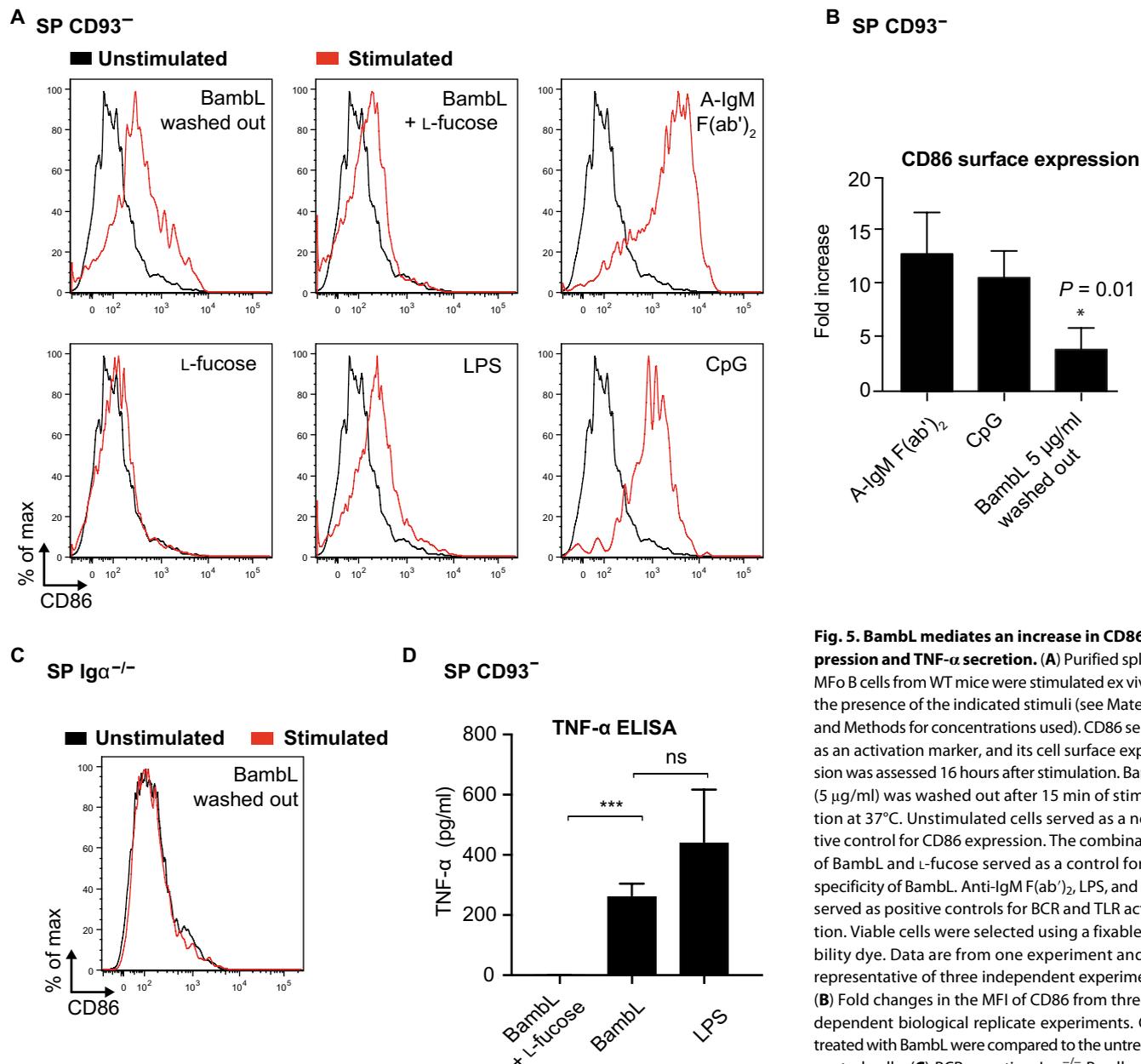


Fig. 5. BambL mediates an increase in CD86 expression and TNF- α secretion. (A) Purified splenic MFO B cells from WT mice were stimulated *ex vivo* in the presence of the indicated stimuli (see Materials and Methods for concentrations used). CD86 served as an activation marker, and its cell surface expression was assessed 16 hours after stimulation. BambL (5 μ g/ml) was washed out after 15 min of stimulation at 37°C. Unstimulated cells served as a negative control for CD86 expression. The combination of BambL and L-fucose served as a control for the specificity of BambL. Anti-IgM F(ab')₂, LPS, and CpG served as positive controls for BCR and TLR activation. Viable cells were selected using a fixable viability dye. Data are from one experiment and are representative of three independent experiments. (B) Fold changes in the MFI of CD86 from three independent biological replicate experiments. Cells treated with BambL were compared to the untreated control cells. (C) BCR-negative, Ig α ^{-/-} B cells were treated as described in (A) and examined by flow cytometry to determine the cell surface expression of CD86. The cells were obtained 10 days after tamoxifen treatment. Data are representative of at least three experiments. (D) B cells were stimulated for 16 hours with BambL (2.5 μ g/ml), LPS (10 μ g/ml), or BambL (2.5 μ g/ml) + L-fucose (25 mg/ml). The cell culture media were collected and analyzed by enzyme-linked immunosorbent assay (ELISA) to determine the amount of TNF- α released. Data are means \pm SD of three independent experiments and were analyzed by Student's *t* test to assess the statistical significance of observed differences. ****P* < 0.001; ns, not significant.

the para-caspase mucosa-associated lymphoid tissue lymphoma translocation protein 1 (MALT-1) have been described to play essential roles in B cell differentiation, death, and survival (59, 66, 67). We analyzed the BambL- and QVD-treated animals at day 3 after injection, when the full potential of the inhibitor could still be assumed. However, co-application of QVD *in vivo* did not statistically significantly affect either the size of the spleens or the B cell numbers in the spleen (Fig. 7, A to D).

In the bone marrow, lymphopenia was detected on day 3 after injection of BambL, which was marked by the loss of total B cells among

which the pre-, immature, and recirculation B cell subpopulations were statistically significantly reduced (Fig. 7, E and F; for gating strategy, see fig. S8C), whereas the total numbers of bone marrow-derived cells were not changed (fig. S8D). Similar to its effects in the spleen, QVD did not rescue the BambL-induced reduction in B cell numbers in the bone marrow (Fig. 7, E and F). The numbers of mature recirculating B cells were statistically significantly increased by day 7 compared to day 3 after injection with BambL alone or with QVD (Fig. 7E), suggesting that the B cell numbers were likely restored over time. Together, these data suggest that BambL induced a transient

Fig. 6. BAFF partially reverses B cell death induced by BambL. (A) Purified splenic MFO B cells from WT mice were stimulated with the indicated concentrations of BambL. Cell viability was assessed after 24 hours in parallel with that of an unstimulated control by flow cytometric analysis using the viability dye. Data are means \pm SD from three independent experiments and were analyzed by Student's *t* test to assess statistical significance. ****P* < 0.001; *****P* < 0.0001. (B) Purified splenic MFO B cells from WT mice were stimulated ex vivo in the presence of BambL (2.5 μ g/ml) with or without BAFF costimulation. The cells were analyzed 24 hours later by flow cytometry to generate forward scatter (FSC) and side scatter (SSC) plots and viability dye histograms. Cells negative for the viability dye were considered to be alive, whereas those cells positive for the dye were considered to be dead. Data are representative of four independent experiments. (C) Flow cytometric analysis of the binding of BambL-Cy5 to MFO B cells in the presence and absence of soluble BAFF (100 ng/ml). Data are representative of three independent experiments.

splénomegaly and expansion of B cells in the spleen with simultaneous loss of B cells from the bone marrow.

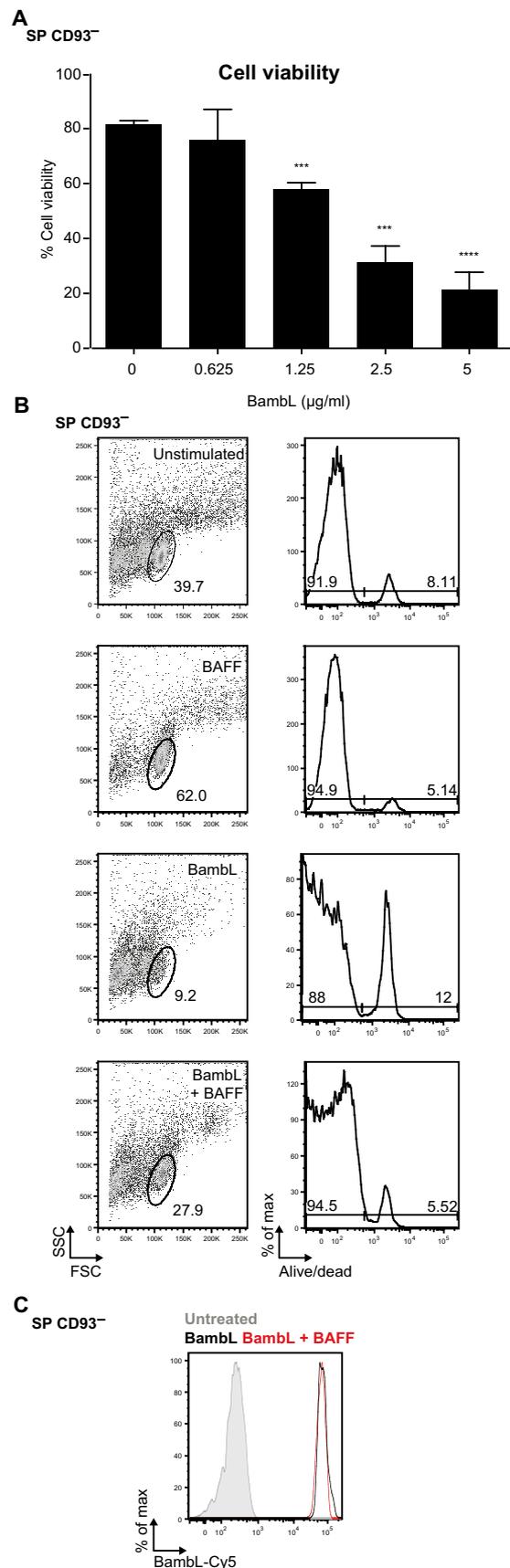
BambL administration induces the accumulation of myeloid cells in the spleen

During the analysis of BambL-injected mice, we observed an accumulation of large and granulated cells by flow cytometric analysis (fig. S8E). To determine the exact lineage of this population, we stained the splenocytes and bone marrow–derived cells for myeloid surface markers and calculated the absolute cell counts of myeloid populations based on a previously described gating strategy (68, 69). On day 3 after BambL injection, we found that the amount of total CD11b⁺ cells was statistically significantly increased in the spleen, specifically neutrophils, eosinophils, macrophages, and dendritic cells (DCs) (Fig. 8, A and B; for gating strategy, see fig. S9A). The cell numbers of these populations, with the exception of eosinophils, were reduced by day 7 (Fig. 8, A and B). We further distinguished inflammatory and circulating monocyte populations, which were still increased in number on day 7 (Fig. 8C). The addition of QVD did not substantially affect the numbers of total CD11b⁺ cells in BambL-treated mice. The inflammatory monocytes were statistically significantly increased in number after coinjection with QVD compared to those from mice treated only with BambL. In the bone marrow, the numbers of myeloid cells were not statistically significantly changed upon injection with BambL alone or together with QVD (Fig. 8, D to F; gating strategy, fig. S9B).

We stained the splenic and bone marrow–derived cells with the Red Flica active polycaspases kit and Sytox viability dye to assess caspase activity. On the basis of a similar gating strategy as one described earlier (fig. S8, A and B), we identified the individual B cell compartments as well as the total non-B cell population. We subsequently analyzed these populations by plotting Red Flica versus Sytox, which enabled us to assess the caspase activity (Red Flica⁺), to detect later stages of apoptosis (Red Flica⁺Sytox⁺), and to exclude necrotic cells (Sytox⁺). We found statistically significantly increased caspase activity (Red Flica⁺) in splenic non-B cell populations, which was reduced after QVD application (fig. S10, A and B, left). In the bone marrow, only the pro-B cells exhibited caspase activity, wherein the Red Flica⁺Sytox⁺ population (late apoptotic cells) was statistically significantly increased (fig. S10, C to F).

In vivo expansion of splenic B cells is independent of BCR signaling and of B cell development in the bone marrow

To assess whether the observed in vivo phenotype after BambL injection depended on BCR signaling, we injected mice with BambL,



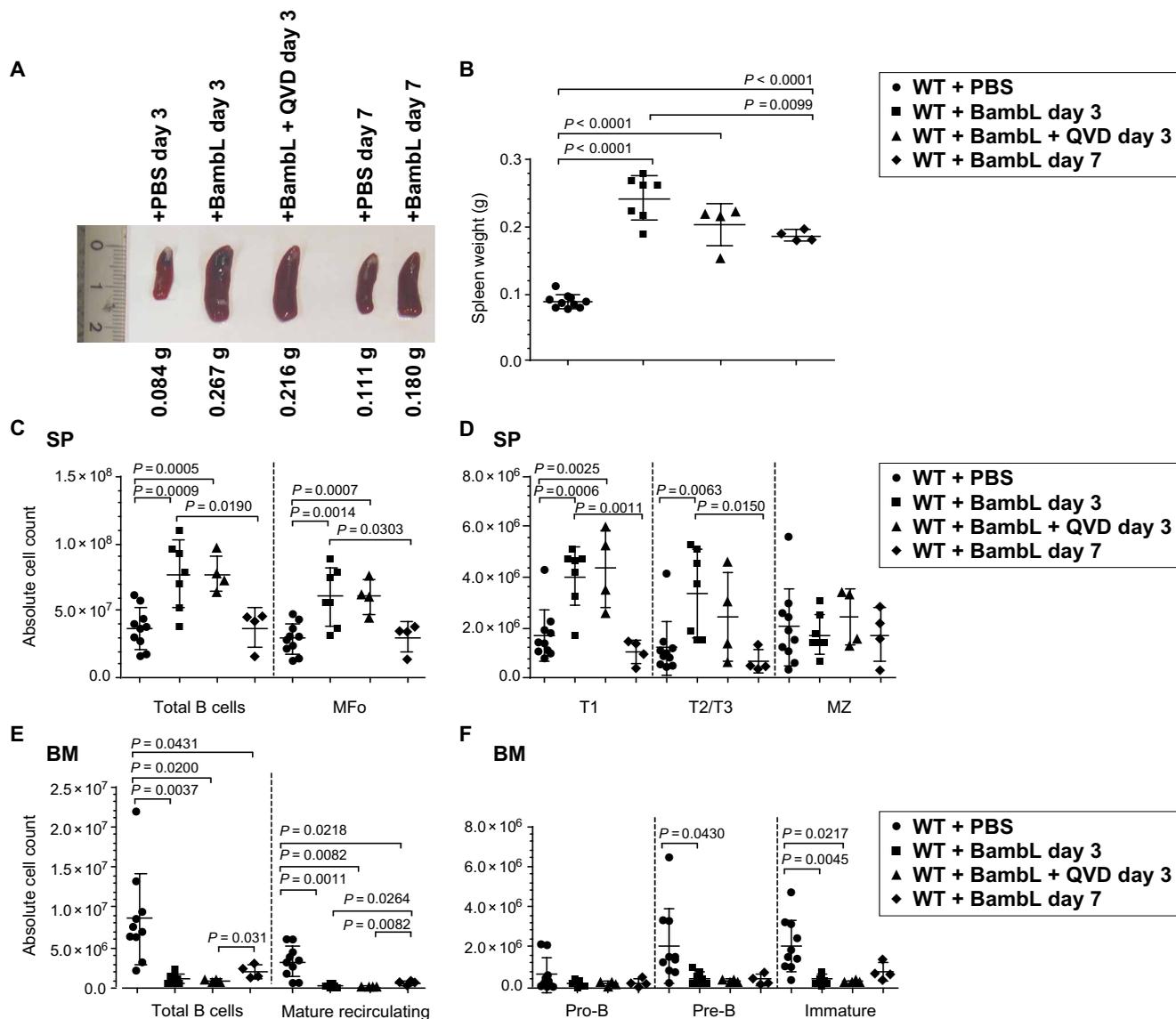


Fig. 7. Intrapерitoneal injection of mice with BambL leads to splenomegaly and accumulation of B cells in the spleen with simultaneous lymphopenia in the bone marrow. (A) Images of spleens for the indicated times and treatment conditions after mice were injected with BambL. The sizes of the spleens were directly compared after extraction. Scale bar is given in centimeters. (B) Statistical analysis of spleen weights from the indicated groups of treated mice. (C and D) Statistical analysis of the numbers of the indicated B cell populations in the indicated groups of treated mice. (E and F) Absolute counts of the indicated B cell populations in the bone marrow of the indicated groups of treated mice. In (B) to (F), each dot indicates an individual mouse: PBS, $n = 10$; BambL d3, $n = 7$; BambL + QVD d3, $n = 4$; BambL d7, $n = 4$. The data represent a compilation of values from four to seven independent experiments. All P values were obtained using a two-tailed Student's t test.

wherein we inactivated BCR expression by conditionally deleting $Ig\alpha$. We used the $Ig\alpha^{TMF}$ mouse model (70), which we crossed with the $mb1-CreER^{T2}$ mice. At day 10 after tamoxifen application, these mice had inactivated $Ig\alpha$ expression and subsequently lost BCR surface expression. We chose this mouse line for the in vivo experiments because the deletion of the $Ig\alpha$ -encoding locus in these mice was more efficient (about 70 to 90% BCR-negative B cells) compared to the inactivation of the HC in the $mb1-CreER^{T2}$; $B1-8^d$ mice (about 15 to 20% BCR-negative B cells), which we had used in our earlier in vitro experiments. Analogously to WT mice, the $mb1-CreER^{T2}$; $Ig\alpha^{TMF}$ mice (referred to as $Ig\alpha^{TMF}$ mice for simplicity) were injected with BambL (15 mg/kg). On day 3 after injection, these mice exhibited

splenomegaly with splenic weights increased twofold in BambL-treated versus PBS-treated animals (fig. S11, A and B). By staining the B cells from the $Ig\alpha^{TMF}$ mice with anti-IgM and anti-IgD antibodies, we confirmed that about 70 to 90% of all B cells were BCR negative (fig. S11C). We further observed an increase in the number of total B cells in the spleen, most likely due to an increased number of MFo B cells, because the transitional and MZ compartments were not substantially affected (fig. S11, D and E). Similar to WT mice, the T cell numbers were not changed (fig. S8B). In the bone marrow, the total cell numbers were not markedly altered, whereas the absolute B cell counts were reduced; here, specifically pro-, pre-, and immature B cell compartments were affected (fig. S11, F and G; for gating strategies for

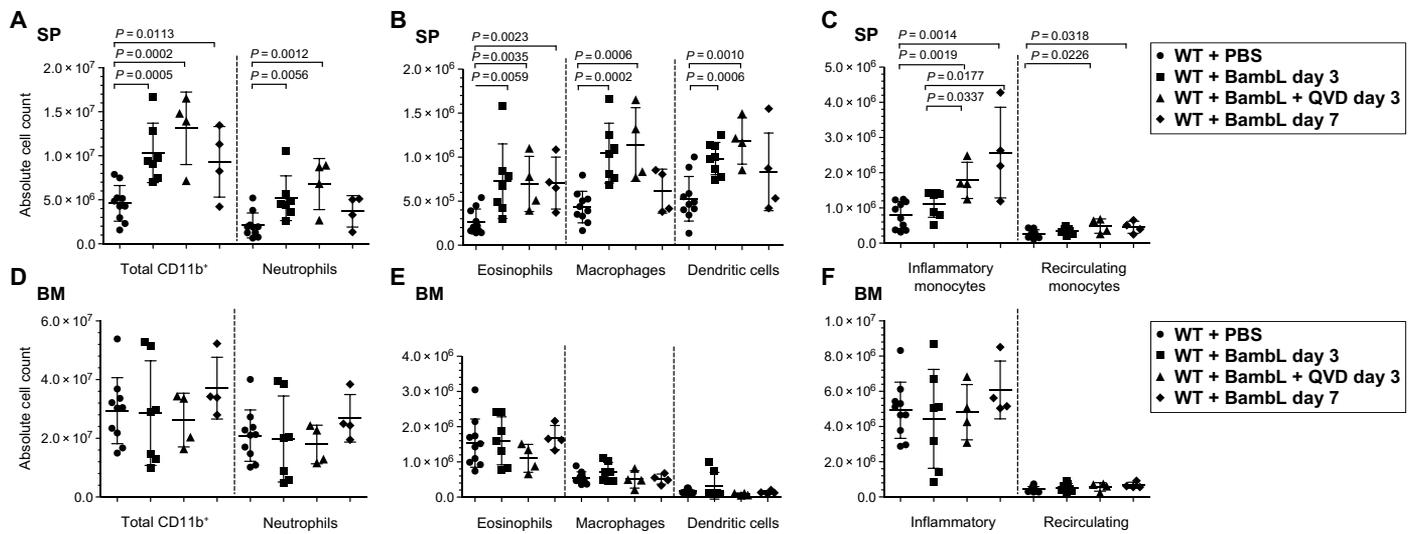


Fig. 8. Myeloid cell counts are increased in mice after injection with BambL. (A to C) Absolute counts of the indicated myeloid cell populations in the spleens of mice from the indicated treatment groups. (D to F) Absolute counts of the indicated myeloid cell populations in the bone marrow of mice from the indicated treatment groups. In (A) to (F), each dot indicates an individual mouse: PBS, $n = 10$; BambL d3, $n = 7$; BambL + QVD d3, $n = 4$; BambL d7, $n = 4$. The data represent a compilation of values from four to seven independent experiments. All P values were obtained using a two-tailed Student's t test. Gating strategy was performed as described previously (67, 68).

splenic and bone marrow cells, see fig. S11, H and I, respectively). In addition, myeloid cells in the spleen (fig. S11, J to L), as well as myeloid compartments in the bone marrow were not substantially changed (fig. S11, M to O).

To investigate the origin of the B cell expansion in the spleen, we blocked interleukin-7 receptor (IL-7R) signaling by injecting the mice with anti-IL-7R antibodies 7 days before we administered BambL, thereby arresting B cell development in the bone marrow. On day 3 after BambL injection (thus, day 10 after anti-IL-7R application), we observed splenomegaly (fig. S12, A and B) and increased numbers of total B cells, most likely due to an expansion of the MFO B cell compartment (fig. S12C). Whereas T1 B cell numbers in the spleen were statistically significantly increased in anti-IL-7R- and BambL-treated mice (fig. S12, D and E), their average numbers were 20-fold lower compared to those of BambL-treated WT mice, which had not received anti-IL-7R antibodies (Fig. 7D). Consistent with this finding, the T2/T3 population was also reduced in numbers; however, it was not further affected by BambL treatment (fig. S12, D and E).

In the bone marrow of anti-IL-7R-treated mice, the total B cell numbers were about 4.5-fold less than those of untreated WT mice (Fig. 7E and fig. S12F). Furthermore, lymphopenia was observed after BambL administration, wherein the total B cell counts and the numbers of mature recirculating B cells in the bone marrow were reduced (fig. S12, F to H). The numbers of pro-, pre-, and immature B cells were markedly reduced because of anti-IL-7R treatment but were not further changed by BambL application (fig. S12, G and H). Last, we detected an increase in the number of neutrophils in the spleens and bone marrow of anti-IL-7R- and BambL-treated mice, whereas other myeloid lineage cells were not substantially affected (fig. S12, I to N). These results suggest that the BambL-induced accumulation of B cells in the spleen was most likely due to an increase in the number of MFO cells rather than a shift in lymphopoiesis from

the bone marrow to the spleen. Furthermore, the lack of the mature recirculating population in the bone marrow demonstrates that mature B cells did not circulate but remained in the spleen.

BambL is localized in the cytosol of *B. ambifaria*

The cellular localization of BambL in *B. ambifaria* has not yet been investigated. We therefore assessed the subcellular localization of BambL in *B. ambifaria* biofilms on agar plates and found that the lectin was not released but was exclusively retained in the cytosol of the bacteria (fig. S13).

DISCUSSION

Pathogens express adhesins and toxins to aid in infection and the colonization of host tissues. Glycan-binding factors, such as the influenza A hemagglutinin, were identified by their ability to aggregate red blood cells (71, 72), whereas toxins, such as Shiga toxin, were shown to use glycolipids on epithelial cells as docking sites for entry (11, 73). Here, we provide insights into the interaction between fucose-binding bacterial lectins and B cells and demonstrate that the lectins LecB and BambL trigger BCR-dependent activation in vitro, ultimately resulting in activation-induced death of B cells. This signaling cascade depended mainly on CD19 and IgD because CD19- and IgD-deficient B cells failed to mobilize intracellular Ca^{2+} upon exposure to BambL. To a lesser extent, BambL also bound to IgM, as seen in the experiments using high-lectin concentrations, wherein preference for IgD was lost. Note that studies on monovalent versus multivalent BCR antigens suggest that a strong burst of Ca^{2+} release only takes place after stimulation with soluble multivalent antigens (30, 74, 75). Interesting in this context is the finding that the hinge region of IgD is more flexible compared to the more rigid IgM-BCR, rendering the IgD molecule better suited to bind to multivalent antigens (30, 33). Consistent with our experiments are reports showing that BCR

signaling upon multivalent antigen triggering relies, to a large extent, on the function of IgD and CD19 and less on IgM (33). Whereas antigen binding involves the complementarity determining region (CDR) loops of the variable domains of the HC and LC, BamBL most likely binds to glycan residues at the constant and hinge regions of the BCR, indicating a difference in mechanism, despite similarities in target receptor affinities. In addition, the signaling of the chemokine receptor CXCR4 engages BCR cross-talk by activating the actin cytoskeleton and also depends on IgD and CD19 expression (76). Both IgM and IgD possess five N-glycosylation sites at each constant region, which differ in their positions. The influence of carbohydrates on the regulation and activation of the BCR is not well understood at present. However, a study by Chen *et al.* demonstrated that the glycoproteins that surround IgD- and IgM-BCRs strongly differ and that this might explain their divergent signaling behaviors (77). BamBL-induced activation resembles that induced by B cell superantigens, such as the microbial toxins *Staphylococcus* protein A or *Peptostreptococcus* protein L, which bind to conserved regions of the BCR by protein-protein interactions (78, 79) but are glycosylation independent.

In addition to Ca²⁺ release, short-term stimulation of B cells with BamBL initiated increased cell surface CD86 expression and the activation of effector functions, such as the secretion of TNF- α . In canonical B cell activation, the surface expression of CD86 is increased to enable T cell costimulation by CD28. We found that long-term stimulation with BamBL overnight again decreased CD86 cell surface expression and caused cell death. The loss of surface CD86 might be explained by a direct interaction with BamBL: If CD86 is fucosylated, it will likely bind to BamBL after prolonged exposure with the lectin, which could lead to its internalization.

After initial B cell activation, BamBL induced B cell death, whereas cotreatment with soluble BAFF, which activates pro-survival signals of the noncanonical NF- κ B and PI3K pathways, partially protected the lectin-treated cells (28, 80, 81). In addition, the polycaspase inhibitor QVD, which blocks caspases-1, -3, -8, -9, and -12, the major players during apoptosis (82), and the necroptosis inhibitor Nec-1 rescued B cells from BamBL-induced cell death in vitro.

To address the role of BamBL in vivo, we injected BamBL intraperitoneally into WT and I α ^{TMF} mice. We observed a transient increase in B cell numbers in the spleen, as well as splenomegaly in WT and I α ^{TMF} mice, which lack the surface BCR, suggesting that the effect of BamBL on B cells in vivo was independent of BCR signaling but might result from stimulation of other B cell surface receptors (Fig. 7 and fig. S11). This phenotype is reminiscent of reports ascribing splenomegaly to various infections, injections with BCR-targeting antibodies, and the application of excess BAFF (83–85). The increase in splenic B cell numbers was accompanied by a loss of B cells in the bone marrow (Fig. 7). To reconcile the lymphopenia in the bone marrow with the simultaneous accumulation of B cells in the spleen 3 days after BamBL application, we first assumed that BamBL triggered an activation-induced migration of B cells from the bone marrow to the spleen. However, blocking B lymphopoiesis in the bone marrow with anti-IL-7R antibodies revealed that the accumulation of mature B cells most likely occurred directly in the spleen due to the hyperproliferation of MFO cells (fig. S12). The loss of B cells in the bone marrow was likely due to BamBL-induced enhanced apoptosis at the pro-B cell stage (fig. S10). Note that the B cell populations in the bone marrow were in the process of recovering, and the spleen sizes and weights, as well as splenic B cell numbers, decreased to almost normal levels by day 7 (Fig. 7). This is, to some

extent, consistent with our in vitro observation that BamBL triggered the activation-induced death of B cells (Fig. 6). However, although we did not detect enhanced apoptosis of splenic B cells in vivo, we cannot exclude the possibility that BamBL induced other cell death mechanisms. A study identified a previously uncharacterized proliferation-limiting mechanism: a DNA damage response-induced cell cycle arrest upon the initial hyperproliferation of Epstein-Barr virus- or CpG-stimulated B cells. This cell cycle arrest mechanism inhibits proliferation without inducing apoptosis (86). Another study demonstrated that B cells transiently activated with antigen return to a quiescent state in the absence of T cell help (87). It is feasible that similar mechanisms control the initial hyperproliferation and subsequent proliferation arrest upon treatment with BamBL, particularly because we did not observe any changes in the numbers of T cell populations after BamBL injection, indicating a lack of enhanced T cell activity (fig. S8B).

We also observed a transient accumulation of myeloid lineage cells in the spleens of BamBL-treated WT mice, including neutrophils, eosinophils, macrophages, and DCs (Fig. 8). This indicates that BamBL targets not only B cells in vivo but also other cell types carrying fucose-bearing receptors. It is likely that the temporary increase in the numbers of these cells further contributes to splenomegaly. In addition, myeloid lineage cells produce B cell stimulatory factors, including BAFF (28, 88). Because BAFF dampened BamBL-induced B cell death in vitro (Fig. 6), increased amounts of BAFF, secreted by activated myeloid cells, might account for the BamBL-mediated proliferation of B cells and protection from BamBL-induced B cell death in vivo. Note that because caspase activity was enhanced in splenic non-B cell populations, we assume that BamBL induced a temporary increase in the number of myeloid cells with subsequent apoptosis, thereby reducing their numbers until day 7 (Fig. S10). Because we partially reversed BamBL-induced B cell death with the polycaspase inhibitor QVD in vitro, we coinjected this inhibitor into the BamBL-treated mice but observed no substantial effect. The activity of caspases has been described to play a role not only in the induction of cell death but also during the differentiation of immune cells (82). In our experimental setup, we only detected caspase activity in non-B cells and pro-B cells.

B. ambifaria is a member of the *B. cepacia* complex (Bcc). *Burkholderia* species comprising the Bcc are often found in patients with CF, and they release multiple lectins, which have not been investigated to date (4, 89). It is likely that these bacteria share common features. For example, BamBL from *B. ambifaria* shares a high protein sequence similarity with other lectins from *B. cepacia* (99%), *Burkholderia pyrocinia* (95%), and *Burkholderia contaminans* (94%) [according to a UniProt-based Basic Local Alignment Search Tool (BLAST) analysis]. The expression dynamics and cellular localization of BamBL in *B. ambifaria* are poorly studied to date. Previous studies reported that LecB from the *B. ambifaria*-related bacterium *P. aeruginosa* was located in the outer surface of the bacterial cell wall (10, 90). We found BamBL exclusively in the cytosol of *B. ambifaria* extracts. This finding is expected because other pathogenic bacteria withhold their soluble lectins; for example, the lectin Shiga toxin B subunit is limited to the cytoplasm and periplasm (91) and is released by a triggered, phage-mediated bacterial lysis (11, 92, 93). Similarly, *B. ambifaria* may release BamBL in response to specific environmental changes that are yet to be discovered.

Another study has correlated the expression of LecA and LecB from *P. aeruginosa* with its pathogenicity, wherein bacterial cytotoxicity

and adhesion were reduced in mutant LecA and LecB strains in vitro and the bacterial burden and dissemination of the mutant strains were decreased in vivo (94). Moreover, our group previously reported that the invasiveness of *P. aeruginosa* depended directly on the expression of LecA (12). The role of BambL in infection in general and in CF in particular remains elusive, and the site of B cell encounter with BambL in vivo is not known to date. The lymph nodes may not be the only sites where B cells encounter their antigens. Direct interaction of B cells with bacterial proteins in the periphery has been previously described (95, 96). For example, Chen *et al.* demonstrated the existence of B cells bearing IgD alone in the mucosa of the upper respiratory tract that become class-switched from IgM in a noncanonical manner of B cell maturation (97). It is possible that the binding of BambL to soluble fucosylated proteins, such as antibodies, may dampen its effectiveness in vivo. However, a study demonstrated that circulating IgG molecules do not inhibit the function of *Staphylococcus aureus* superantigen SpA in vivo but rather potentiate its effects (98). We are aware that SpA binding to B cells is glycosylation independent; therefore, we cannot directly compare it to the binding of BambL to fucosylated epitopes. However, we cannot exclude the possibility that similar mechanisms occur during an infection with *B. ambifaria* or after injection with purified BambL.

It is important to emphasize that all of the observed changes in cellular physiology are likely due to the ability of the lectin to bind to fucosylated host cell receptors. Although BambL may bind to various targets in vivo, our observations suggest that the effect on mature BCR signaling through its glycosylated structures is underestimated. For the murine pre-BCR, core fucosylation has been reported to be indispensable for functional HC expression (99). The increase in fucosylation of surface receptors on epithelial cells protects against many intestinal bacterial infections (100) but increases the susceptibility to other (for example, viral) pathogens (101). However, a direct link between fucosylation and host-pathogen signaling has been difficult to prove, partly due to the strong polymorphism of fucosyltransferases (102). Fucosylated epitopes have been identified as functional receptors for the B subunit of cholera toxin, which binds specifically to GM1 and is often used as a lipid raft marker (13). Furthermore, a viral lectin, the hemagglutinin of influenza A virus, has been classified as a pan-B cell activator through its binding to the sialic acid residues of a reconstituted BCR (103), resulting in the hypothesis that this carbohydrate-dependent and superantigen-like B cell activation mechanism might be evolutionarily conserved; however, bacterial lectins have not been included in the list of typical danger- or pathogen-associated molecular patterns so far.

We propose a mechanism of B cell activation by bacteria. In vitro, BambL specifically binds to the fucose residues of BCRs. This interaction initiates diverse, crucial B cell signaling processes, which depend on the presence of the surface receptors IgD and CD19, and eventually cause B cell death. In vivo, BambL application caused a rapid cellular immune response, which involved not only B cells, but also innate immune cells, and subsided within a week. This might pose a risk to immunocompromised patients unable to mount an effective response to pathogenic particles. Whether our finding can be translated to a natural infection remains to be elucidated. However, this study opens up the possibility for further in-depth research of host-pathogen interactions and reveals that highly regulated mechanisms of B cell signaling and complex BCR expression programs can be hijacked by glycan-targeting bacterial lectins.

MATERIALS AND METHODS

Production of recombinant bacterial lectins

BambL (UniProt-ID: Q0B4G1_BURCM) and LecB (UniProt-ID: Q9HYN5_PSEAE) were expressed in *Escherichia coli* and purified by affinity chromatography, as previously described (8, 9). The freeze-dried powder was dissolved in PBS without Ca^{2+} or Mg^{2+} at 1 mg/ml. To avoid contamination by LPS, the lectins were further purified on endotoxin removal traps (hyglos), sterile-handled, and regularly checked for LPS contamination with the Pierce Limulus Amebocyte Lysate (LAL) chromogenic assay (Thermo Scientific). Activation experiments (measurement of Ca^{2+} release and increased CD86 expression) included additional controls with boiled BambL (15 min at 95°C) to exclude possible effects of heat-stable LPS contaminants originating from the recombinant protein production. For fluorescent labeling, the purified lectins were covalently tagged with the Alexa Fluor 488 (Thermo Scientific), Cy-3, or Cy-5 (GE Healthcare) labeling kits according to the manufacturers' instructions and repurified on a Pierce ZebaSpin D-Salt polyacrylamide desalting column (Thermo Scientific).

Mice

All animal experiments were conducted with mice aged 8 to 12 weeks, were performed in accordance with the German Animal Welfare Act, and were reviewed by the regional council and approved under license number G-09/103. Mice were bred in the animal facility of the Max Planck Institute of Immunology and Epigenetics (Freiburg, Germany) and the University of Ulm (Ulm, Germany). For inducible, B cell-specific deletion, mice were orally administered tamoxifen citrate (Ratiopharm) dissolved in 20% ClinOleic (Baxter) as previously described (80). The course of tamoxifen treatments and the interval between doses were pre-optimized for maximal recovery of the modified cells and are outlined below (81). In this study, the following mice strains were used: C57BL/6 WT or heterozygous mEGFP/mb1^{inv} (104) (GFP⁺ B cells); B cells from IgM^{-/-} (47) and IgD^{-/-} mice were also used (105). CD19^{-/-} B cells were obtained from homozygous CD19-CreER^{T2} mice (provided by M. Schmidt-Suppran; Technical University Munich, Germany) (81, 106). To generate CD19/IgD double-deficient B cells, homozygous CD19-CreER^{T2} mice were crossed with homozygous IgD^{-/-} mice and then intercrossed. HC^{-/-} B cells were generated by crossing HC^{B1-8fl/jhT} mice (44) with mb1-CreER^{T2} mice and were analyzed 7 days after tamoxifen application. For the BambL injection experiments, Igα^{TMF} mice (70) were crossed with mb1-CreER^{T2} mice; their offspring are termed Igα^{TMF} mice in this manuscript for simplicity. The mice were injected with BambL at day 7 after tamoxifen treatment, and cells were extracted and analyzed 3 days later. Igα^{-/-} B cells were generated by crossing mb1/mEGFP^{inv} (104) mice with mb1-CreER^{T2} mice (45, 81) and were obtained on day 10 after tamoxifen treatment. Furthermore, we used Lyn^{-/-} mice (107), mice with B cell-specific conditional Syk deletion (Syk^{-/-}) (81), and Blimp^{GFP} mice (49). The animals were euthanized using CO₂, and the spleens and bone marrow were extracted under nonsterile conditions. B cells were then purified by magnetic or flow cytometry-based separation.

Isolation of murine B cells by magnetic-activated cell sorting depletion and flow cytometry-based sorting

Cell separation was performed by magnetic-activated cell sorting (MACS) depletion or flow cytometry-based sorting; spleen-derived B cells were obtained by MACS-based negative selection using the B cell isolation kit (Miltenyi Biotec) according to the manufacturer's instructions. The cells were then separated using an AutoMACS

(Miltenyi Biotec). Spleen-derived B cells from heterozygous mEGFP/*mb1^{intv}* mice as well as GFP⁺ B cells from *Igα^{-/-}* mice were separated by fluorescence-activated cell sorting (FACS) based on their abundance of GFP with a FACSAria (BD) cell sorter. HC-deficient B cells were sorted based on the lack of IgM and IgD expression, as described elsewhere (45). After separation, B cells were analyzed for purity and viability with an anti-CD19 antibody (6D5, BioLegend) and 7-AAD (BioLegend) or a fixable viability dye (eBioscience).

Flow cytometric analysis

Extracellular staining for flow cytometric (FACS) analysis was performed with 0.5×10^6 to 1×10^6 cells/ml in FACS buffer [PBS; 3% fetal calf serum (FCS) and 0.05% NaN_3] on ice for 20 min. Before staining, the cells were incubated with the anti-Fc receptor-blocking antibody (CD16/32, Clone 2.4G2; own production) on ice for at least 5 min. For the extracellular staining of B cells, the following antibodies were used: anti-CD19 Alexa Fluor 647 or phycoerythrin (PE) (eBio1D3; eBioscience and 6D5; BioLegend), anti-IgD fluorescein isothiocyanate (FITC) or PE (11-26c.2a; BioLegend), anti-CD23 Alexa Fluor 647 or PE-Cy7 (B3B4; BioLegend or 2G8; Southern Biotech), anti-CD24 (M1/69; BioLegend), anti-CD25 (PC61.5, eBioscience), anti-BAFF-R biotin (polyclonal; R&D Systems), anti-IgM eFluor 450 (eB121-15F9), anti-CD93 PE or allophycocyanin (APC) (AA4.1), and anti-CD86 PE (GL1) (all eBioscience). For the myeloid stainings, we used anti-CD11c (N418, BioLegend), anti-F4/80 (BM8, eBioscience), anti-CD11b (M1/70; eBioscience), anti-Ly6C (HK1.4; eBioscience), and anti-CD3e (145-2C11, eBioscience). The cells were washed with 700 μl of FACS buffer to remove the residual antibodies and centrifuged at 300g at 4°C for 5 min. All antibodies used in this study were tested for lectin cross-reactivity before staining. CyAn Advanced Digital Processing (ADP) (Beckman Coulter), LSRII, and LSRFortessa (both from Becton Dickinson) cell analyzers were used for data acquisition. For intracellular staining, cells were fixed and permeabilized using the Fix & Perm Kit (ADG Bio Research) and stained with anti-TNF- α antibody (clone: MP6-XT22; eBioscience) dissolved in saponin-based buffer. Cytometry data were analyzed with Flow Jo software (TreeStar).

Caspase-activity assay

Cells were stained with the SR-FLICA Caspase Assay Kit (ImmunoChemistry Technologies) according to the manufacturer's instructions. Briefly, the cells were isolated, stained with detection reagent for 1 hour at room temperature, and washed with the specific apoptosis buffer containing PBS, which was provided with the kit.

Ca²⁺ analysis and binding assays

Ca²⁺ flux in purified mature B cells was measured by flow cytometry using the cell-permeable fluorescent dye Indo-1 (Life Technologies) cells loaded as described previously (45). The following stimuli were used: anti-IgM F(ab')₂ (goat polyclonal; Jackson ImmunoResearch), anti-kappa F(ab')₂, and anti-IgD F(ab')₂ (both polyclonal; Southern Biotech). The cells were maintained in culture medium containing 1% FCS and were treated with lectins at different concentrations, ranging from 250 ng/ml to 10 $\mu\text{g}/\text{ml}$. Intracellular Ca²⁺ flux and extracellular binding of BambL-Cy5 were measured simultaneously on an LSRII flow cytometer (Becton Dickinson).

Lectin-dependent increase in CD86 cell surface abundance

B cells were seeded at 1×10^6 cells/ml in complete medium and stimulated in a 96-well U-bottom plate with the lectin dissolved in

complete medium. For the binding assays, B cells were stimulated on ice for 15 min and then analyzed for lectin binding by flow cytometry. For the "wash out" procedure, cells were stimulated with BambL at 37°C for 15 min, which was followed by two washing steps with 2 ml of PBS. As a positive control, we used polyclonal anti-IgM F(ab')₂ (goat polyclonal; Jackson ImmunoResearch, 10 $\mu\text{g}/\text{ml}$), unmethylated CpG oligonucleotides (Invivogen; ODN 1826, 2.5 μM), and LPS (Sigma; 10 $\mu\text{g}/\text{ml}$). The cells were harvested 16 hours after stimulation, and the cell surface abundance of the activation marker CD86 was analyzed by flow cytometry using anti-CD86 (clone, GL1; eBioscience).

Measurement of TNF- α by ELISA

The concentration of secreted TNF- α was measured by ELISA. The extracted splenic B cells were treated with BambL overnight, and the cell culture medium was collected by centrifugation and subsequent filtration. The amount of TNF- α released was determined using the anti-mouse TNF- α Ready-Set-Go ELISA Kit (eBioscience) according to the manufacturer's instructions.

Survival assays

Primary, spleen-derived murine B cells were purified and cultured at 37°C and 7.5% CO₂ in complete medium, Iscove's modified Dulbecco's medium (IMDM) (Biochrom, Merck) supplemented with 10% heat-inactivated FCS, 1% Gibco Penicillin-Streptomycin [penicillin (100 U/ml)-streptomycin (100 $\mu\text{g}/\text{ml}$)], and 2 mM Gibco L-glutamine (all from Thermo Scientific). The cells were incubated either in medium alone or in the presence of human recombinant BAFF (100 ng/ml) (hrBAFF, ImmunoTools) over a period of 4 days. The medium was exchanged every second day and supplemented with fresh hrBAFF. Cell viability was assessed with a fixable viability dye (eBioscience). Annexin V and propidium iodide staining was not applicable for the survival assays because of reported false positivity by catfish egg lectin (108). For the viability analysis, the percentages were obtained from the ungated, viability dye-negative (healthy) cells. The following inhibitors were first titrated on B cells and then used in this study: necrostatin (Nec-1), polycaspase inhibitor QVD (Q-VD-PH), and caspase-1 inhibitor Z-WEHD-FMK (all from R&D Biosystems).

Intraperitoneal injection of BambL

Expecting that a major fraction of BambL would be intercepted by soluble Igs and carbohydrates expressed on erythrocytes, we decided to inject BambL in PBS (without adjuvants) intraperitoneally instead of intravenously. In a dose-dependency study, we determined the concentration of 15 mg/kg to be comparable to the doses of BCR-targeting antibodies that had been used previously (84). Control mice received the same volume of PBS intraperitoneally. Seven days after injection, we extracted the spleens and bone marrow to further analyze the cells by flow cytometry. Experimental mice did not suffer at any point of the intraperitoneal treatment experiments and did not lose any body weight during the study. QVD was coapplied at 20 mg/kg, as suggested by the manufacturer (R&D Biosystems).

BambL localization in *B. ambifaria*

Bacteria were fractionated on the basis of the LecB localization protocol for *P. aeruginosa* published by Tielker *et al.* (10). *B. ambifaria* strain HSJ1 was grown as a biofilm on agar plates [tryptic soy broth (30 g liter⁻¹) and agar (15 g liter⁻¹); Carl Roth] for 72 hours. Bacteria were detached and collected in 1 ml of 140 mM NaCl, which yielded

an optical density of ~160 at 600 nm. A sample (100 μ l) of this suspension was subjected to fractionation, whereas another 100- μ l sample was used to prepare whole-cell lysates. Bacteria were collected by centrifugation (5 min at 3000g), and the supernatant was sterile-filtered before the BamBL concentration in the extracellular space was determined. The bacterial outer membrane was lysed for 30 min at room temperature in 500 μ l of 100 mM tris-HCl (pH 8) containing 20% (w/v) sucrose, 2.5 mM EDTA, and 20 μ g of lysozyme (Sigma-Aldrich). Spheroblasts were collected by centrifugation (20 min at 10,000g); the supernatant served as the periplasmic fraction. The spheroblasts were sonicated (Bandelin Sonopuls mini20, probe MS 2.5) in 500 μ l of 100 mM tris-HCl (pH 8), and undisrupted cells were separated by centrifugation (5 min at 5000g). The supernatant was centrifuged at 20,000g for 45 min to collect the membrane fraction; the resulting supernatant was used as the cytoplasmic fraction. For Western blotting analysis, equivalent volumes of each cellular fraction were used to directly compare the relative amounts of BamBL. Recombinantly produced BamBL was loaded as a positive control. Samples were boiled with SDS-polyacrylamide gel electrophoresis sample buffer and separated in an 18% polyacrylamide gel, followed by electrophoretic transfer to a nitrocellulose membrane. After transfer, proteins were stained reversibly with Ponceau S solution (Sigma-Aldrich) and photographed. BamBL was detected with a polyclonal antibody raised against its C terminus (Eurogentec) and an anti-rabbit IgG-horseradish peroxidase conjugate using the Clarity ECL kit (Bio-Rad).

Software and statistical analysis

Adobe Illustrator CC 2017 (Adobe) and Prism7 (GraphPad) software were used for figure preparation and statistical analysis, respectively. The statistical significance of the observed differences was evaluated by an unpaired, two-tailed Student's *t* test (if not indicated otherwise). **P* < 0.05, ***P* < 0.01, ****P* < 0.001, *****P* < 0.0001; ns, not significant. In the *in vivo* experiments, the exact *P* values are indicated; differences that were not statistically significantly different are not depicted.

SUPPLEMENTARY MATERIALS

www.sciencesignaling.org/cgi/content/full/12/571/eaao7194/DC1

- Fig. S1. Lectin binding and gating of HC^{-/-} B cells.
 Fig. S2. Gating schemes and IgD surface expression of specific B cell populations.
 Fig. S3. Characterization of CD19-, IgM-, and IgD-deficient B cells.
 Fig. S4. B220 is not internalized upon BamBL binding.
 Fig. S5. Internalization of CD86 and intracellular increase in TNF- α in BamBL-treated primary B cells.
 Fig. S6. Stimulation of cells with BamBL in the presence of cell death inhibitors.
 Fig. S7. BamBL binds to soluble Igs.
 Fig. S8. Analysis of lymphoid lineage cells after *in vivo* injection of BamBL.
 Fig. S9. Analysis of myeloid lineage cells after *in vivo* injection of BamBL.
 Fig. S10. Non-B cells in the spleen show an increase in caspase activity upon BamBL injection.
 Fig. S11. BamBL injection leads to splenomegaly and B cell accumulation in Ig α ^{TMF} mice.
 Fig. S12. Mice treated with anti-IL-7R antibody exhibit splenomegaly and B cell proliferation in the spleen.
 Fig. S13. *B. ambifaria* biofilms do not secrete BamBL but retain it in the cytosol.

REFERENCES AND NOTES

1. T. Coenye, E. Mahenthalingam, D. Henry, J. J. LiPuma, S. Laevens, M. Gillis, D. P. Speert, P. Vandamme, *Burkholderia ambifaria* sp. nov., a novel member of the *Burkholderia cepacia* complex including biocontrol and cystic fibrosis-related isolates. *Int. J. Syst. Evol. Microbiol.* **51**, 1481–1490 (2001).
2. P. D. Lister, D. J. Wolter, N. D. Hanson, Antibacterial-resistant *Pseudomonas aeruginosa*: Clinical impact and complex regulation of chromosomally encoded resistance mechanisms. *Clin. Microbiol. Rev.* **22**, 582–610 (2009).
3. T. Coenye, P. Vandamme, Diversity and significance of *Burkholderia* species occupying diverse ecological niches. *Environ. Microbiol.* **5**, 719–729 (2003).
4. M. S. Saldias, M. A. Valvano, Interactions of *Burkholderia cenocepacia* and other *Burkholderia cepacia* complex bacteria with epithelial and phagocytic cells. *Microbiology* **155**, 2809–2817 (2009).
5. K. A. Ramsay, C. A. Butler, S. Paynter, R. S. Ware, T. J. Kidd, C. E. Wainwright, S. C. Bell, Factors influencing acquisition of *Burkholderia cepacia* complex organisms in patients with cystic fibrosis. *J. Clin. Microbiol.* **51**, 3975–3980 (2013).
6. G. Pradenas, B. N. Ross, A. G. Torres, *Burkholderia cepacia* complex vaccines: Where do we go from here? *Vaccine* **4**, 10 (2016).
7. E. C. Nannini, A. Ponessa, R. Muratori, P. Marchiaro, V. Ballerini, L. Flynn, A. S. Limansky, Polyclonal outbreak of bacteremia caused by *Burkholderia cepacia* complex and the presumptive role of ultrasound gel. *Braz. J. Infect. Dis.* **19**, 543–545 (2015).
8. A. Imberty, M. Wimmerova, E. P. Mitchell, N. Gilboa-Garber, Structures of the lectins from *Pseudomonas aeruginosa*: Insights into the molecular basis for host glycan recognition. *Microbes Infect.* **6**, 221–228 (2004).
9. A. Audfray, J. Claudinon, S. Abounit, N. Ruvoën-Clouet, G. Larson, D. F. Smith, M. Wimmerová, J. Le Pendu, W. Römer, A. Varrot, A. Imberty, Fucose-binding lectin from opportunistic pathogen *Burkholderia ambifaria* binds to both plant and human oligosaccharidic epitopes. *J. Biol. Chem.* **287**, 4335–4347 (2012).
10. D. Tielker, S. Hacker, R. Loris, M. Strathmann, J. Wingender, S. Wilhelm, F. Rosenau, K.-E. Jaeger, *Pseudomonas aeruginosa* lectin LecB is located in the outer membrane and is involved in biofilm formation. *Microbiology* **151**, 1313–1323 (2005).
11. L. Johannes, W. Römer, Shiga toxins—From cell biology to biomedical applications. *Nat. Rev. Microbiol.* **8**, 105–116 (2010).
12. T. Eierhoff, B. Bastian, R. Thuenauer, J. Madl, A. Audfray, S. Aigal, S. Juillot, G. E. Rydell, S. Müller, S. de Bentzmann, A. Imberty, C. Fleck, W. Römer, A lipid zipper triggers bacterial invasion. *Proc. Natl. Acad. Sci. U.S.A.* **111**, 12895–12900 (2014).
13. A. M. Wands, A. Fujita, J. E. McCombs, J. Cervin, B. Dedic, A. C. Rodriguez, N. Nischan, M. R. Bond, M. Mettlen, D. C. Trudgian, A. Lemoff, M. Quiding-Järbrink, B. Gustavsson, C. Steentoft, H. Clausen, H. Mirzaei, S. Teneberg, U. Yrlid, J. J. Kohler, Fucosylation and protein glycosylation create functional receptors for cholera toxin. *eLife* **4**, e09545 (2015).
14. C. Cott, R. Thuenauer, A. Landj, K. Kühn, S. Juillot, A. Imberty, J. Madl, T. Eierhoff, W. Römer, *Pseudomonas aeruginosa* lectin LecB inhibits tissue repair processes by triggering β -catenin degradation. *Biochim. Biophys. Acta Mol. Cell Res.* **1863**, 1106–1118 (2016).
15. D. Avichezer, N. Gilboa-Garber, PA-II, the L-fucose, and D-mannose binding lectin of *Pseudomonas aeruginosa* stimulates human peripheral lymphocytes and murine splenocytes. *FEBS Lett.* **216**, 62–66 (1987).
16. N. Sharon, History of lectins: From hemagglutinins to biological recognition molecules. *Glycobiology* **14**, 53R–62R (2004).
17. R. Übelhart, M. P. Bach, C. Eschbach, T. Wossning, M. Reth, H. Jumaa, N-linked glycosylation selectively regulates autonomous precursor BCR function. *Nat. Immunol.* **11**, 759–765 (2010).
18. J. Haimovich, N. Ben Moshe, Y. Raviv, N. Hollander, All oligosaccharide moieties of the μ chains in the pre-BCR are of the high-mannose type. *Mol. Immunol.* **48**, 351–355 (2010).
19. D. Schneider, M. Dühren-von Minden, A. Alkhatib, C. Setz, C. A. M. van Bergen, M. Benkißer-Petersen, I. Wilhelm, S. Villringer, S. Krysov, G. Packham, K. Zirlak, W. Römer, C. Buske, F. K. Stevenson, H. Veelken, H. Jumaa, Lectins from opportunistic bacteria interact with acquired variable-region glycans of surface immunoglobulin in follicular lymphoma. *Blood* **125**, 3287–3296 (2015).
20. W. G. Kerr, L. M. Hendershot, P. D. Burrows, Regulation of IgM and IgD expression in human B-lineage cells. *J. Immunol.* **146**, 3314–3321 (1991).
21. K. Rajewsky, Clonal selection and learning in the antibody system. *Nature* **381**, 751–758 (1996).
22. R. Geisberger, M. Lamers, G. Achatz, The riddle of the dual expression of IgM and IgD. *Immunology* **118**, 429–437 (2006).
23. J. M. Hombach, T. Tsubata, L. Leclercq, H. Stappert, M. G. Reth, Molecular components of the IgM antigen receptor complex. *Prog. Immunol.* 659–662 (1989).
24. A. Weiss, D. R. Littman, Signal transduction by lymphocyte antigen receptors. *Cell* **76**, 263–274 (1994).
25. J. C. Cambier, C. M. Pleiman, M. R. Clark, Signal transduction by the B cell antigen receptor and its coreceptors. *Annu. Rev. Immunol.* **12**, 457–486 (1994).
26. R. A. Barrington, M. Zhang, X. Zhong, H. Jonsson, N. Holodick, A. Cherukuri, S. K. Pierce, T. L. Rothstein, M. C. Carroll, CD21/CD19 coreceptor signaling promotes B cell survival during primary immune responses. *J. Immunol.* **175**, 2859–2867 (2005).
27. D. A. Tuveson, R. H. Carter, S. P. Soltoff, D. T. Fearon, CD19 of B cells as a surrogate kinase insert region to bind phosphatidylinositol 3-kinase. *Science* **260**, 986–989 (1993).
28. P. Schneider, F. Mackay, V. Steiner, K. Hofmann, J.-L. Bodmer, N. Holler, C. Ambrose, P. Lawton, S. Bixler, H. Acha-Orbea, D. Valmori, P. Romero, C. Werner-Favre, R. H. Zubler, J. L. Browning, J. Tschopp, BAFF, a novel ligand of the tumor necrosis factor family, stimulates B cell growth. *J. Exp. Med.* **189**, 1747–1756 (1999).

29. A. Craxton, K. E. Draves, A. Gruppi, E. A. Clark, BAFF regulates B cell survival by downregulating the BH3-only family member Bim via the ERK pathway. *J. Exp. Med.* **202**, 1363–1374 (2005).
30. C. Volkman, N. Brings, M. Becker, E. Hobeika, J. Yang, M. Reth, Molecular requirements of the B cell antigen receptor for sensing monovalent antigens. *EMBO J.* **35**, 2371–2381 (2016).
31. F. D. Batista, D. Iber, M. S. Neuberger, B cells acquire antigen from target cells after synapse formation. *Nature* **411**, 489–494 (2001).
32. E. Hobeika, P. C. Maity, H. Jumaa, Control of B cell responsiveness by isotype and structural elements of the antigen receptor. *Trends Immunol.* **37**, 310–320 (2016).
33. R. Übelhart, E. Hug, M. P. Bach, T. Wosning, M. Dühren-von Minden, A. H. C. Horn, D. Tsiantoulas, K. Kometani, T. Kurosaki, C. J. Binder, H. Sticht, L. Nitschke, M. Reth, H. Jumaa, Responsiveness of B cells is regulated by the hinge region of IgD. *Nat. Immunol.* **16**, 534–543 (2015).
34. J. N. Arnold, M. R. Wormald, R. B. Sim, P. M. Rudd, R. A. Dwek, The impact of glycosylation on the biological function and structure of human immunoglobulins. *Annu. Rev. Immunol.* **25**, 21–50 (2007).
35. E. Maverakis, K. Kim, M. Shimoda, M. E. Gershwin, F. Patel, R. Wilken, S. Raychaudhuri, L. R. Ruhaak, C. B. Lebrilla, Glycans in the immune system and the altered glycan theory of autoimmunity: A critical review. *J. Autoimmun.* **57**, 1–13 (2015).
36. K.-T. C. Shade, R. M. Anthony, Antibody glycosylation and inflammation. *Antibodies* **2**, 392–414 (2013).
37. V. Coelho, S. Krysov, A. M. Ghaemmaghami, M. Emara, K. N. Potter, P. Johnson, G. Packham, L. Martinez-Pomares, F. K. Stevenson, Glycosylation of surface Ig creates a functional bridge between human follicular lymphoma and microenvironmental lectins. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 18587–18592 (2010).
38. F. Vuillier, G. Dumas, C. Magnac, M.-C. Prevost, A. I. Lalanne, P. Oppezio, E. Melanitou, G. Dighiero, B. Payelle-Brogard, Lower levels of surface B cell–receptor expression in chronic lymphocytic leukemia are associated with glycosylation and folding defects of the μ and CD79a chains. *Blood* **105**, 2933–2940 (2005).
39. T. Kurosaki, Functional dissection of BCR signaling pathways. *Curr. Opin. Immunol.* **12**, 276–281 (2000).
40. L. B. King, B. D. Freedman, B-lymphocyte calcium influx. *Immunol. Rev.* **231**, 265–277 (2009).
41. Y. Baba, M. Matsumoto, T. Kurosaki, Calcium signaling in B cells: Regulation of cytosolic Ca^{2+} increase and its sensor molecules, STIM1 and STIM2. *Mol. Immunol.* **62**, 339–343 (2013).
42. C. Lamagna, Y. Hu, A. L. DeFranco, C. A. Lowell, B cell–specific loss of lyn kinase leads to autoimmunity. *J. Immunol.* **192**, 919–928 (2014).
43. V. W. F. Chan, C. A. Lowell, A. L. DeFranco, Defective negative regulation of antigen receptor signaling in Lyn-deficient B lymphocytes. *Curr. Biol.* **8**, 545–553 (1998).
44. K.-P. Lam, R. Kühn, K. Rajewsky, In vivo ablation of surface immunoglobulin on mature B cells by inducible gene targeting results in rapid cell death. *Cell* **90**, 1073–1083 (1997).
45. E. Levit-Zerdoun, M. Becker, R. Pohlmeier, I. Wilhelm, P. C. Maity, K. Rajewsky, M. Reth, E. Hobeika, Survival of Ig α -deficient mature B cells requires BAFF-R function. *J. Immunol.* **196**, 2348–2360 (2016).
46. M. Kraus, L. I. Pao, A. Reichlin, Y. Hu, B. Canono, J. C. Cambier, M. C. Nussenzweig, K. Rajewsky, Interference with immunoglobulin (Ig) α immunoreceptor tyrosine–based activation motif (ITAM) phosphorylation modulates or blocks B cell development, depending on the availability of an Ig β cytoplasmic tail. *J. Exp. Med.* **194**, 455–470 (2001).
47. C. Lutz, B. Ledermann, M. H. Kosco-Vilbois, A. F. Ochsenbein, R. M. Zinkernagel, G. Köhler, F. Brombacher, IgD can largely substitute for loss of IgM function in B cells. *Nature* **393**, 797–801 (1998).
48. J. G. Monroe, W. L. Havran, J. C. Cambier, B lymphocyte activation: Entry into cell cycle is accompanied by decreased expression of IgD but not IgM. *Eur. J. Immunol.* **13**, 208–213 (1983).
49. K. A. Fairfax, L. M. Corcoran, C. Pridans, N. D. Huntington, A. Kallies, S. L. Nutt, D. M. Tarlinton, Different kinetics of Blimp-1 induction in B cell subsets revealed by reporter gene. *J. Immunol.* **178**, 4104–4111 (2007).
50. A. Kallies, J. Hasbold, D. M. Tarlinton, W. Dietrich, L. M. Corcoran, P. D. Hodgkin, S. L. Nutt, Plasma cell ontogeny defined by quantitative changes in Blimp-1 expression. *J. Exp. Med.* **200**, 967–977 (2004).
51. P. C. Maity, A. Blount, H. Jumaa, O. Ronneberger, B. F. Lillemeier, M. Reth, B cell antigen receptors of the IgM and IgD classes are clustered in different protein islands that are altered during B cell activation. *Sci. Signal.* **8**, ra93 (2015).
52. K. Kläsener, P. C. Maity, E. Hobeika, J. Yang, M. Reth, B cell activation involves nanoscale receptor reorganizations and inside-out signaling by Syk. *eLife* **3**, e02069 (2014).
53. P. Hou, E. Araujo, T. Zhao, M. Zhang, D. Massenbourg, M. Veselits, C. Doyle, A. R. Dinner, M. R. Clark, B cell antigen receptor signaling and internalization are mutually exclusive events. *PLoS Biol.* **4**, e200 (2006).
54. P. K. A. Mongini, S. Tolani, R. J. Fattah, J. K. Inman, Antigen receptor triggered upregulation of CD86 and CD80 in human B cells: Augmenting role of the CD21/CD19 co-stimulatory complex and IL-4. *Cell. Immunol.* **216**, 50–64 (2002).
55. D. Frasca, M. Romero, A. Diaz, S. Alter-Wolf, M. Ratliff, A. M. Landin, R. L. Riley, B. B. Blomberg, A molecular mechanism for TNF- α -mediated downregulation of B cell responses. *J. Immunol.* **188**, 279–286 (2012).
56. R. C. Rickert, J. Jellusova, A. V. Miletic, Signaling by the tumor necrosis factor receptor superfamily in B cell biology and disease. *Immunol. Rev.* **244**, 115–133 (2011).
57. C. J. Hanson, M. D. Bootman, H. L. Roderick, Cell signalling: IP3 receptors channel calcium into cell death. *Curr. Biol.* **14**, R933–R935 (2004).
58. A. M. Scharenberg, L. A. Humphries, D. J. Rawlings, Calcium signalling and cell-fate choice in B cells. *Nat. Rev. Immunol.* **7**, 778–789 (2007).
59. D. Donjerković, D. W. Scott, Activation-induced cell death in B lymphocytes. *Cell Res.* **10**, 179–192 (2000).
60. I. Ceballos-Olvera, M. Sahoo, M. A. Miller, L. del Barrio, F. Re, Inflammasome-dependent pyroptosis and IL-18 protect against *Burkholderia pseudomallei* lung infection while IL-1 β is deleterious. *PLoS Pathog* **7**, e1002452 (2011).
61. R. Rosales-Reyes, A. Pérez-López, C. Sánchez-Gómez, R. R. Hernández-Mote, D. Castro-Eguiluz, V. Ortiz-Navarrete, C. M. Alpuche-Aranda, *Salmonella* infects B cells by macropinocytosis and formation of spacious phagosomes but does not induce pyroptosis in favor of its survival. *Microb. Pathog.* **52**, 367–374 (2012).
62. L. Schoenlaub, R. Cherla, Y. Zhang, G. Zhang, *Coxiella burnetii* avirulent nine mile phase II induces Caspase-1–dependent pyroptosis in murine peritoneal B1a B cells. *Infect. Immun.* **84**, 3638–3654 (2016).
63. V. A. K. Rathinam, K. A. Fitzgerald, Inflammasome complexes: Emerging mechanisms and effector functions. *Cell* **165**, 792–800 (2016).
64. V. Y. Melnikov, S. Faubel, B. Siegmund, M. S. Lucia, D. Ljubanovic, C. L. Edelstein, Neutrophil-independent mechanisms of caspase-1– and IL-18–mediated ischemic acute tubular necrosis in mice. *J. Clin. Invest.* **110**, 1083–1091 (2002).
65. T. M. Caserta, A. N. Smith, A. D. Gultice, M. A. Reedy, T. L. Brown, Q-VD-OPH, a broad spectrum caspase inhibitor with potent antiapoptotic properties. *Apoptosis* **8**, 345–352 (2003).
66. B. Boulianne, O. L. Rojas, D. Haddad, A. Zahaen, A. Kapelnikov, T. Nguyen, C. Li, R. Hakem, J. L. Gommerman, A. Martin, AID and caspase 8 shape the germinal center response through apoptosis. *J. Immunol.* **191**, 5840–5847 (2013).
67. P. Lee, Z. Zhu, J. Hachmann, T. Nojima, D. Kitamura, G. Salvesen, R. C. Rickert, Differing requirements for MALT1 function in peripheral B cell survival and differentiation. *J. Immunol.* **198**, 1066–1080 (2017).
68. W. P. Lafuse, R. Gearinger, S. Fisher, C. Nealer, A. R. Mackos, M. T. Bailey, Exposure to a social stressor induces translocation of commensal lactobacilli to the spleen and priming of the innate immune system. *J. Immunol.* **198**, 2383–2393 (2017).
69. S. Rose, A. Misharin, H. Perlman, A novel Ly6C/Ly6G-based strategy to analyze the mouse splenic myeloid compartment. *Cytometry* **81A**, 343–350 (2012).
70. M. Kraus, M. B. Alimzhanov, N. Rajewsky, K. Rajewsky, Survival of resting mature B lymphocytes depends on BCR signaling via the Ig α / β heterodimer. *Cell* **117**, 787–800 (2004).
71. G. K. Hirst, The agglutination of red cells by allantoic fluid of chick embryos infected with influenza virus. *Science* **94**, 22–23 (1941).
72. A. Gottschalk, S. M. Patridge, Interaction between simple sugars and amino-acids. *Nature* **165**, 684–685 (1950).
73. T. Schubert, W. Römer, How synthetic membrane systems contribute to the understanding of lipid-driven endocytosis. *Biochim. Biophys. Acta Mol. Cell Res.* **1853**, 2992–3005 (2015).
74. Y.-M. Kim, J. Y.-J. Pan, G. A. Korbel, V. Peperzak, M. Boes, H. L. Ploegh, Monovalent ligation of the B cell receptor induces receptor activation but fails to promote antigen presentation. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 3327–3332 (2006).
75. S. Mukherjee, J. Zhu, J. Zikherman, R. Parameswaran, T. A. Kadlecik, Q. Wang, B. Au-Yeung, H. Ploegh, J. Kuriyan, J. Das, A. Weiss, Monovalent and multivalent ligation of the B cell receptor exhibit differential dependence upon Syk and Src family kinases. *Sci. Signal.* **6**, ra1 (2013).
76. M. Becker, E. Hobeika, H. Jumaa, M. Reth, P. C. Maity, CXCR4 signaling and function require the expression of the IgD-class B cell antigen receptor. *Proc. Natl. Acad. Sci. U.S.A.* **114**, 5231–5236 (2017).
77. J. Z. Chen, A. M. Stall, L. A. Herzenberg, Differences in glycoprotein complexes associated with IgM and IgD on normal murine B cells potentially enable transduction of different signals. *EMBO J.* **9**, 2117–2124 (1990).
78. G. J. Silverman, C. S. Goodyear, A model B cell superantigen and the immunobiology of B lymphocytes. *Clin. Immunol.* **102**, 117–134 (2002).
79. G. J. Silverman, C. S. Goodyear, Confounding B cell defences: Lessons from a staphylococcal superantigen. *Nat. Rev. Immunol.* **6**, 465–475 (2006).
80. J. Jellusova, A. V. Miletic, M. H. Cato, W.-W. Lin, Y. Hu, G. A. Bishop, M. J. Shlomchik, R. C. Rickert, Context-specific BAFF-R signaling by the NF- κ B and PI3K pathways. *Cell Rep.* **5**, 1022–1035 (2013).
81. E. Hobeika, E. Levit-Zerdoun, V. Anastasopoulou, R. Pohlmeier, S. Altmeier, A. Alsadeq, M.-W. Dobenecker, R. Pelanda, M. Reth, CD19 and BAFF-R can signal to promote B cell survival in the absence of Syk. *EMBO J.* **34**, 925–939 (2015).

82. X. Chen-Deutsch, A. Kutner, J. S. Harrison, G. P. Studzinski, The pan-caspase inhibitor Q-VD-OPH has anti-leukemia effects and can interact with vitamin D analogs to increase HPK1 signaling in AML cells. *Leuk. Res.* **36**, 884–888 (2012).
83. T. G. Nguyen, J. M. Morris, Signals from activation of B-cell receptor with anti-IgD can override the stimulatory effects of excess BAFF on mature B cells in vivo. *Immunol. Lett.* **161**, 157–164 (2014).
84. S. E. Spurgeon, G. Coffey, L. B. Fletcher, R. Burke, J. W. Tyner, B. J. Druker, A. Betz, F. DeGuzman, Y. Pak, D. Baker, A. Pandey, S. J. Hollenbach, U. Sinha, M. M. Loriaux, The selective syk inhibitor P505-15 (PRT062607) inhibits B cell signaling and function in vitro and in vivo and augments the activity of fludarabine in chronic lymphocytic leukemia. *J. Pharmacol. Exp. Ther.* **344**, 378–387 (2013).
85. P. M. Higgins, Splenomegaly in acute infections due to group A streptococci and viruses. *Epidemiol. Infect.* **109**, 199–209 (1992).
86. P. A. Nikitin, A. M. Price, K. McFadden, C. M. Yan, M. A. Luftig, Mitogen-induced B cell proliferation activates Chk2-dependent G1/S cell cycle arrest. *PLOS ONE* **9**, e87299 (2014).
87. J. S. Turner, M. Marthi, Z. L. Benet, I. Grigorova, Transiently antigen-primed B cells return to naive-like state in absence of T cell help. *Nat. Commun.* **8**, 15072 (2017).
88. B. Nardelli, O. Belvedere, V. Roschke, P. A. Moore, H. S. Olsen, T. S. Migone, S. Sosnovtseva, J. A. Carrell, P. Feng, J. G. Giri, D. M. Hilbert, Synthesis and release of B-lymphocyte stimulator from myeloid cells. *Blood* **97**, 198–204 (2001).
89. C. Chaparro, J. Maurer, C. Gutierrez, M. Krajden, C. Chan, T. Winton, S. Keshavjee, M. Scavuzzo, E. Tullis, M. Hutcheon, S. Kesten, Infection with *Burkholderia cepacia* in cystic fibrosis: Outcome following lung transplantation. *Am. J. Respir. Crit. Care Med.* **163**, 43–48 (2001).
90. H. Funken, K.-M. Bartels, S. Wilhelm, M. Brocker, M. Bott, M. Bains, R. E. W. Hancock, F. Rosenau, K.-E. Jaeger, Specific association of lectin LecB with the surface of *Pseudomonas aeruginosa*: Role of outer membrane protein OprF. *PLOS ONE* **7**, e46857 (2012).
91. P. L. Wagner, J. Livny, M. N. Neely, D. W. K. Acheson, D. I. Friedman, M. K. Waldor, Bacteriophage control of Shiga toxin 1 production and release by *Escherichia coli*. *Mol. Microbiol.* **44**, 957–970 (2002).
92. P. L. Wagner, D. W. K. Acheson, M. K. Waldor, Human neutrophils and their products induce Shiga toxin production by enterohemorrhagic *Escherichia coli*. *Infect. Immun.* **69**, 1934–1937 (2001).
93. S. B. Calderwood, J. J. Mekalanos, *Genetic Analysis of Regulation and Structure of Shiga-Like Toxin I in Escherichia coli* (Springer US, 1990), pp. 57–64.
94. C. Chemani, A. Imbert, S. de Bentzmann, M. Pierre, M. Wimmerová, B. P. Guery, K. Faure, Role of LecA and LecB lectins in *Pseudomonas aeruginosa*-induced lung injury and effect of carbohydrate ligands. *Infect. Immun.* **77**, 2065–2075 (2009).
95. K. Nothelfer, P. J. Sansonetti, A. Phalipon, Pathogen manipulation of B cells: The best defence is a good offence. *Nat. Rev. Microbiol.* **13**, 173–184 (2015).
96. K. Nothelfer, E. T. Arena, L. Pinaud, M. Neunlist, B. Mozeleski, I. Belotserkovsky, C. Parsot, P. Dinadayala, A. Burger-Kentscher, R. Raqib, P. J. Sansonetti, A. Phalipon, B lymphocytes undergo TLR2-dependent apoptosis upon *Shigella* infection. *J. Exp. Med.* **211**, 1215–1229 (2014).
97. K. Chen, W. Xu, M. Wilson, B. He, N. W. Miller, E. Bengtén, E.-S. Edholm, P. A. Santini, P. Rath, A. Chiu, M. Cattalini, J. Litzman, J. B. Bussel, B. Huang, A. Meini, K. Riesbeck, C. Cunningham-Rundles, A. Plebani, A. Cerutti, Immunoglobulin D enhances immune surveillance by activating antimicrobial, proinflammatory and B cell-stimulating programs in basophils. *Nat. Immunol.* **10**, 889–898 (2009).
98. A. J. Ulloa-Morales, C. S. Goodyear, G. J. Silverman, Essential domain-dependent roles within soluble IgG for in vivo superantigen properties of staphylococcal protein A: Resolving the B cell superantigen paradox. *Front. Immunol.* **9**, 2011 (2018).
99. W. Li, Q. Liu, Y. Pang, J. Jin, H. Wang, H. Cao, Z. Li, X. Wang, B. Ma, Y. Chi, R. Wang, A. Kondo, J. Gu, N. Taniguchi, Core fucosylation of μ heavy chains regulates assembly and intracellular signaling of precursor B cell receptors. *J. Biol. Chem.* **287**, 2500–2508 (2012).
100. A. L. Morrow, J. Meinen-Derr, P. Huang, K. R. Schibler, T. Cahill, M. Keddache, S. G. Kallapur, D. S. Newburg, M. Tabangin, B. B. Warner, X. Jiang, Fucosyltransferase 2 non-secretor and low secretor status predicts severe outcomes in premature infants. *J. Pediatr.* **158**, 745–751 (2011).
101. B. Carlsson, E. Kindberg, J. Buesa, G. E. Rydell, M. F. Lidón, R. Montava, R. A. Mallouh, A. Grahn, J. Rodriguez-Diaz, J. Bellido, A. Arnedo, G. Larson, L. Svensson, The G428A nonsense mutation in FUT2 provides strong but not absolute protection against symptomatic GII.4 norovirus infection. *PLOS ONE* **4**, e5593 (2009).
102. M. Fumagalli, R. Cagliani, U. Pozzoli, S. Riva, G. P. Comi, G. Menozzi, N. Bresolin, M. Sironi, Widespread balancing selection and pathogen-driven selection at blood group antigen genes. *Genome Res.* **19**, 199–212 (2008).
103. R. F. Villar, J. Patel, G. C. Weaver, M. Kanekiyo, A. K. Wheatley, H. M. Yassine, C. E. Costello, K. B. Chandler, P. M. McTamney, G. J. Nabel, A. B. McDermott, J. R. Mascola, S. A. Carr, D. Lingwood, Reconstituted B cell receptor signaling reveals carbohydrate-dependent mode of activation. *Sci. Rep.* 36298 (2016).
104. R. Pelanda, U. Braun, E. Hobeika, M. C. Nussenzweig, M. Reth, B cell progenitors are arrested in maturation but have intact VDJ recombination in the absence of Ig- α and Ig- β . *J. Immunol.* **169**, 865–872 (2002).
105. L. Nitschke, M. H. Kosco, G. Köhler, M. C. Lamers, Immunoglobulin D-deficient mice can mount normal immune responses to thymus-independent and -dependent antigens. *Proc. Natl. Acad. Sci. U.S.A.* **90**, 1887–1891 (1993).
106. M. Inaoki, S. Sato, B. C. Weintraub, C. C. Goodnow, T. F. Tedder, CD19-regulated signaling thresholds control peripheral tolerance and autoantibody production in B lymphocytes. *J. Exp. Med.* **186**, 1923–1931 (1997).
107. M. L. Hibbs, D. M. Tarlinton, J. Armes, D. Grail, G. Hodgson, R. Maglitto, S. A. Stacker, A. R. Dunn, Multiple defects in the immune system of *Lyn*-deficient mice, culminating in autoimmune disease. *Cell* **83**, 301–311 (1995).
108. S. Sugawara, M. Hosono, Y. Ogawa, M. Takayanagi, K. Nitta, Catfish egg lectin causes rapid activation of multidrug resistance 1 P-glycoprotein as a lipid translocase. *Biol. Pharm. Bull.* **28**, 434–441 (2005).

Acknowledgments: We thank M. Becker, P. C. Maity, and the whole Reth group for material and technical support, particularly for access to the animal facility of the Max Planck Institute of Immunology and Epigenetics (Freiburg, Germany). Furthermore, we appreciate support by the BIOS ToolBox, especially P. Salavei, in the purification of the bacterial lectins. We thank M. Schmidt-Supprian (Technical University Munich, Germany) for the CD19-CreER^{T2} mice. We further thank P. Hennecke for helpful discussions during thesis committee meetings, as well as S. Samodelov and R. Pohlmeier for proofreading the manuscript. We would like to acknowledge A. Nyström of the Bruckner-Tudermann group and C. Molnár from the Erlacher group at the University Medical Centre Freiburg for their supply of the murine WT spleens for some in vitro experiments. We also thank J. M. Weiss from the Erlacher group for help with the myeloid gating strategy and G. Andrieux for the evaluation of the statistical analysis. **Funding:** The research group of W.R. was supported by the Excellence Initiative of the German Research Foundation (EXC 294 and GSC-4); the Ministry of Science, Research and the Arts Baden-Württemberg (Az: 33-7532.20); and a starting grant from the European Research Council (Programme “Ideas”, ERC-2011-StG 282105-lec&lip2invade). A.I. and W.R. acknowledge support by the French National Research Agency (ANR) and the German Federal Ministry of Education and Research (BMBF) in the framework of the EU ERASynBio project SynGlycTis (ANR-14-SYNB-0002-02; BMBF 031A464). A.I. was further supported by Glyco@Alps (ANR-15-IDEX-02). This study was partially funded by the German Research Agency (DFG) through the grants RTG 2202 and RO 4341/2-1. I.W. and R.U. were funded by the Excellence Initiative of the DFG (GSC-4, Spemann Graduate School), and E.H. was partly supported by the SFB 1074 P N09. E.L.-Z. and M.F. were funded by the International Max Planck Research School for Molecular and Cellular Biology, Max Planck Institute of Immunobiology and Epigenetics (Freiburg, Germany). R.T. was supported by the State Ministry of Sciences, Research and the Arts Baden-Württemberg (Az: 7533-30-10/25/36). **Author contributions:** I.W., W.R., and E.H. designed this interdisciplinary study. The experiments were conducted by I.W., M.F., and J.J. under the supervision of E.H., E.L.-Z., and W.R. Furthermore, P.M., S.V., and A.L. provided preliminary data. A.I., R.T., and J.C. established the lectin experiments and supervised the respective data analysis. R.U. helped perform Ca²⁺ experiments and establish the methodology. I.W. and E.L.-Z. performed data analysis and wrote the manuscript. M.R., H.E., and H.J. provided experimental advice and B cell-specific expertise. W.R. and E.H. supervised the study. **Competing interests:** The authors declare that they have no competing interests. **Data and materials availability:** All data needed to evaluate the conclusions in the paper are present in the paper or the Supplementary Materials.

Submitted 18 August 2017
Resubmitted 11 September 2017
Accepted 15 February 2019
Published 5 March 2019
10.1126/scisignal.aao7194

Citation: I. Wilhelm, E. Levit-Zerdoun, J. Jakob, S. Villringer, M. Frensch, R. Übelhart, A. Landi, P. Müller, A. Imbert, R. Thuenauer, J. Claudinon, H. Jumaa, M. Reth, H. Eibel, E. Hobeika, W. Römer, Carbohydrate-dependent B cell activation by fucose-binding bacterial lectins. *Sci. Signal.* **12**, eaao7194 (2019).

Carbohydrate-dependent B cell activation by fucose-binding bacterial lectins

Isabel Wilhelm, Ella Levit-Zerdoun, Johanna Jakob, Sarah Villringer, Marco Frensch, Rudolf Übelhart, Alessia Landi, Peter Müller, Anne Imberty, Roland Thuenauer, Julie Claudinon, Hassan Jumaa, Michael Reth, Hermann Eibel, Elias Hobeika and Winfried Römer

Sci. Signal. **12** (571), eaao7194.
DOI: 10.1126/scisignal.aao7194

Sweet B cell activation

Burkholderia and *Pseudomonas* species of proteobacteria secrete carbohydrate-binding proteins called lectins, enabling adhesion to host tissues and biofilm formation, which can lead to opportunistic infections. Wilhelm *et al.* showed that fucose-binding bacterial lectins bound to and activated mouse B cells *in vitro*, which depended on the B cell antigen receptor and the kinase Syk. *In vivo*, these lectins induced a strong but transient immune response characterized by increased numbers of B cells and myeloid cells in the spleen. These data suggest that bacterial lectins may act like antigens to hijack B cell signaling and stimulate rapid immune responses, which may pose a risk to immunocompromised individuals.

ARTICLE TOOLS

<http://stke.sciencemag.org/content/12/571/eaao7194>

SUPPLEMENTARY MATERIALS

<http://stke.sciencemag.org/content/suppl/2019/03/01/12.571.eaao7194.DC1>

RELATED CONTENT

<http://stke.sciencemag.org/content/sigtrans/12/564/eaar5514.full>
<http://stke.sciencemag.org/content/sigtrans/10/508/eaan3676.full>
<http://stke.sciencemag.org/content/sigtrans/11/532/eaal1506.full>
<http://science.sciencemag.org/content/sci/361/6400/402.full>
<http://science.sciencemag.org/content/sci/363/6427/649.full>

REFERENCES

This article cites 105 articles, 38 of which you can access for free
<http://stke.sciencemag.org/content/12/571/eaao7194#BIBL>

PERMISSIONS

<http://www.sciencemag.org/help/reprints-and-permissions>

Use of this article is subject to the [Terms of Service](#)